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JCS25 U.S. PTO

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INTELLECTUAL PROPERTY LAW
(PATENT, BIOTECHNOLOGY, COMPUTER,
TRADEMARK & TRADE SECRET LAW)

JCS25 U.S. PTO
09/320156
05/26/99

May 26, 1999

Docket No.: D5425CIP2

The Assistant Commissioner of Patents
BOX PATENT APPLICATION
Washington, DC 20231

Dear Sir:

Transmitted herewith for filing is the continuation-in-part patent application which claims benefit of priority of non-provisional application 08/404,499, filed March 17, 1995, which is a continuation in part of US Serial No. 08/300,082, filed September 2, 1994, now abandoned, which is a continuation of US Serial No. 08/164,638, filed 12/09/93, now abandoned, which is a continuation of US Serial No. 07/867,728, filed 04/10/92, now abandoned.

Name of: **Rosenblum, Michael G. and Shawver, Laura K.**
For: ***Immunotoxins Directed Against *cerbB-2* (HER-2/neu)
Related Surface Antigens***

CLAIMS AS FILED

Fee for:	Small entity	Amount
Basic fee	\$ 3 8 0	\$ 3 8 0
Each independent claim in excess of 3 (0)		
Each claim excess of 20 (1) \$ 9		\$ 9
Multiple dependent claim		

TOTAL FILING FEE \$ 3 8 9

____ Please charge my Deposit Account No. _____ in the total
amount of the filing fee and the assignment recordation fee if any.

X The Commissioner is hereby authorized to charge any additional fees which may be required, or credit any overpayment to Deposit Account No. 07-1185.

X Any additional fees under 37 CFR 1.16.

X Any application processing fees under 37 CFR 1.17.

X Small Entity Statement

A small entity statement is enclosed and its benefit under 37 CFR 1.28(a) is hereby claimed.

X Relate Back--35 U.S.C. 120

This continuation-in-part application claims benefit of priority of continuation in part application 08/404,499, filed March 17, 1995, which is a continuation in part of US Serial No. 08/300,082, filed September 2, 1994, now abandoned, which is a continuation of US Serial No. 08/164,638, filed 12/09/93, now abandoned, which is a continuation of US Serial No. 07/867,728, filed 04/10/92, now abandoned.

Assignment

The application is assigned by the inventors to the_____.

X Sequence Listing

The sequence listing is enclosed, including a paper copy, a copy on disk for the computer readable form and a compliance letter indicating that the sequence listing on the paper copy and the disk are one and the same.

X Power of Attorney

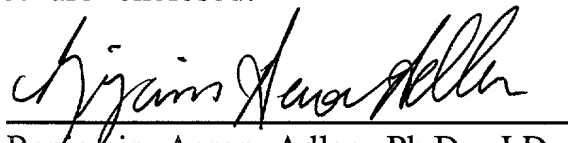
X is attached.

X Address all future communications to:

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X Two photocopies of this sheet are enclosed.

Date: May 26, 1999


Benjamin Aaron Adler, Ph.D., J.D.
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IN THE UNITED STATES PATENT AND TRADEMARK OFFICE

APPLICANT: Rosenblum, *et al.* § ART UNIT:
§
FILED: May 26, 1999 §
§ EXAMINER:
SERIAL NO.: §
§
FOR: Immunotoxins Directed Against § DOCKET:
c-erbB-2 (HER-2/Neu) Related § D5425CIP2
Surface Antigens §

The Assistant Commissioner of Patents and Trademarks
BOX PATENT APPLICATION
Washington, D.C. 20231

COMPLIANCE OF REQUIREMENTS FOR PATENT APPLICATIONS
CONTAINING NUCLEOTIDE AND/OR AMINO ACID SEQUENCE
DISCLOSURES

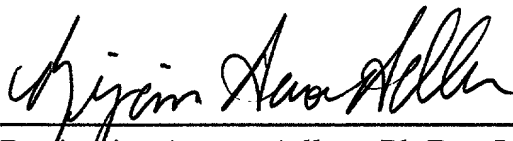
Dear Sir:

Applicant provides a computer readable form of the Sequence Listing on the enclosed 3.5 inch disk and a paper copy thereof for the above-referenced application. The disk is a 1.44 mb Apple Macintosh-formatted disk. The file is stored as D5425CIP2/SEQ in text format. I hereby state that the content of the paper copy of the Sequence Listing filed as part of the above-captioned application and the enclosed computer readable copy of the Sequence Listing are the same.

Respectfully submitted,

Date:

May 26, 1999



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Applicant or Patentee: Rosenblum et al Attorney's
Serial or Patent No.: _____ Docket No.: D5425C1P4

Filed or Issued:

For: Immunotoxins Directed Against c-erbB-2 (HER-2/Neu) Related Surface Antigens

**VERIFIED STATEMENT (DECLARATION) CLAIMING SMALL ENTITY
STATUS (37 CFR 1.9(f) and 1.27(c)) - NONPROFIT ORGANIZATION**

I hereby declare that I am an official of the nonprofit organization empowered to act on behalf of the concern identified below:

NAME OF ORGANIZATION: Research Development Foundation

ADDRESS OF CONCERN: 402 North Division Street
Carson City, Nevada 89703

TYPE OF ORGANIZATION:

☒ TAX EXEMPT UNDER INTERNAL REVENUE SERVICE CODE (26 USC 501(a) AND 501(c)(3))

I hereby declare that the above identified nonprofit organization qualifies as a nonprofit organization as defined in 37 CFR 1.9(e), for purposes of paying reduced fees under section 41(a) and (b) of Title 35, United States Code.

I hereby declare that rights under contract or law have been conveyed to and remain with the nonprofit organization identified above with regard to the invention, entitled as above by inventor(s) as above described in

- ☒ the specification filed herewith
☐ application serial no. _____, filed _____
☐ patent no. _____, issued _____

If the rights held by the above identified nonprofit organization are not exclusive, each individual, concern or organization having rights to the invention is listed below* and no rights to the invention are held by any person, other than the inventor, who could not qualify as a small business concern under 37 CFR 1.9(d) or by any concern which would not qualify as a small business concern under 37 CFR 1.9(d) or a nonprofit organization under 37 CFR 1.9(e).

*NOTE: Separate verified statements are required from each named person, concern or organization having rights to the invention averring to their status as small entities. (37 CFR 1.27)

NAME _____
ADDRESS _____

☐ INDIVIDUAL ☐ SMALL BUSINESS CONCERN ☐ NONPROFIT ORGANIZATION

NAME _____
ADDRESS _____

☐ INDIVIDUAL ☐ SMALL BUSINESS CONCERN ☐ NONPROFIT ORGANIZATION

I acknowledge the duty to file, in this application or patent, notification of any change in status resulting in loss of entitlement to small entity status prior to paying, or at the time of paying, the earliest of the issue fee or any maintenance fee due after the date on which status as a small entity is no longer appropriate. (37 CFR 1.28(b))

I hereby declare that all statements made herein of my own knowledge are true and that all statements made on information and belief are believed to be true; and further that these statements were made with knowledge that willful false statements and the like so made are punishable by fine or imprisonment, or both, under section 1001 of Title 18 of the United States Code, and that such willful false statements may jeopardize the validity of the application, any patent issuing thereon, or any patent to which this verified statement is directed.

NAME OF PERSON SIGNING: JAMES F. WEILER

TITLE OF PERSON OTHER THAN OWNER: VICE PRESIDENT

ADDRESS OF PERSON SIGNING ONE RIVERWAY, SUITE 1560, HOUSTON, TEXAS 77056

SIGNATURE: JAMES F. WEILER

DATE: 5.19.99

**IMMUNOTOXINS DIRECTED AGAINST
c-erbB-2(HER-2/neu) RELATED SURFACE ANTIGENS**

5

BACKGROUND OF THE INVENTION

10 **Cross Reference to Related Application**

 This application is a continuation in part of US Serial No.
08/404,499, filed March 17, 1995, which is a continuation in part of
US Serial No. 08/300,082, filed September 2, 1994, now abandoned,
which is a continuation of US Serial No. 08/164,638, filed
12/09/93, now abandoned, which is a continuation of US Serial No.
07/867,728, filed 04/10/92, now abandoned.

Field of the Invention

 The present invention relates generally to the field of
treatment of neoplastic disease. More specifically, the present
invention relates to novel immunoconjugates and their use in the
treatment of neoplastic disease.

Description of the Related Art

Neoplastic disease is one of the leading causes of mortality and morbidity in the Western World. Neoplastic conditions, *e.g.*, diseases or "cancers", share at least one characteristic, i.e., the involvement of defects in the cellular growth regulatory process. The process by which normal cells are transformed into malignant cells has been a subject of intense study for decades. More recently, study has focused on the role of oncogenes in the cancer process. Oncogenes are genes that have the ability to transform eukaryotic cells so that they grow in a manner analogous to tumor cells.

An oncogene is created when a normal gene or proto-oncogene is mutated, rearranged, or amplified. One such oncogene is the c-erbB-2(HER-2/*neu*) proto-oncogene. Hereinafter, this oncogene will be referred to as c-erbB-2. This gene encodes a protein similar to epidermal growth factor receptor. Amplification of this proto-oncogene can result in a cascade of cellular events leading to unregulated cell growth.

Antibodies are proteins produced by the immune system of an animal, normally in response to foreign antigens or antigenic determinants. Antibodies bind to the specific antigen to which they

are directed. The development of specific monoclonal antibodies has provided investigators with a possible means of selectively targeting therapeutic agents to cells which overexpress defined antigens.

Overexpression of the c-erbB-2 proto-oncogene in
5 neoplastic transformation has been postulated. Several types of human cancers including some mammary carcinomas and some ovarian carcinomas have an amplified c-erbB-2 gene. Moreover, amplification and subsequent overexpression of the c-erbB-2 gene has been correlated with poor disease prognosis. Thus, there exists a
10 great need and desire in this art for a method of selectively targeting a chemotherapeutic agent to a cell which exhibits overexpression of the c-erbB-2 oncogene to modulate growth of cells which overexpress the protein. The present invention provides means for accomplishing
15 this.

SUMMARY OF THE INVENTION

The present invention provides a composition comprising a conjugate of a cellular targeting moiety, *e.g.*, an antigen binding
20 region, exhibiting binding specificity for the c-erbB-2 protein and a cell growth modulator, *e.g.*, a toxin or growth suppressing reagent.

This composition acts as an immunotoxin to specifically target a cell growth modulator to tumor cells overexpressing the c-erbB-2 protein.

Thus, in one embodiment of the present invention, there is provided a new composition of matter comprising a conjugate of a
5 targeting moiety with binding specificity for the c-erbB-2 protein, *e.g.*, TAb 250 monoclonal antibody, and a cytotoxic moiety. The cytotoxic moiety may be a toxin, a cytocidal drug, a cytostatic drug, or a biological response modifier. In one particular embodiment, the cytotoxic moiety is gelonin.

Another embodiment of the present invention provides a
10 method of treating a neoplastic condition, *e.g.*, disease, which is characterized by amplification or overexpression of the c-erbB-2 oncogene, comprising administering a cytocidally effective dose of an immunotoxin of the present invention to an individual in need of
15 such treatment.

In still another embodiment of the present invention, there is provided compositions of matter comprising fusion constructs of targeting moieties with binding-affinity for c-erbB-2 protein and a cytotoxic moiety. Preferably, the targeting moiety is
20 an antibody which recognizes an extracellular epitope of c-erbB-2, *e.g.*, TAb 250, and the cytotoxic moiety is relatively inert when

applied separately from the targeting moiety, *e.g.*, gelonin. In other embodiments of the present invention there are provided methods of extending the survival time of a tumor bearing mammal by administration of targeted toxins of the present invention to this mammal and also a method of retarding the rate of growth of tumors by administering targeted toxins of the present invention. Typically, the targeted toxins will be targeted by an immunological binding region, *e.g.*, an antibody binding segment. Additionally provided is a pharmaceutical composition comprising an immunotoxin consisting essentially of a cytotoxic moiety conjugated to a monoclonal antibody. Most preferably, the antibody is TAb 250 and the cytotoxic moiety is gelonin.

In another embodiment of the instant invention, there is provided a conjugate of tumor necrosis factor to an antibody exhibiting binding specificity for an extracellular epitope of c-erbB-2 protein. The antibody may be an intact full length antibody with either the heavy chain or light chain peptide conjugated to tumor necrosis factor. Alternatively, the antibody may be or a Fv fragment with the toxin linked to either the V_L or V_H peptide. In the preferred embodiment, conjugate is a fusion protein between a single chain antibody and tumor necrosis factor which is preferably produced by

recombinantly fusing a gene encoding a single chain antibody to a gene encoding tumor necrosis factor. One possible sFv is scFv-23.

Finally, another embodiment of the instant invention comprises a method of treating neoplastic cells with an antibody-TNF conjugate. Possible target cells include mammary carcinoma cells, ovarian carcinoma cells, lung carcinoma cells, salivary gland carcinoma cells, gastric tumor cells, colon adenocarcinoma cells, and bone marrow leukemia cells. A specific example entailing breast carcinoma cells is provided.

BRIEF DESCRIPTION OF THE DRAWINGS

Figure 1 demonstrates the effects of ZME antibody, TAb 250 antibody or immunonconjugates of TAb 250 and gelonin on SKOV-3 cells as measured by ELISA.

Figure 2 demonstrates the cytotoxicity of the TAb 250 gelonin construct on SKOV-3 cells.

Figure 3 demonstrates the competition of relevant versus irrelevant antibody with the conjugate on SKOV-3 cells.

Figure 4 demonstrates the dose response relationship and the effects of the TAb 250-gelonin conjugate on SKOV-3 cells.

Figure 5 demonstrates the cytotoxicity of TAb 250 and the TAb 250–gelonin conjugate on SKOV–3 cells.

Figure 6 demonstrates the ability of the TAb 250 antibody to internalize in various cell lines.

5 **Figure 7** demonstrates the cytotoxicity of the TAb 250–gelonin immunoconjugate in the MTT assay.

Figure 8 shows a schematic of the construction of the scFv23-gelonin immunotoxin gene. The first PCR syntheses were performed in separate tubes and one tenth of each of the reaction products were subsequently combined and used as templates for the second PCR (Davis et al., BioTechnology, 9:165-9 (1991). Nco I and Hind III restriction sites were included into the sequences of the "outer" flanking primers and 1 and 4 for future cloning purposes. Homology between bases in the PCR primers 2 and 3 is indicated by the filled boxes at the 5' ends of each primer and represented DNA sequences encoding the Gly-Gly-Gly-Gly-Ser (G4S) (Seq ID No: 1) linker between the antibody and gelonin fragments of the immunotoxin.

10 **Figure 9** shows the configuration and sequence of the scFv-gelonin immunotoxin. The single chain antibody fragment was cloned in the V_L-212 linker-VH orientation and was fused to the toxin

gelonin by a short, nonstructured five amino acid peptide (G4S). Figure 9B shows the DNA sequence encoding the immunotoxin is shown with the corresponding protein sequence. The six antibody complementarity determining (CDRs) and the 212 linker sequence between the VL and VH domains are highlighted: the G4S sequence linking the antibody and toxin and fragments is italicized. DNA encoding the synthetic gene for gelonin was used as a template for the gene fusion and in addition, encoded the targeting signal KDEL incorporated into the 3' end.

Figure 10 shows a binding analysis of sixteen individual scFv-gelonin immunotoxin clones. Bacterial culture supernatants and periplasmic extracts of individually expressed immunotoxins were added to wells of a 96 well ELISA plate coated with either (A) mouse anti-gelonin antibody 13A3, or (B) 5×10^4 antigen positive SK-OV-3 cells. Bound immunotoxin was detected with a rabbit anti-gelonin polyclonal antibody followed by addition of a horseradish peroxidase (HRPO) conjugate of a goat anti-rabbit IgG secondary antibody. Signals were developed with the horse radish peroxidase substrate ABTS and measured at 405 nm. The negative and positive controls were free gelonin (at 50 micrograms/ml, 10 micrograms/ml and 1 microgram/ml) and a gelonin conjugate of TAb 250, a murine

anti-HER-2/neu antibody (tested at 50 ng/ml, 10 ng/ml, and 1 ng/ml), respectively.

Figure 11 shows a western blot analysis of the scFvZME-gelonin and scFv-23-gelonin proteins refolded and purified from insoluble inclusion bodies. Refolded GST fusions of the two immunotoxins were concentrated, bound to glutathione-agarose (GSH-ag) and digested with thrombin protease. The supernatant containing cleaved protein was retreated with GSH-ag rebind any GST and immediately bound to Blue Sepharose (B.S.) resin. Immunotoxins were subsequently eluted with 2 M NaCl and dialyzed into TBS. Nitrocellulose filters used in the western analysis were developed using the Amersham ECL system with an exposure time of 30 seconds. Molecular weight are indicated on the left. Lane 1: Refolded GST-fusion of scFvZME-Gel; Lane 2: Flowthrough after binding of scFvZME-Gel to BLUE SEPHAROSE; Lane 3: scFvAME-gelonin eluate from BLUE SEPHAROSE; Lane 4: GST-scFvAME-gelonin molecular weight marker; Lane 5: refolded GST-fusion of scFv-gelonin; Lane 6: flowthrough after binding scFv-23-gelonin to BLUE SEPHAROSE; Lane 7: scFv-23-gelonin eluate from BLUE SEPHAROSE; Lane 8: gelonin standard (40 ng).

Figure 12 demonstrates binding of TAb-250/recombinant gelonin, TAb-250, or an irrelevant murine antibody (ZME) to adherent SKOV-3 cells. Binding was measured in an ELISA, in which various concentrations of each reagent were added to each well and the plates were assayed for the presence of murine antibody. The values shown are the means (\pm SEM) of quadruplicate determinations.

Figure 13 shows displacement of either BACH-250 or BACH-250/recombinant gelonin on SKOV-3 cells. Both BACH-250/recombinant gelonin and unconjugated BACH-250 showed similar binding curves, with affinity constants of 6.12×10^{-10} and 8.42×10^{-10} respectively, indicating that the conjugation of recombinant gelonin to BACH-250 did not significantly interfere with the ability of the conjugate to bind to the c-erbB-2 protein on the cell surface.

Figure 14 shows dose-response curves for BACH-250/recombinant gelonin on SKOV-3 cells. Various concentrations of BACH-250/recombinant gelonin were added to log-phase SKOV-3 cells. The immunotoxin was also admixed with fixed concentrations of free BACH-250 and then added to cells. Cells were incubated for

72 hours and cell concentrations were assessed. The values shown are the means (\pm SEM) for octuplicate determinations.

Figure 15 demonstrates the effect of BACH-250 on SKOV-3 colony formation. SKOV-3 cells were treated with 1.0, 0.5, 0.1, 0.05, and 0.01 ug/ml of BACH-250/rGel and plated in soft agar. Colonies were counted for up to 10 days after plating. The values shown are the means (\pm SEM) of quadruplicate determinations.

Figure 16 demonstrates the effect of HER-2 expression level on the cytotoxicity of TAb-250/rGel. Various human cell lines in log-phase culture were treated with concentrations of the immunotoxin for 72 hours. The cell number was assessed and expressed as the percentage of untreated control wells. The values shown are the means (\pm SEM) of quadruplicate determinations.

Figure 17 demonstrates plasma clearance of BACH-250 and BACH-250/rGel. Both BACH-250 and BACH-250/rGel were radiolabeled and administered separately to groups of balb/c mice by an intravenous bolus (tail vein). Six mice per group were sacrificed at various times after administration and blood was collected from the chest cavity and weighed and the radioactivity counted. The values shown are the means (\pm SEM) of six mice. The

curves represent the least-squares, best-fit lines through the data points.

Figure 18 demonstrates the tissue disposition of BACH-250 and BACH-250/rGel 24 hours after administration. Radiolabeled antibody and immunotoxin were administered intravenously into the tail vein in nude mice bearing well-developed SKOV-3 subcutaneous tumors. Twenty-four hours after administration, 6 mice in each group were sacrificed, and various organs were excised, and weighed and the radioactivity counted. The values shown are the means (\pm SEM) of six mice.

Figure 19 demonstrates the tissue disposition of BACH-250 and BACH-250/rGel 48 hours after administration. Radiolabeled antibody and immunotoxin were administered intravenously into the tail vein in nude mice bearing well-developed SKOV-3 subcutaneous tumors. Forty-eight hours after administration, 6 mice in each group were sacrificed and various organs were excised, weighed and the radioactivity counted. The values shown are the means (\pm SEM) of 6 mice.

Figure 20 demonstrates the tissue disposition of BACH-250 and BACH-250/rGel 72 hours after administration. Radiolabeled antibody and immunotoxin were administered intravenously into the

tail vein in nude mice bearing well-developed SKOV-3 subcutaneous tumors. Seventy-two hours after administration, 6 mice/group were sacrificed and various organs were excised, weighed and counted. The values shown are the means (\pm SEM) of six mice.

Figure 21 demonstrates the tissue disposition of BACH-250 and BACH-250/rGel 96 hours after administration. Radiolabeled antibody and immunotoxin were administered intravenously into the tail vein in nude mice bearing well-developed SKOV-3 subcutaneous tumors. Ninety-six hours after administration, 6 mice in each group were sacrificed and various organs were excised, and weighed and the radioactivity counted. The values shown are the means (\pm SEM) of six mice.

Figure 22 demonstrates tumor:blood ratios of BACH-250 and BACH-250/rGel at various times after administration. Analysis of the radioactivity of blood and tumor after administration of ^{125}I BACH-250 or BACH-250/Gel conjugate showed that the maximal tumor concentration of either antibody or immunotoxin occurs approximately 48 hours after administration and remains fairly constant up to 48 hours thereafter.

Figure 23 demonstrates the treatment of subcutaneous tumors in balb/c mice with the immunotoxin conjugates. Balb/c

athymic mice received subcutaneous implants of the ovarian tumor cell line SKOV-3 in the right hind flank. One week after implantation, the mice were randomly divided into groups of 8 and treated with PBS, 42 μ g TAb-250 plus 8 μ g gelonin, or 50 μ g TAb-250/rGel via intraperitoneal injection. Treatments were administered 2 times per week for three weeks. Tumor volume (L x W x H) was determined by caliper measurements taken twice per week and averaged for each treatment group. This figure shows that tumor growth was significantly more inhibited than in the PBS control mice and also in animals receiving the TAb-250/rGel conjugate (99% at day 35 after implantation, 94% at day 49) and also more than it was in animals given equimolar amounts of unconjugated TAb-250 plus free rGel (4% at day 35, 56% at day 49).

Figure 24 demonstrate the results of immunotoxin treatment of intraperitoneally implanted tumors in balb/c athymic mice. Mice (7-8 mice/group) were implanted (day 0) subcutaneously with SKOV-3 tumor cells. Treatments were administered intraperitoneally 3 times per week for 3 weeks starting on day 5 after implantation (tumor volumes were 0-30 mm³). Mice received either PBS (control), TAb-250 (42 ug/dose) plus gelonin (42 ug/dose), or the TAb-250/rGel conjugate (50 ug/dose). Tumors were

measured twice per week with vernier calipers and volumes (mm^3) were calculated as a product of length x width x height. Tumor volumes were plotted as a function of day of growth. As shown in Figure 29, TAb-250/rGel treatment enhanced the survival of mice with intraperitoneal SKOV-3 tumors by an average of 12.4 days as compared with survival in mice treated with unconjugated TAb-250 or TAb-250 plus gelonin.

Figure 25 shows a schematic representing the design of the sfv23/TNF fusion construct and the assembly of the expression vector.

Figure 26 demonstrates the expression and purification of both the sfv23 antibody as well as the sfv23/TNF fusion construct from *E. coli* utilizing an immobilized metal affinity column(IMAC).

Figure 27 demonstrates the SDS-PAGE and Coomassie staining profile of bacteria prior to induction (left lane) and following induction (right lane) by addition of IPTG. The position of the scFv23/TNF fusion construct is indicated by the arrow. Western analysis using anti-sfv23 antibody demonstrates reactivity against purified scFv23 antibody (left lane) and the scFv23/TNF fusion construct (right lane) while probing the same Western blot with an

anti-TNF antibody demonstrates reactivity only with the scFv23/TNF construct.

Figure 28 shows ELISA binding of either sfv23 or sfv23/TNF to SKBR3 immobilized cells.

5 **Figure 29** shows ELISA binding of scFv23/TNF construct to either antigen-positive SKBR3 or antigen-negative A-373 cells.

Figure 30 demonstrates western analysis of two variants of SKBR3 cells (insert) showing that the SKBR3 (LP) cells express approximately 5-fold lower HER2 protein than the SKBR3(HP) cell line. Direct comparison of the cytotoxic effects of continuous exposure of various concentrations of TNF demonstrate that the cell line expressing higher levels of HER2 was effectively resistant to TNF while the cells expressing low levels of HER2 were sensitive to the cytotoxic effects of TNF.

10
15 **Figure 31** demonstrates the cytotoxicity of either TNF or purified sfv23/TNF against log-phase SKBR3-HP cells in culture. As this figure demonstrates, the IC_{50} of the sfv23/TNF construct (20 U/ml) was approximately 2000-fold lower than that of free TNF ($IC_{50} = 4,000$ U/ml).

20 **Figure 32** shows the cytotoxicity of TNF and sfv23/TNF against log-phase SKBR3-LP cells in culture. As this figure illustrates,

the IC₅₀ of TNF against these resistant cells was 5×10^7 U/ml while that of the sfv23/TNF construct was approximately 20-fold lower (2×10^6 U/ml).

5

DETAILED DESCRIPTION OF THE INVENTION

As used herein, a cellular targeting moiety is capable to selectively binding to a c-erbB-2 protein which is expressed on a cell, typically on its surface. This includes both a ligand specific to binding to the c-erbB-2 protein and antigen binding region, *e.g.*, intact antibodies or epitope binding fragments thereof. This encompasses both classical antibody molecules, chimeric versions, single chain, and modified antibody fragments which retain epitope binding specificity and affinity.

As used herein, the term "immunoglobulin" or "antibody peptide(s)" refers to an entire immunoglobulin or antibody or any functional binding fragment of an immunoglobulin molecule. Examples of such peptides include complete antibody molecules, antibody fragments, such as Fab, F(ab')₂, CDRs, V_L, V_H, and any other portion of an antibody, particularly those exhibiting antigen binding

specificity or affinity. For instance, an IgG antibody molecule is composed of two light chains, each linked by disulfide bonds to two heavy chains. The two heavy chains are, in turn, linked to one another by disulfide bonds in an area known as the hinge region of the antibody. A single IgG molecule typically has a molecular weight of approximately 150–160 kD and contains two antigen binding sites. Fragments of these molecules, *e.g.*, heavy or light chains alone, can sometimes bind antigen. Antibodies, fragments of antibodies, and individual chains can be functionally equivalent to immunoglobulins.

A normal antibody heavy or light chain has an N-terminal (NH₂) variable (V) region, and a C-terminal (–COOH) constant (C) region. The heavy chain variable region is referred to as V_H (including, for example, V_γ), and the light chain variable region is referred to as V_L (including V_κ or V_λ). The variable region is the part of the molecule that binds to the antibody's cognate antigen, while the Fc region (the second and third domains of the C region) determines the antibody's effector function (*e.g.*, complement fixation, opsonization). Full-length immunoglobulin or antibody "light chains" (generally about 25 Kd, about 214 amino acids) are encoded by a variable region gene at the N-terminus (generally about 110

amino acids) and a κ (kappa) or λ (lambda) constant region gene at the COOH-terminus. Full-length immunoglobulin or antibody "heavy chains" (generally about 50 Kd, about 446 amino acids), are similarly encoded by a variable region gene (generally encoding about 116 amino acids) and one of the constant region genes, *e.g.* gamma (encoding about 330 amino acids). Typically, the "V_L" will include the portion of the light chain encoded by the V_L and/or J_L (J or joining region) gene segments, and the "V_H" will include the portion of the heavy chain encoded by the V_H, and/or D_H (D or diversity region) and J_H gene segments. *See, Roitt, et al., Immunology, Ch. 6, (2d ed. 1989) and Paul, Fundamental Immunology, Raven Press (2d ed. 1989).*

An immunoglobulin light or heavy chain variable region consists of a "framework" region interrupted by three hypervariable regions, also called complementarity-determining regions or CDRs. The extent of the framework region and CDRs have been defined (*see, "Sequences of Proteins of Immunological Interest," E. Kabat, et al., U.S. Department of Health and Human Services, (1987)*). The sequences of the framework regions of different light or heavy chains are relatively conserved within a species. The framework

regions of an antibody, that is the combined framework regions of the constituent light and heavy chains, serve to position and align the CDRs in three dimensional space. The CDRs are primarily responsible for binding to an epitope of an antigen. The CDRs are typically referred to as CDR1, CDR2, and CDR3, numbered sequentially starting from the N-terminus.

The two types of light chains, κ and λ , are referred to as isotypes. Isotypic determinants typically reside in the constant region of the light chain, also referred to as the C_L in general, and C_κ or C_λ in particular. The constant region of the heavy chain molecule, also known as C_H , determines the isotype of the antibody. Antibodies are classified as IgM, IgD, IgG, IgA, and IgE depending on the heavy chain isotype. The isotypes are encoded in the mu, delta, gamma, alpha, and epsilon segments of the heavy chain constant region, respectively. In addition, there are a number of gamma subtypes.

The heavy chain isotype determines different effector functions of the antibody, such as opsonization or complement fixation. In addition, the heavy chain isotype determines the secreted form of the antibody. Secreted IgG, IgD, and IgE isotypes are typically found in single unit or monomeric form. Secreted IgM

isotype is found in pentameric form; secreted IgA can be found in both monomeric and dimeric form.

An $F(ab')_2$ fragment lacks the C-terminal portion of the heavy chain constant region, and usually has a molecular weight of approximately 110 kD. It retains two antigen binding sites and the interchain disulfide bonds in the hinge region, but it does not have the effector functions of an intact IgG molecule. An $F(ab')_2$ fragment may be obtained from an IgG molecule by proteolytic digestion with pepsin at pH 3.0–3.5 using standard methods such as those described in Harlow and Lane, *infra*.

An "Fab" fragment comprises a light chain and the N-terminal portion of the heavy chain which are linked together by disulfide bonds. It typically has a molecular weight of approximately 50 kD and contains a single antigen binding site. Fab fragments may be obtained from $F(ab')_2$ fragments by limited reduction, or from whole antibody by digestion with papain in the presence of reducing agents. (See, Harlow and Lane, *infra*.) In certain cases, the concentration of reducing agent necessary to maintain the activity of papain in the presence of atmospheric oxygen is sufficient to fully reduce the interchain disulfide bonds to the antibody. This can result in loss of antigen recognition. To circumvent this problem,

papain may be activated and then exchanged into buffer containing a concentration of reducing agent compatible with maintaining antigen binding activity. The antibody digestion is typically carried out under an inert atmosphere to prevent deactivation of the papain.

5 The following protocol is an example of this process: (A)

Activation of papain: Papain, supplied as 10 mg/ml NH_4SO_4 suspension, is dissolved in 10 mM EDTA, 20 mM cysteine, pH=8.0, to a final concentration of 2 mg/ml. The solution is degassed and allowed to incubate 2 hours at room temperature under nitrogen. (B)

10 The activated papain is exchanged into 20 mM NaPO_4 , pH=7.0, 150 mM NaCl, 10 mM EDTA, 30 μM DTT. (C) Digestion of antibody: 1 mg of activated papain is added for every 100 mg of antibody, and the solution is dialyzed against a large excess of 20 mM NaPO_4 , pH=7.0, 150 mM NaCl, 10 mM EDTA, 30 μM DTT, with continuous helium
15 sparging. Dialysis is used to maintain a molar excess of reducing agent during the course of the digestion. (D) After 2–4 hours at room temperature the digestion is terminated by addition of iodoacetamide. (E) Fab fragments are separated from undigested or partially digested antibody using standard chromatography methods.

20 As used herein, the terms "Fab", or any other antibody fragments, have similar classifications when applied to the present

invention as to the general terms "antibodies" or "immunoglobulins". Thus, "mammalian" Fab protein, "chimeric Fab", and the like are used analogously to the corresponding definitions in general usage, and as set forth in the subsequent paragraphs.

5 As used herein, the term "chimeric antibodies" or "chimeric peptides" refer to those antibodies or antibody peptides wherein one portion of the peptide has an amino acid sequence that is derived from, or is homologous to, a corresponding sequence in an antibody or peptide derived from a first gene source, while the
10 remaining segment of the chain(s) is homologous to corresponding sequences of another gene source. For example, a chimeric heavy chain antibody peptide may comprise a murine variable region and a human constant region. The two gene sources will typically be two separate species, but will occasionally involve one species.

15 Chimeric antibodies or peptides are typically produced using recombinant molecular and/or cellular techniques. In many cases, chimeric antibodies have variable regions of both light and heavy chains that mimic the variable regions of antibodies derived from one mammalian species, while the constant and/or framework
20 portions are homologous to the sequences in antibodies derived from a second, different mammalian species.

As used herein, the definition of chimeric antibody, however, is not limited to this example. A chimeric antibody is any antibody in which either or both of the heavy or light chains are composed of combinations of sequences mimicking the sequences in antibodies of different sources, whether these sources be differing classes, differing antigen responses, or differing species of origin, and whether or not the fusion point is at the variable/constant boundary. For example, chimeric antibodies can include antibodies where the framework and CDRs are from different sources. For example, non-human CDRs are integrated into human framework regions linked to a human constant region to make "humanized antibodies." See, e.g., PCT Application WO 87/02671; U.S. Patent No. 4,816,567; EP Patent Application 0173494; Jones, *et al.*, *Nature*, 321:522-525 (1986); and Verhoeyen, *et al.*, *Science*, 239:1534-1536 (1988).

As used herein, the term "human-like framework region" is a framework region for each antibody chain, and it usually comprises at least about 70 amino acid residues, typically 75 to 85 or more residues. The amino acid residues of the human-like framework region are at least about 80%, preferably about 80-85%, and most preferably more than 85% homologous with those in a human immunoglobulin. This shared feature with other endogenous

antibodies is useful in generating a targeting moiety which introduces only a minor immune reaction, *e.g.*, a mechanism which minimizes response to "self" markers.

As used herein, the term "humanized" or "human-like immunoglobulin" refers to an immunoglobulin comprising a human-like framework region and a constant region that is substantially homologous to a human immunoglobulin constant region, *e.g.*, having at least about 80% or more, preferably about 85-90% or more and most preferably about 95% or more homology. Hence, most parts of a human-like immunoglobulin, except possibly the CDRs, are substantially homologous to corresponding parts of one or more native human immunoglobulin sequences.

As used herein, the term "hybrid antibody" refers to an antibody wherein each chain is separately homologous with reference to a mammalian antibody chain, but the combination represents a novel assembly so that two different antigens are recognized by the antibody. In hybrid antibodies, one heavy and light chain pair is homologous to that found in an antibody raised against one antigen recognition feature, *e.g.*, epitope, while the other heavy and light chain pair is homologous to a pair found in an antibody raised against another epitope. This results in the property

of multi-functional valency, i.e., ability to bind at least two different epitopes simultaneously. Such hybrids may, of course, also be formed using chimeric chains.

As used herein, the term "monoclonal antibody" means an antibody composition recognizing a discrete antigen determinant. It is not intended to be limited as regards the source of the antibody or the manner in which it is made.

Using standard methods known in the art, the variable regions and CDRs may be derived from a hybridoma that produces a monoclonal antibody that is specific for c-erbB-2. The nucleic acid sequences of the present invention capable of ultimately expressing the desired chimeric antibodies can be formed from a variety of different nucleotide sequences (genomic or cDNA, RNA, synthetic oligonucleotides, etc.) and components (*e.g.*, V, J, D, and C regions), as well as by a variety of different techniques. Joining appropriate-genomic sequences is presently a common method of production, but cDNA sequences may also be utilized (*see*, European Patent Publication No. 0239400 and Reichmann, *et al.*, *Nature*, 332:323-327 (1988)).

Human constant region DNA sequences are preferably isolated from immortalized B-cells, *see e.g.*, Heiter, *et al.*, *Cell*,

22:197-207 (1980) but can be isolated or synthesized from a variety of other sources. The nucleotide sequence of a human immunoglobulin C γ 1 gene is described in Ellison, *et al.*, *Nucl. Acid. Res.*, 10:4071 (1982); Beidler, *et al.*, *J. Immunol.*, 141:4053 (1988);
5 Liu, *et al.*, *Proc. Natl. Acad. Sci. USA*, 84:3439 (1987).

The CDRs for producing the immunoglobulins of the present invention preferably are derived from monoclonal antibodies capable of binding to the desired antigen, c-erbB-2 protein, and produced in any convenient mammalian source, including, mice, rats, rabbits, hamsters, or other vertebrate host cells capable of producing antibodies by well known methods. Suitable source cells for the DNA sequences and host cells for immunoglobulin expression and secretion can be obtained from a number of sources, such as the American Type Culture Collection ("ATCC").

In addition to the chimeric antibody peptides specifically described herein, other "substantially homologous" modified immunoglobulins can be readily designed and manufactured utilizing various recombinant DNA techniques known to those skilled in the art. Modifications of the genes may be readily accomplished by a variety of well-known techniques, such as site-directed mutagenesis (see, Gillman and Smith, *Gene*, 8:81-97 (1979) and Roberts, S., *et al.*,

Nature, 328:731–734 (1987)). These modifications can include amino acid additions, deletions, substitutions, preferably conservative, and other changes in the sequence of the polypeptide while retaining the appropriate property or biological activity. Alternatively, polypeptide fragments comprising only a portion of the primary antibody structure and possessing binding and/or effector activities may be produced. Also because, like many genes, the immunoglobulin-related genes contain separate functional regions, each having one or more distinct biological activities, the genes may be fused to functional regions from other genes to produce fusion proteins (*e.g.*, immunotoxins) having novel properties or novel combinations of properties.

The cloned variable and constant regions can be isolated from plasmids and ligated together into a mammalian expression vector, *e.g.*, pSV2-neo, or pRSV-gpt, to form a functional transcription unit. These expression vectors can then be transfected into host cells. Mouse myeloma cells, such as SP 2/O or P3X cells, are a preferred host because they do not secrete endogenous immunoglobulin protein and contain all of the components used in immunoglobulin expression. Myeloma cells can be transfected using appropriate techniques as described above.

Other types of promoters and enhancers specific for other host cells are known in the art. *See*, Kameyoma, K., *et al.*, *supra*. For example, the DNA sequence encoding the chimeric antibody amino acid sequence can be linked to yeast promoters and enhancers and transfected into yeast by methods well known in the art. *See*, Kriegler, *supra*. This same approach can be taken to isolate the c-erbB-2 specific CDRs from one source such as one mammalian species and the framework regions of another source, such as a different mammalian species. The CDRs can then be ligated to the framework regions and constant regions to form a chimeric antibody. The CDRs could be cloned in an expression vector comprising, for example, human framework and constant regions.

Another example is a recombinant DNA sequence comprising the heavy and/or light chain CDR1, CDR2, and CDR3 of one species, such as mouse, and the framework regions of human heavy chain to encode an antibody specific for c-erbB-2. Other possibilities include using CDRs specific for c-erbB-2; using part of the variable region encompassing CDR1 and CDR2 from one mammalian species, and then ligating this sequence to another encoding the framework portions of a second mammalian species to the CDR3 of the first; or transfecting a host cell line with a recombinant DNA sequence

encoding a c-erbB-2 specific heavy chain CDRs derived from a first mammalian species, interspersed within the framework of a second mammalian species with a light chain containing a variable region DNA sequence derived from the first species and the constant region
5 derived from the second species.

Recombinant DNA expression vectors comprising antibody sequences may be transfected by electroporation into host cells. Standard selection procedures are used to isolate clones that produce the c-erbB-2 specific chimeric antibody.

Antibodies may be expressed in an appropriate folded form, including single chain antibodies, from bacteria such as *E. coli*. See, Pluckthun, *Biotechnology*, 9:545 (1991); Huse, *et al.*, *Science*, 246:1275 (1989); and Ward, *et al.*, *Nature*, 341:544 (1989). The antibody peptide sequences may be amplified for cloning by use of polymerase chain reaction, or PCR, a technique used to amplify a DNA sequence of interest using a thermostable DNA polymerase, such as Taq polymerase, and polymerase and oligonucleotide primers, all as described in *PCR Protocols*, ed. Innis, *et al.*, Academic Press, Inc. (1990), incorporated by reference herein. See also Orlandi, *supra*
15 and Larrick, *et al.*, *Biotechnology*, 7:934 (1989).
20

The c-erbB-2 protein (also referred to here simply as c-erbB-2) is a 185 Kd (Kilodalton) membrane glycoprotein having tyrosine kinase activity and is related to, but distinct from, the epidermal growth factor receptor (EGFR). Like the EGFR protein, the c-erbB-2 protein has an extracellular domain that includes two cysteine-rich repeat clusters, a transmembrane domain, and an intracellular kinase domain. In addition, the amino acid sequence of the c-erbB-2 protein as well as the nucleotide sequence has been described by Coussens, *et al.*, *Science*, 230:1132 (1985).

The c-erbB-2 protein is encoded by the c-erbB-2 oncogene described in 1985 by three different research groups: Semba, *et al.*, *Proc. Natl. Acad. Sci. USA*, 82:6497 (designating the gene as c-erbB-2); Coussens, *et al.*, *supra*, (designating the gene as HER-2); and King, *et al.*, *Science*, 229:1132 (designating the gene as v-erbB related). Thus, the c-erbB-2 gene sequence and its corresponding protein sequence are well-known and described in the art. The c-erbB-2 protein has a defined intracellular design, a transmembrane region, and an extracellular region. Typically, the targeting moieties of the present invention will bind to the extracellular region, which should be exposed to the exterior of the neoplastic cell. The targeting moiety will generally recognize a

feature or features found there, including a ligand binding region, or antigen recognition sites, *e.g.*, epitopes. The epitopes will often be directed to pure polypeptide epitopes, either linear peptide sequence determinants or conformational determinants, but can also be directed to epitopes having carbohydrate components. The epitopes can thus include combined protein/carbohydrate components, or carbohydrate components alone. Other modifications to the protein, normal or abnormal, will present important epitopic determinants, also.

Detection of the c-erbB-2 protein may be accomplished by well-known immunoassays employing antibodies specific to the c-erbB-2 protein, such as those described here. Such antibodies are commercially available, for example, from Chemicon International, Inc., Temecula, CA or may be prepared by standard immunological procedures. *See, e.g.*, Harlow and Lane, *Antibodies: A Laboratory Manual*, Cold Spring Harbor Publications, N.Y. (1988).

It is intended herein that the c-erbB-2 protein definition will also include those proteins developed from other host systems, *e.g.*, proteins that are immunologically related to the human c-erbB-2 protein. For example, a related rat gene (designated *neu*) has been reported in Schecter, *et al.*, *Science*, 229:976 (1985).

Useful epitopes to which antibodies may be easily raised include extracellular epitopes found on target cells. These epitopes will generally be protein epitopes, *e.g.*, linear or conformational epitopes of the protein as found on neoplastic cells. Other useful
5 epitopes will include non-proteineous components including carbohydrate or other modifications, usually post-translational, found on the c-erbB-2 protein. Antibodies and other binding regions which exhibit binding specificity for overexpressed c-erbB-2 may be raised against fragments of the protein.

10 Mouse monoclonal antibodies have been made against the extracellular portion of c-erbB-2. One example of such an antibody is TAb 250, which is deposited with the American Type Culture Collection, (ATCC) bearing Accession No. HB10646.

15 Alternatively, a targeting moiety may be derived from any other targeting method which exhibits affinity and specificity for a c-erbB-2 expressing cell. For example, a ligand which is recognized and bound by the c-erbB-2 protein would be a useful targeting moiety. *See, e.g.,* Ciccodicola, *et al.* (1989) *Embo J.* 8:1987-1991; Ciardiello, *et al.* (1991) *Cancer Research* 51:1051-1054; and Ciardiello,
20 *et al.* (1991) *P.N.A.S. USA* 88:7792-7796, which describe CRIPTO, a

molecule which appears to serve as a ligand for the EGF receptor, and will likely also bind with specificity to the c-erbB-2 protein.

In the case of the sequences described herein, it should be understood that variants of these sequences are also included, such as substitution, addition, and/or deletion mutations, or any other sequence possessing substantially similar binding activity to the sequences from which they are derived or otherwise similar to.

For this invention, an antibody or other peptide is specific for a c-erbB-2 protein if the antibody or peptide binds or is capable of binding c-erbB-2, *e.g.*, protein as measured or determined by standard antibody-antigen or ligand-receptor assays, for example, competitive assays, saturation assays, or standard immunoassays such as ELISA or RIA. This definition of specificity applies to single heavy and/or light chains, CDRs, fusion proteins or fragments of heavy and/or light chains, that are also specific for c-erbB-2 protein if they bind c-erbB-2 protein alone or if, when properly incorporated into immunoglobulin conformation with complementary variable regions and constant regions as appropriate, are then capable of binding c-erbB-2 protein with specificity.

In competition assays the ability of an antibody or peptide fragment to bind an antigen can be determined by detecting

the ability of the peptide to compete with the binding of a compound known to bind the antigen. Numerous types of competitive assays are known and are discussed herein. Alternatively, assays that measure binding of a test compound in the absence of an inhibitor
5 may also be used. For instance, the ability of a molecule or other compound to bind the c-erbB-2 protein can be detected by labeling the molecule of interest directly or it may be unlabeled and detected indirectly using various sandwich assay formats. Numerous types of binding assays such as competitive binding assays are known (*see*,
10 *e.g.*, U.S. Patent Nos. 3,376,110, 4,016,043, Harlow and Lane, *Antibodies: A Laboratory Manual*, Cold Spring Harbor Publications, N.Y. (1988), and Coligan, *et al.* (eds), *Current Protocol in Immunology*, Wiley and Sons, N.Y.). Assays for measuring binding of a test compound to one component alone rather than using a competition
15 assay are also available. For instance, immunoglobulins can be used to identify the presence of the c-erbB-2 protein. Standard procedures for monoclonal antibody assays, such as ELISA, may be used (*see*, Harlow and Lane, *supra*). For a review of various signal producing systems which may be used, *see*, U.S. Patent No. 4,391,904.

20 Further, the specificity of the binding moieties to c-erbB-2 can be determined by their affinity. Such specificity exists

if the dissociation constant ($K_D=1/K$, where K is the affinity constant) of the moiety is $<1\mu\text{M}$, preferably $<100\text{ nM}$, and most preferably $<1\text{ nM}$. Antibody molecules will typically have a K_D in the lower ranges. $K_D = [R-L]/[R][L]$ where $[R]$, $[L]$, and $[R-L]$ are the concentrations at equilibrium of the receptor or c-erbB-2 (R), ligand, antibody, or peptide (L) and receptor-ligand complex (R-L), respectively. Typically, the binding interactions between ligand or peptide and receptor or antigen include reversible noncovalent associations such as electrostatic attraction, Van der Waals forces, and hydrogen bonds.

Other assay formats may involve the detection of the presence or absence of various physiological or chemical changes that result from the interaction, such as down modulation, internalization, or an increase in phosphorylation. *See also, Receptor-Effector Coupling – A Practical Approach*, ed. Hulme, IRL Press, Oxford (1990).

A preferred peptide specific for c-erbB-2 protein induces an increase in the phosphorylation of the c-erbB-2 protein when placed in contact with tumor cells expressing the c-erbB-2 protein. A molecule that "induces an increase in the phosphorylation of c-erbB-2 protein" is one that causes a detectable increase in the incorporation of phosphate into the protein over that which occurs in the absence of the molecule. Typically this detectable increase will be a two-fold

or greater increase in phosphorylation, preferably greater than a three-fold increase over controls. Phosphorylation may be measured by those methods known in the art for detecting phosphorylation of receptors. See, for example Cooper, *et al.*, *Methods in Enzymology*, 99:387-402 (1983); Antoniadis and Pantazis, *Methods in Enzymology*, 147:36-40 (1987); and Lesniak, *et al.*, *Methods in Enzymology*, 150:717-723 (1987).

Typically, phosphorylation can be measured by *in vivo* phosphorylation of intact cells (Lesniak, *supra*) or by an *in vitro* autophosphorylation reaction (Antonaidis, *supra*). For measuring *in vivo* phosphorylation, for example, assays may be conducted where cells bearing the c-erbB-2 protein are placed into contact with radioactive labeled phosphate. To detect phosphorylation of the c-erbB-2 protein receptor in the *in vivo* assay, it is advantageous to incubate the test cells for about 12 to about 18 hours, with the labeled phosphate. The cells are divided into two or more batches, where some are exposed to the molecule expected to increase the phosphorylation of the receptor and some are separated out as controls. The aliquots are subsequently immunoprecipitated, the receptor is recognized, for example, by SDS polyacrylamide gel or autoradiography methods, and an increase in phosphorylation is

considered statistically significant when there is a two-fold or greater increase in the background of the aliquot exposed to the test molecule over the control aliquots. To measure *in vitro* autophosphorylation, for example, cells or cell extracts may be
5 incubated in the presence or absence of the peptide specific for c-erbB-2. Following immunoprecipitation with an anti-c-erbB-2 antibody, the immune complex may be incubated with $\gamma^{32}\text{P}$ -ATP and analyzed by SDS-PAGE autoradiography.

Another preferred peptide specific for c-erbB-2 protein is one that causes down modulation of the c-erbB-2 protein. "Down modulation of the c-erbB-2 protein" is determined by a detectable decrease in the presence on the tumor cells of the c-erbB-2 receptor. Such down modulation is detected by a decrease in the ability of antibodies or other specific binding moieties to bind to or recognize
10 the c-erbB-2 receptor protein on the tumor cells. For example, down modulation can be determined by incubating tumor cells bearing the c-erbB-2 protein receptor with the peptide of interest, washing the cells, then contacting the cells with labeled (preferably radiolabeled) antibodies specific for the c-erbB-2 protein. The extent of binding of
15 the labeled anti-c-erbB-2 antibodies to the cells exposed to the peptide specific for c-erbB-2 protein is compared to the extent of
20

binding of the antibodies to control cells (i.e., not exposed to the c-erbB-2 specific peptide). Preferably for these assays, the cells are directly subjected to the labeled anti-c-erbB-2 antibodies after washing.

5 The down modulation observed is typically dose dependent, i.e., the extent of down modulation increases with the amount of peptide specific for c-erbB-2 protein exposed to the c-erbB-2 protein. A peptide that causes a decrease in 90% or greater of binding of the treated cells versus control cells to anti-c-erbB-2 antibodies is preferred.

10 Another preferred peptide specific for c-erbB-2 protein is one that binds tumor cells expressing c-erbB-2 protein and is internalized when placed in contact with such tumor cells. "Internalization" occurs when the peptide becomes sequestered in the cytoplasm of the cells. Once internalized, the receptor and/or peptide may be degraded in the cell lysosomes or may be recycled to the cell surface. A method for determining internalization of a ligand- receptor complex is also described in Haigler, *et al.*, *J. Biol. Chem.*, 255:1239-1241 (1980).

15 A cell growth modulator is a molecule which affects the growth of a cell to which it is targeted. Typically, the modulator

must be internalized into the target cell, but this function is usually provided by internalization which results from the targeting moiety. The modulation will typically be a decrease in metabolism or growth rate, preferably a cytotoxic effect, but a significant increase in metabolism or growth rate will also be useful. When a significant increase in metabolism or growth rate is effected, a short term poison might be used in combination to kill only those cells exhibiting such.

For modulators which decrease metabolism or growth rate, it is preferred that the modulator be highly potent, *e.g.*, have a very high activity. The toxin may include inorganic or simple organic molecules, but biological molecules will usually be more potent. Although viral and fungal toxins exist, particular bacterial or plant toxins have among the highest specific activities known. Growth arrest may occur by preventing any of a number of essential cellular functions including nucleic acid synthetic, protein synthesis, and cellular metabolism, general or specific. For example, *pseudomonas* exotoxin and diphtheria toxin function by irreversibly arresting protein synthesis in eukaryotic cells. Both examples enzymatically inactivate elongation factor 2, which is an essential component of protein synthesis. Other elongation factors may be targets for other

toxins. Ricin, in contrast, is a plant toxin which acts directly on the ribosome, acting on the 28S rRNA. Preferably, the growth modulators will have enzymatic activities with high turnover numbers so internalization of very few molecules can kill the target cell. See, Pastain, *et al.*, *Science* 254:1173– 1177.

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Gelolin is a glycoprotein (M.W. approximately 29–30,000 Kd) purified from the seeds of *Gelonium multiflorum* and belongs to a class of potent ribosomal– inactivating plant toxins. Other members of this class include chains of abrin, ricin, and modeccin. Gelonin, like abrin and ricin, inhibits protein synthesis by damaging the 60S subunit of mammalian ribosomes. Gelonin appears to be stable to chemical and physical treatment. Furthermore, gelonin itself does not bind to cells and is normally non-toxic (except in high concentrations) when administered alone, and is safe to manipulate in the laboratory. The inactivation of ribosomes is irreversible, does not appear to involve co-factors, and occurs with an efficiency which suggests that gelonin acts enzymatically.

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Gelolin and ricin inhibit protein synthesis and are among the most active toxins on a protein weight basis. Gelonin is 10 to 1000 times more active in inhibiting protein synthesis than ricin A chain. Peptides like ricin and abrin are composed of two chains, an A

chain which is the toxic unit and a B chain which acts by binding to cells. Unlike ricin and abrin, gelonin is composed of a single chain, and, because it lacks a B chain for binding to cells, it is itself relatively inert, or non-toxic to intact cells. This feature of having a much lower cellular effect when not conjugated to a binding or targeting moiety is an important feature of various embodiments of the present invention. This differential toxicity is important in high specificity for c-erbB-2 expressing cells.

Mammalian cells apparently lack the ability to bind and/or to internalize the native gelonin molecule. Conjugates of gelonin with a tumor-targeting reagent, such as the monoclonal antibody TAb 250 directed to a tumor associated antigen present on certain tumor cells, provide both a specific method for binding the gelonin to the cell and a route for internalization of the gelonin-antibody complex.

The cytotoxic moiety of the immunotoxin may be a cytotoxic drug or an enzymatically active toxin of bacterial or plant origin, or an enzymatically active fragment ("A chain") of such a toxin. Enzymatically active toxins and fragments thereof are preferred and are exemplified by gelonin, diphtheria A chain, nonbinding active fragments of diphtheria toxin, exotoxin A chain

(from Pseudomonas aeruginosa), ricin A chain, abrin A chain, modeccin A chain, alpha-sarcin, Aleurites fordii proteins, dianthin proteins, Phytolacca americana proteins (PAPI, PAPII, and PAP-S), momordica charantia inhibitor, curcin, crotin, saponaria officinalis inhibitor, mitogellin, restrictocin, phenomycin, and enomycin. The active toxins can function by any of a number of mechanisms, each of which affects cellular physiology and growth. The toxins may be metabolic inhibitors or poisons, nucleic acid synthesis inhibitors, protein synthesis inhibitors, or any other mediators of abnormal or deleterious functions. Most preferred is conjugation with gelonin.

Active fragments and derivatives include any compounds which have the same core structure as the full length structure of gelonin but lack the entire primary sequence. These fragments or derivatives will have the same or improved biological or cytotoxic activity as gelonin. The cytotoxicity of the gelonin fragments or derivatives may be routinely determined by those with skill in the art using the rabbit reticulocyte lysate assay.

Biological response modifiers which may be coupled to the TAb 250 antibody and used in the present invention include, but are not limited to, lymphokines and cytokines such as IL-1, IL-2, interferons (α , β or γ), TNF, LT, TGF- β , and IL-6. These biological

response modifiers have a variety of effects on tumor cells. Among these effects are increased tumor cell killing by direct action as well as increased tumor cell killing by increased host defense mediated processes. Conjugation of antibody TAb 250 to these biological
5 response modifiers will allow selective localization or targeting to tumors or cells overexpressing c-erbB-2 and, hence, improved anti-proliferative effects. Non-specific effects leading to toxicity of non-target cells will be minimized since the selected cell growth mediator is ineffective absent a targeting component.

Cytotoxic drugs (and derivatives thereof) which are useful in the present invention include, but are not limited to, adriamycin, cis-platinum complex, bleomycin and methotrexate. These cytotoxic drugs are useful for clinical management of recurrent tumors, but their use is complicated by severe side effects and
10 damage caused to non-target cells. The TAb 250 antibody may serve as a useful carrier of such drugs providing an efficient means of both delivery to the tumor and enhanced entry into the tumor cells themselves. In addition, specific antibody delivery of cytotoxic drugs to tumors will provide protection from the deleterious action of the
15 chemotherapeutic agents of sensitive sites which do not overexpress c-erbB-2 such as the liver and bone marrow. Use of drugs conjugated

to the TAb 250 antibody as a delivery system allows lower dosage of the drug itself, since all drug moieties are conjugated to antibodies which concentrate on neoplastic cells, and will usually be internalized therein.

5 The targeting moiety and the cell growth modulator may be conjugated using a variety of bifunctional protein coupling agents. Representative reagents are N-succinimidyl 3-(2-pyridyldithio)(propionate)(SPDP), 2-IT, 4-succinimidylloxycarbonyl- α -methyl- α -(2-pyridyldithio) toluene (SMPT) bifunctional derivatives of imidoesters such as dimethyl adipimidate, HCl, active esters such as disuccinimidyl suberate, aldehydes such as glutaraldehyde, bis-azido compounds such as bis(p-azidobenzoyl) hexanediamine, bis-diazonium derivatives such as bis-(p-diazoniumbenzoyl)- ethylenediamine, diisocyanates such as tolylene 2,6- diisocyanate, and bis-active fluorine compounds such as a 1,5-difluoro-2,4-dinitrobenzene.

10 Prior to use in these studies, the Sp2/0-Ag14 cells is grown initially in the presence of 0.1 μ g/ml of native gelonin. Over several months, the concentration of gelonin is gradually increased
15 until the cells can be maintained in up to 10 mg/ml. Cells will then be cloned by limiting dilution in the presence of 10 mg/ml gelonin
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and the resulting colonies resistant to gelonin is expanded. Gelonin is then removed from the culture media for two passages and the cells challenged again with gelonin exposure to confirm development of stably-resistant clones. After tests to confirm the production and activity of chimeric TAb-250, gelonin-resistant SP2/0 cell producing antibody is grown and the cDNA for the TAb 250 antibody removed from the total DNA by incubation with restriction endonuclease. In parallel, the cDNA from JM105 E-Coli expressing optimized gelonin is removed, purified and the DNA encoding gelonin released after digestion with HindIII and Eco RI. The gelonin gene is ligated into the heavy-chain fragment and the insert replaced into gelonin resistant SP2/0 cells. Cells is sub-cloned by limiting dilution and the clones screened for both chimeric antibody production and gelonin content. Finally, positive clones will be expanded and the recombinant fusion protein will be purified and tested in both *in vitro* cytotoxicity assays and *in vivo* tissue distribution, pharmacokinetics, therapeutics and toxicity trials. A comparison of TAb-250-gelonin fusion protein properties to the characteristics of the previously described TAb 250 gelonin constructs is performed to determine the advantages and drawbacks of each. Based upon these

studies a Phase I clinical study of chimeric TAb 250 gelonin fusion protein is performed in patients with advanced breast cancer.

Administration of the immunotoxins of the present invention to an individual who has been diagnosed as having
5 neoplastic cells, *e.g.*, a tumor with an undesirable level of expression of the c-erbB-2 oncogene, allows targeting and concentration of the cytotoxic agent at the site where it is needed to kill them. By so targeting the cytotoxic agents, non-specific toxicity to other organs, tissues and cells will be eliminated, minimized or at least decreased.

When used *in vivo* for therapy, the immunotoxins of the present invention are administered to the patient or an animal in therapeutically effective amounts, *i.e.*, amounts that eliminate or reduce the tumor burden. They are administered parenterally, preferably intravenously, but other routes of administration are used
10 as appropriate. The dose and dosage regimen depend upon the nature of the cancer (primary or metastatic) and its population, the characteristics of the particular immunotoxin, *e.g.*, its therapeutic index, the patient, the patient's history and other factors. The amount of immunotoxin administered typically is in the range of
15 about 0.1 to about 10 mg/kg of patient weight. The schedule is continued to optimize effectiveness while balanced against negative
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effects of treatment. See Remington's Pharmaceutical Science, 17th Ed. (1990) Mark Publishing Co., Easton, Penn.; and Goodman and Gilman's: The Pharmacological Basis of Therapeutics 8th Ed (1990) Pergamon Press.

5 For parenteral administration the immunotoxins are most typically formulated in a unit dosage injectable form (solution, suspension, emulsion) in association with a pharmaceutically acceptable parenteral vehicle. Such vehicles are preferably non-toxic and non-therapeutic. Examples of such vehicles are water, saline, Ringer's solution, dextrose solution, and 5% human serum albumin. Nonaqueous vehicles such as fixed oils and ethyl oleate may also be used. Liposomes may be used as carriers. The vehicle may contain minor amounts of additives such as substances that enhance isotonicity and chemical stability, e.g., buffers and preservatives. 10 The immunotoxin will typically be formulated in such vehicles at concentrations of about 0.1 mg ml to 10 mg ml. 15

The immunotoxins of the present invention may also be used to extend the survival time of tumor bearing mammals and to retard the rate of growth of tumors comprised of cancer cells carried by a mammal. For example, nude mice bearing xenografts of human 20 tumors growing subcutaneously or intraperitoneally can be treated

with doses of immunotoxin, antibody alone, toxin alone or saline at a dose between 25 and 100 mg/kg. Tumor growth inhibition can be measured by the change in physical size of the subcutaneous tumors or by prolongation of survival in mice-bearing intraperitoneally tumors such as SKOV-3 cells. Such studies are useful or indicative of methodologies and which might be applicable to other mammals, including primates.

The present invention also provides an immunotoxin comprised of a single chain antibody raised against an epitope on HER-2/neu. In one embodiment, the single chain antibody is raised against the 185 kD antigen of HER-2/neu. For example, such a preferred single chain antibody is antibody e-23. Preferably, the composition of the present invention is made by recombinantly fusing the gene encoding said antibody to the gene encoding gelonin.

The instant invention also provides a conjugate of tumor necrosis factor to an antibody specific for c-erbB-2 protein. The antibody may be an intact full-length immunoglobulin in which either the heavy chain or light chain is conjugated to tumor necrosis factor. Alternatively, the antibody may be an Fv fragment with tumor necrosis factor conjugated to either the V_H or the V_L peptide. Preferably, the conjugate is a fusion protein between a single chain

antibody and tumor necrosis factor produced by fusing a gene encoding the single chain antibody to the gene encoding tumor necrosis factor. Specific example with single chain antibody scFv-23 are described herein.

5 The following examples provide a detailed description of the preparation, characterization, and use of the immunotoxins of this invention. These examples are not intended to limit the invention in any manner.

EXAMPLE 1

Purification of Gelonin

10 A procedure for the isolation of gelonin is available in Stirpe, *et al.*, *J. Biol. Chem.*, 255, 6947– 53 (1980). Seeds of *Gelonium*
15 *multiflorum* were shelled and the nuts ground in a homogenizer with eight volumes of 0.14 M NaCl containing a 5 mM sodium phosphate (pH 7.4). The homogenate was left overnight at 4°C with continuous stirring, cooled on ice and centrifuged at 35,000 times g for 20
20 minutes at 0°C. The supernatant was removed, dialyzed against 5 mM sodium phosphate (pH 6.5) and concentrated using a PM10 filter. The sample was layered on a CM-52 ion-exchange column (20 x 1.5

cm) equilibrated with 5 mM sodium phosphate (pH 6.5). Material which bound to the ion exchange resin was eluted with 400 ml of a linear NaCl gradient from 0 to 0.3 M at a rate of 25 ml per hour at 4°C. Five ml fractions were collected. The fractions were monitored at 280 nm in a spectrophotometer. The gelonin eluted in about fractions 55–70 and was the last major elution peak. Fractions 55–70 were pooled, dialyzed against double distilled water and concentrated by lyophilization. The purity and the molecular weight of each preparation was checked on high pressure liquid chromatography using a TSK 3000 gel permeation column with 50 mM sodium phosphate buffer, pH 7.4 and 15% sodium dodecylsulphate–polyacrylamide gel electrophoresis (SDS–page). Gelonin migrated as a single band with an approximate molecular weight of 29– 30,000 daltons.

EXAMPLE 2

Assay of Gelonin Activity

The gelonin activity was monitored in a cell– free protein synthesis inhibition assay. The cell–free protein synthesis inhibition assay was performed by sequentially adding to 50 µl rabbit

reticulocyte lysate, mixing after each addition, the following components: 0.5 ml of 0.2 M Tris-HCl (pH 7.8), 8.9 ml of ethylene glycol, and 0.25 ml of 1 M HCl).

Twenty microliters of a salt-amino acid- energy mixture (SAEM) consisting of: 0.375 M KCl, 10 mM $\text{Mg}(\text{CH}_3\text{CO}_2)_2$, 15 mM glucose, 0.25–10 mM amino acids (excluding leucine), 5 mM ATP, 1 mM GTP, 50 mM Tris-HCl (pH 7.6), 10 μl Creatinine phosphate-creatinine phosphokinase, 12 μl ^3H leucine (Amersham, 74 mCi/mmol), and adding 1.5 μl of solutions containing varying concentrations of the gelonin mixture. The mixture was incubated for 60 minutes at 30°C. ^3H - leucine incorporation was monitored in an aliquot of the mixture by precipitating synthesized protein on glass fiber filters, washing in 10% TCA and acetone, and monitoring the radioactivity in a Beta-counter using Aquasol scintillation fluid.

Gelonin with a specific activity no lower than 4×10^9 U/mg was used for conjugation with the antibodies. A unit of gelonin activity is the amount of gelonin protein which causes 50% inhibition of incorporation of [^{14}C] leucine into protein in the cell free assay.

EXAMPLE 3

Conjugation of TAb250 With A Preparation of 2-IT Modified Gelonin

Gelonin in phosphate buffered saline was concentrated to approximately 10 mg/ml in a Centriprep 10 concentrator. 5 Triethanolamine hydrochloride (TEA/HCl), pH 8.0, and EDTA were added to a final concentration of 60 mM TEA/HCl and 1 mM EDTA, pH 8.0. A 2-iminothiolane stock solution (500 mM in 60 mM TEA/HCl buffer containing 1 mM EDTA, pH 8.0) was added to a final concentration of 1 mM and the sample was incubated for 90 min at 10 4°C under a stream of nitrogen gas with stirring. Excess iminothiolane was removed by gel filtration on a column of Sephadex G-25 (1 x 24 cm) pre-equilibrated with phosphate-EDTA buffer, pH 15 7.5, containing 0.01 M Na_2HPO_4 , 0.0018 M KH_2PO_4 , 0.0034 M KCl, 0.001 M EDTA and 0.17 M NaCl. Fractions were analyzed for protein content in microtiter plates using Bio-Rad assay. Gelonin eluted at the void volume (about fractions 21-23). These fractions were pooled and stored at 4°C.

EXAMPLE 4

Preparation of Monoclonal Antibodies

BALB/c mice were immunized intraperitoneally (i.p.) and subcutaneously (s.c.) with 2×10^6 – 1×10^7 NIH3T3_T (NIH3T3 cells transfected with c-erbB-2) cells emulsified 1:1 in complete Freund's adjuvant. Animals were boosted every 2 to 4 weeks. When positive titers in an ELISA (described below) were detected, a final i.p. or intravenous (i.v.) boost was given 4 days before fusion. Spleen cells were fused with P3–X63Ag8.653 myeloma cells maintained in RPMI 1640, 10% FBS, and 2 mM L-glutamine. Hybridoma supernatants were tested for positive reactivity in an ELISA (see below), and extracellular domain reactivity was determined by indirect immunofluorescence using unfixed NIH3T3 and NIH3T3_T cells at 4°C followed by flow cytometric analysis. The monoclonal antibody TAb 250 may be obtained from any source. Most preferably, the anti-c-erbB-2 antibody used in the present invention is either a human antibody or a murine antibody.

Hybridoma cells producing TAb 250 are grown in a 2L continuous perfusion bioreactor. The cell supernatant from the bioreactor is filtered and then passed through a Protein-G Trio system followed by ion exchange chromatography. The material is

then concentrated and sterile filtered. Testing of final product includes tests for total DNA, protein purity, pH, (IEF), total protein, endotoxin, potency, identity and protein-G antigen. The present invention, *inter alia*, utilizes a chimeric antibody, including a hybrid antibody or a humanized or human-like antibody. In a preferred embodiment, the variable sequence originates from and is substantially identical to a sequence of the murine TAb 250 antibody. Such a chimeric antibody is BACH- 250.

EXAMPLE 5

ELISA Assay

Sterile 96-well plates were pretreated for 2 hours at 37°C with bovine collagen at 1 mg/ml in sterile PBS. NIH3T3_T (NIH3T3 cells transformed with the vector) cells were grown to 80% confluence and harvested with warm Puck's Versene (0.02% EDTA in PBS), washed, and plated at $0.5-1 \times 10^6$ cells/ml in the treated wells overnight at 37°C. Plates were gently washed and treated with 10% neutral buffered formalin followed by a blocking step with 1% bovine serum albumin BSA/PBS. Sample supernatants or antibody dilutions were then added to the plates and incubated for 2 hours at 37°C followed by incubation with an alkaline phosphatase-conjugated

goat anti-mouse IgG Fc-specific secondary antibody and incubated for 1 hour at 37°C. Plates were washed with PBS, a para-nitrophenyl phosphate and diethanolamine substrate were added and incubated for 15 minutes at room temperature, and A₄₀₅ was measured.

- 5 Supernatants or antibodies that reacted with the transfected cells at an absorbance of 0.2–1.0 greater than the absorbance for a negative control antibody were considered positive.

EXAMPLE 6

Preparation and Handling of ¹²⁵I-TAb 250

10 TAb 250 was radiolabeled using Iodobeads (Pierce) according to the manufacturer's specifications. Carrier-free Na¹²⁵I (400 µCi of IMS. 30, Amersham) was reacted with 25 µg TAb 250 in 100 mM Na-phosphate buffer (200 µl, pH 7.4) in the presence of 3
15 Iodobeads. This resulted in an approximate ratio of one iodine atom per IgG molecule. The incorporation was allowed to proceed at room temperature for 7.5 minutes with intermittent mixing. The reaction mixture was removed from the beads, and after 5 minutes, the volume was adjusted to 0.5 ml with Na-phosphate buffer and 2 µl
20 were taken to estimate specific activity (see below). The remaining volume was desalted by gel filtration using a NAP-5 column

(Pharmacia) equilibrated with PBS containing 0.1% BSA and 0.02% azide. The radiolabeled antibody was eluted in 1 ml column buffer and was stored at 4°C for up to six weeks with no apparent loss of binding activity. The de-salted material was essentially free of
5 unincorporated iodine since >95% was TCA-precipitable.

The specific activity of the radiolabeled antibody was estimated by TCA precipitation of the material before the de-salting step. Thus, 2 µl of the reaction mixture was diluted 500-fold in column buffer and duplicate aliquots mixed with an equal volume of ice-cold 20% TCA. After 25 minutes on ice the precipitated material was collected by centrifugation (10 min, 3000 X g). Supernatants and pellets were counted separately, and the incorporation was expressed as the percent of TCA-precipitable counts. The incorporation obtained in separate iodinations ranged from 27% to
10 45%, yielding specific activity estimates from 3.9 to 7.2 uCi/µg.

Before each binding experiment, an appropriate amount of ^{125}I -TAB
250 was de-salted by gel filtration using a NAP-5 column equilibrated in binding buffer. This procedure removed the azide and yielded material that was routinely >98% TCA-precipitable.

EXAMPLE 7

Cell Culture

The human breast adenocarcinoma cell lines, SKBR-3, MDA-MB-453, and MDA-MB-231, and the human ovarian adenocarcinoma cell line SKOV-3, were used. SKBR-3, MDA-MB-231, MDA-MB-453 cells were maintained in minimal essential medium supplemented with 10% FBS and 2 mM L-glutamine. Medium for MDA-MB-453 cells also contained 1% non-essential amino acid and 1% vitamins. SKOV-3 cells were cultured in Iscove's modified Dulbecco's medium supplemented with 10% FBS and 2 mM L-glutamine. All cultures were incubated at 37°C in either 5 or 10% CO₂ as required.

EXAMPLE 8

Internalization of ¹²⁵I-TAb 250

Internalization of ¹²⁵I-TAb 250 was assessed by determining the amount of radioactivity in acid sensitive and insensitive compartments. Cells were harvested and resuspended in ice cold binding buffer with ¹²⁵I-TAb 250 alone (from 6 ng/ml to 153 ng/ml) or with excess unlabeled TAb 250 to determine non-specific binding. After the cell surface binding of the

radiolabeled antibody reached equilibrium, the cells were pelleted at 200 x g for 5 minutes 4°C and washed three times with ice cold binding buffer to remove unbound antibody. The cell pellets were resuspended in ice cold binding medium, and aliquots were taken to
5 determine the amount of initial ^{125}I -TAb 250 surface binding. To initiate internalization of the radiolabeled antibody, the cells were warmed to 37° C. At times from 15 to 150 minutes, aliquots were removed and the cells were collected by centrifugation (1400 x g 5 minutes, 4°C). The supernatants which contained dissociated or
10 recycled antibody were collected. The pellets were resuspended twice in an acid wash (100 µl/tube PBS, 1% glucose, pH 1). Supernatants containing the surface-bound antibody were combined and counted. The tips of the tubes containing the remaining cell associated radioactivity were clipped and counted.

15 Figure 6 illustrates that monoclonal antibody TAb 250 is not internalized in MDA-MB-231 cells (figure 6B). In contrast, SKBR-3 cells internalized the TAb 250 antibody most efficiently (figure 6A) while internalization of the antibody into SKOV-3 cells and MDA-MB-453 cells was intermediate (figures 6C and 6D,
20 respectively).

EXAMPLE 9

Modification of Monoclonal Antibody TAb 250 With SPDP

N-succinimidyl 3-(2-pyridyldithio)(propionate)(SPDP) in dimethylformamide was prepared as a stock solution of 3 mg/ml in dry dimethylformamide. Since the crystalline SPDP can undergo hydrolysis, the actual concentration of chemically reactive crosslinker was determined by spectrophotometric methods by analyzing the absorbance at 260 nm in a dual-beam spectrophotometer. The concentration of SPDP stock is calculated from the following equation:

$$\frac{\text{Change in absorbance (260 nm)}}{0.02 \times 10^3 \text{ ml mmol}} \times \frac{(301)}{0.01} = \text{mmoles/ml/SPDP}$$

One milligram of monoclonal antibody TAb 250 in 1.0 ml of phosphate buffered saline (PBS) was added to a glass tube. SPDP stock solution was slowly added at about a 5-fold molar excess to the tube (approximately 10 μ l of stock solution), mixing constantly. The mixture was incubated for 30 minutes at room temperature, mixing every 5 minutes during the incubation period.

Excess unreacted SPDP was removed from the sample by gel filtration chromatography on a Sephadex G-25 column (1 x 24 cm) pre-equilibrated with 100 mM sodium phosphate buffer pH 7.0 containing 0.5 mM EDTA (Buffer A). Fractions (0.5 ml) were collected

and analyzed for protein content using the Bradford dye binding assay (Bradford, *Anal. Biochem.* 72: 248–254 (1976). Absorbance (600 nm) was monitored in a 96-well plate using a Bio-TEK Microplate autoreader. Antibody eluted at the void volume (fractions 14–20) and these fractions were pooled and kept at 4°C. The protein was concentrated in a Centricon–30 microconcentrator. The Centricon retentate was washed with 100 mM sodium phosphate buffer, pH 7.0 containing EDTA (0.5 mM). The antibody was concentrated to a final volume of approximately 0.5–0.75 ml.

EXAMPLE 10

Conjugation of SPDP-Modified TAb250 With Iminothiolane-modified Gelonin: Conjugation of 2-IT Modified Gelonin and TAb 250

TAbs 250–gelonin linked with SMPT is prepared by coupling 2-IT-modified gelonin with SMPT-modified monoclonal antibody TAb 250. Briefly, to modify TAb 250 with SMPT, 10 mg of antibody in 1.0 ml of PBS is diluted 1:1 with 2X borate buffer (0.05 M sodium borate 1.7% sodium chloride, pH 9.0) and 52 µl of 4 mM SMPT in dry DMF is slowly added to the antibody solution. The reaction is incubated at room temperature for 2 hr with stirring under N₂. Excess SMPT is removed by passing the reactions mixture

through a Sephadex G-25 column containing phosphate-EDTA buffer, pH 7.5, and antibody positive fractions are evaluated by Bio-Rad assay. The fractions are pooled and stored at 4°C under N₂. The cross-link with 2-IT is carried out at 27°C under N₂ with stirring for 5 96 hr. The final product is purified as described for SPDP in Example 9.

One milligram of purified gelonin (2 mg/ml in PBS) prepared as described in Example 1 was modified with iminothiolane as described in Example 3. Monoclonal antibody TAb250 modified as described in Example 9 was mixed with an equal weight of the modified gelonin. This proportion corresponded to a 5-fold molar excess of gelonin as compared to antibody. The pH of the mixture was adjusted to 7.0 by the addition of 0.05 M TEA/HCl buffer pH 8.0 and the mixture was incubated for 20 hours at 4°C under nitrogen. 10 Iodoacetamide (0.1 M) was added to a final concentration of 2 mM to block any remaining free sulfhydryl groups and incubation was continued for an additional hour at about 25°C. The reaction mixture was stored at 4°C until purification by gel filtration. 15

EXAMPLE 11

Purification of Gelonin–Monoclonal Antibody TAb 250 Complexes

Non-conjugated gelonin and low molecular weight products were removed from the reaction mixtures of Example 10 by gel filtration on a Sephadex S-300 column (1.6 x 31 cm) pre-equilibrated with PBS. Reaction mixtures from Example 10 were concentrated to approximately 1 ml with a Centricon 30 microconcentrator before loading on the Sephadex column. The column was washed with PBS. One ml fractions were collected and 50 μ l aliquots are analyzed for protein by the Bradford assay.

Non-conjugated antibody was removed from the gelonin conjugated antibody by affinity chromatography on a column (1 x 24 cm) of Blue Sepharose CL-6B pre-equilibrated with 10 mM phosphate buffer, pH 7.2 containing 0.1 M NaCl. After loading the S-300 eluate sample, the column was washed with 30 ml of the same buffer to completely elute non-conjugated antibody.

Gelonin-conjugated antibody bound to the column and was eluted with a linear salt gradient of 0.2 to 2 M NaCl in 10 mM phosphate buffer, pH 7.2. The antibody-gelonin complex eluted at approximately 0.7 M NaCl. Protein content of the eluted fractions was determined by the Bradford assay. The protein-containing

fractions were pooled and the elution pattern confirmed by electrophoresis on a 5 to 20% gradient non-reducing polyacrylamide gel. The flow-through peak (fractions 14–20) contains only free antibody while fractions 50–80, eluted with high salt, contain TAb 250–gelonin conjugate free of unconjugated gelonin or antibody. The final product contained TAb 250 antibody coupled to 1, 2 and 3 gelonin molecules. Average gelonin content was 1.5 molecules per antibody molecule. The rabbit reticulocyte *in vitro* translation system was utilized to estimate the gelonin activity of the essentially pure gelonin–TAB 250 antibody complex. One unit of activity in this assay was defined as the amount of protein required to provide 50% inhibition of protein synthesis as compared to untreated controls. Utilizing this assay, the specific activity of both the native gelonin and the TAb 250 gelonin conjugate were determined to be 2×10^8 U/mg and 8.2×10^5 U/mg, respectively. The essentially pure gelonin–TAB 250 antibody is active in the reticulocyte lysate assay. A 1:1000 dilution of the original sample caused approximately a 50% inhibition of protein synthesis, i.e., a 50% reduction of the incorporation of ^{14}C –leucine into protein. Thus, the activity of the original preparation was 1000 U/ml.

The compositions of the present invention may include fusion constructs of the TAb 250 monoclonal antibody and a cytotoxic moiety. Fusion constructs of the immunotoxin of the present invention may be prepared, *e.g.*, by the following method.

5 The nucleotide sequence of both the H and L chain V regions of TAb 250 are easily determined. For example, total RNA is extracted from TAb 250 producing cells with guanidinium thiocyanate. Poly A+ RNA can be isolated by oligo (dT) cellulose chromatography. The appropriate genes can be isolated using standard techniques, including reverse transcription and PCR techniques. A cDNA strand may be synthesized from isolated mRNA using an oligo-dT primer and reverse transcriptase. Such a cDNA strand can be amplified using standard PCR techniques with appropriate primers. A primer near the poly-A tail of the message can be based either upon the poly-A sequence, or upon common adjacent sequences found in mouse immunoglobulins. See Devereaux, Genetics Computer Group, University of Wisconsin Biotechnology Center and its associated sequence databases. A primer at the other end of the gene may be selected from common sequences found in mouse immunoglobulins.

20 See Orlandi *et al.* (1989) *PNAS* 86:3833-3837 and Larrick *et al.* (1989) *Bio/Technology* 7:934-938. The TAb 250 heavy chain gene

has a 5' upstream sequence of ATATAG CAGGAC CATATG (Seq ID No: 2) and starts coding with ATGAA CTTGG GGCTC (Seq ID No: 3). The TAb 250 light chain gene has a 5' upstream sequence TTTAC TTCCT TATTT (Seq ID No: 4) and starts coding with ATGGG CATCA AGATG (Seq ID No: 5). These primers can be used to amplify the genes by PCR technology, and cloned into plasmid expression vectors.

Transfection of DNA into Mouse Cells by Electroporation

Standard transfection methods can be applied to these genes. For example, DNA can be introduced into murine hybridoma Sp2/0-AG14 cells by electroporation. $1-2 \times 10^3$ actively growing SP2/0-AG14 cells are washed and resuspended in 1.0 ml of sterile PBS. Thirty micrograms of each chimeric, IgK and IgG1, plasmid is added to the cell suspension. The DNA/cells are transferred to a precooled shocking cuvette, incubated on ice at least 5 minutes and then a 0.5 kv/cm electro-pulse is delivered for 10 msec (Transfector 300, BTX). After shocking, the DNA/cell mixture is returned to ice for 10 minutes, diluted in 10 ml of DMEM containing 5% NCTC-109 and 10% FCS, and incubated at room temperature for 10 minutes. Finally, the cells are transferred to a 37°C incubator with 7% CO₂ for 48 hours before plating in selective medium, containing 1 µg/ml Xanthine. Cells can be plated in 96-well plates at 3×10^4 cells/well

and the culture supernatants assayed by ELISA for antibody bound to TAb 250 antigen positive target cells.

EXAMPLE 12

5 MTT Assay

3-(4,5-dimethyl/thiazolyl)-2,5-diphenyltetrazolium bromide (MTT) assays were carried out by removing cells from tissue culture flasks with versene 1:5000, centrifuging at 500 x g for five minutes, and resuspending the cells in medium at a concentration of 1×10^5 cell/ml. Cells were plated at 100 μ l/well into 96-well microtiter plates and incubated in a humidified CO₂ incubator at 37°C for 24 hours.

On the next day, TAb 250 or TAb-250 gelonin was added. Immediately after deposition of the highest antibody concentration into the first column of wells, 1:2 dilutions of antibody were performed directly in the microtiter plates using a multichannel pipette. Plates were then incubated for three days, followed by the addition of 10 μ l/well of MTT. MTT was prepared as a 5 mg/ml solution of PBS, filtered sterilized, and stored at 4°C in the dark.

Plates were kept dark and incubated for an additional 4 hours at 37°

C. The MTT crystals were dissolved by mixing the contents of the wells vigorously with 100 microliters of isopropanol containing 0.04 N HCl and 3% sodium dodecylsulfate. Absorbance at 570 nm was determined using an enzyme-linked immunosorbent assay reader.

5

EXAMPLE 13

CPA

The cell proliferation assay (CPA) measures cell growth by determination of cell number and viability. On Day 0, cells at 80–90% confluency are released from a tissue culture flask, pelleted at 200 x g at 20°C for 6 minutes, and resuspended in Iscove's MEM containing 2 mM glutamine and 10% fetal bovine serum to a concentration of 6000 cells/ml. The cell suspension is added to 24–well plates at 1 ml per well. The plates are incubated at 37°C, 5% CO₂, for 24 hours. On day 1, the cells in three wells are washed with PBS and released from the plate with 1 ml 0.05% trypsin in PBS and 500 µl are counted using a Coulter Counter. To the remaining wells are added 20 µl of PBS, TAb 250, or TAb 250–gelonin, at the concentrations indicated in the figures. The plates are returned to the incubator. At the time–points indicated in the figures, the cells are released with trypsin and counted as on Day 1. The remaining

500 μ l of cells are stained with propidium iodide so that viability can be determined by flow cytometry. For each point on the graph, a mean cell number (n=3) is determined and multiplied by the percent of live cells to find the number of viable cells. This is divided by the number of live cells treated with PBS to the percent of control on the Y axis.

EXAMPLE 14

Cytotoxicity of Gelonin and Gelonin-TAb 250 Antibody Complex

As can be seen in Figure 1, ZME antibody had virtually no effect on SKOV-3 cells. In contrast, TAb 250-gelonin immunoconjugate was highly active. Figure 2 illustrates a cytotoxicity of the TAb 250-gelonin immunoconjugate on SKOV-3 cells compared to gelonin alone. At the same concentration, TAb 250-gelonin immunoconjugate was approximately 10,000x as cytotoxic as gelonin alone. Figure 3 demonstrates that an irrelevant antibody, ZME 18 monoclonal antibody, has no competitive effect on the cytotoxicity of the TAb 250-gelonin immunoconjugate. In contrast, increasing concentration of TAb 250 monoclonal antibody decreases the cytotoxicity of the immunoconjugate in a dose dependent fashion. Figure 4 illustrates a dose response relationship

of the TAb 250–gelonin immunoconjugate on SKOV–3 cells. As seen in the CPA assay, a dose above 0.1 micrograms/ml produced 80% inhibition at six days. Figure 5 illustrates the effects of either the monoclonal antibody TAb 250 alone or the conjugate of TAb 250 with gelonin on SKOV–3 cells. As can be seen, there is a dose dependent inhibition by the TAb 250–gelonin immunoconjugate in the CPA assay. Figure 7 depicts the effects of the TAb 250– gelonin immunoconjugate on four different cell lines. As would be expected, the greatest amount of toxicity occurred in the SKBR–3 cell line. Intermediate toxicity was evidenced in SKOV–3 and MDA–MB–453 cells while virtually no cytotoxicity was seen in MDA–MB–231 cells. Thus, the cytotoxicity of the TAb 250–gelonin immunoconjugate correlates with the number of cell surface receptors in these cells.

EXAMPLE 15

Single Chain Immunotoxins

Single-chain antibodies (scFvs or sFvs), incorporating the binding characteristics of the parent immunoglobulin, consist of the antibody V_L and V_H domains (the Fv fragment) linked by a designed flexible peptide tether. The translation of scFvs as single polypeptides ensures expression of both V_L and V_H chains in

equimolar concentrations and the covalent linking of the two sequences facilitates their association after folding. Compared to intact IgGs or Fab fragments, scFvs have the advantages of smaller size and structural simplicity with comparable antigen-binding 5 affinities. In addition, they are more stable than the analogous two-chain Fv fragments. Furthermore, scFvs have significant advantages in clinical and diagnostic applications currently involving conventional monoclonal antibodies or Fab fragments thereof. The smaller size of scFvs would provide for better penetration of tumor tissue, improved pharmacokinetics, and a reduction in the immunogenicity and high backgrounds observed with intravenously administered Fabs.

Figure 8 shows a schematic of the construction of the scFv23-gelolin immunotoxin gene. A single-chain analogue of the antibody e-23 was raised against an epitope on the 185 kD antigen p185HER-2/neu found on the surface of breast and ovarian carcinomas. The 12 amino acid 212 linker was chosen to tether the two variable regions of the antibody since this sequence was shown to provide for proteolytic stability and functional antibody in several 10 instances. Alternately other linker sequences such as the flexible Gly-rich peptide, linking peptides from multidomain proteins, or 15 20

other designed peptides e.g. the 202, 202', 205, and 218 could have been selected. In addition a functional linker could have been selected from a randomized sequence library using phage display technology or a colony filter-lift hapten-binding assay.

5 Furthermore, short linker sequences used in the construction of diabodies could also have been chosen. Antibodies recognizing tumor cell-surface epitopes have the ability to selectively localize within human tumors after systemic administration and therefore can serve as targeting probes for the site-specific delivery of cytotoxic
10 chemotherapeutic agents such as Pseudomonas exotoxin, ricin or gelonin. An immunotoxin was constructed with sFv-23 and gelonin. In addition, with a view to increased efficacy of the immunotoxin, the carboxyl-terminal endoplasmic reticulum retrieval signal Lys-Asp-Glu-Leu was added to the sequence of gelonin. A person having
15 ordinary skill in this art would readily recognize that certain modifications in the sequence of scFv-23 could be made, e.g., a V_H - Linker - V_L format and CDR grafting to construct a humanized or chimeric antibody to minimize potential immunogenicity problems with this murine antibody.

EXAMPLE 16

Construction of the gene encoding the scFv-23-Gelonin immunotoxin

Figure 9 shows the configuration and sequence of the scFv-gelonin immunotoxin. The genes encoding scFv-23 and gelonin (Gel) were linked together in a 5' Nco I-and 3' Hind III-flanked scFv-gelonin orientation via a short Gly-Gly-Gly-Gly-Ser (Seq ID No. 1) peptide tether to provide flexibility between the two proteins in a PCR-based method. Briefly, DNA encoding sFv-23 was amplified using the primer 1: 5'-GCTGCCCAACCAGCCATGGCGATGTCTGACGTC-3' (Seq ID No: 6) and primer 3: 5'-CCGGAGCCACCGCCACCGCTAGCTGAGGAGACTGTGA-3' (Seq ID No: 7). Simultaneously, DNA encoding gelonin from the vector pRCM1808B was amplified using the following primers: primer 2: 5'-GGTGGCGGTGGCTCCGGTCTAGATACCGTTAGC-3' (Seq ID No: 8) and primer 4: 5'-CGGCCGCAAGCTTAAGTACAGCTCGTCTTTCTCGAGGAATTCAGCAG-3' (Seq ID No: 9)

PCR syntheses were carried out in a Perkin Elmer Thermal Cycler and the profile used for the construction of the complete immunotoxin gene was as follows: the first step involved

25 cycles of 94°C denaturation for 1.5 minutes, 50°C annealing for 1.5 minutes, and a 1 minute extension at 72°C followed by a single 72°C incubation for 5 minutes using all four aforementioned primers. Following this, one tenth volume of the crude PCR product was removed and added to a second PCR mixture containing only primers 1 and 4. This second PCR synthesis also comprised 25 cycles each with a profile identical to that of the previous amplification. This 1,500 bp product was gel purified using GeneClean II (Bio 101), digested overnight at room temperature with Nco I and Hind III, and cloned into the Novagen vector pET-22b (+).

Expression of the scFv-23-Gelonin immunotoxin in *E. coli* pET-22b clones encoding full-length immunotoxin, as judged by restriction digest analysis, were transformed into competent *E. coli* BL21(DE3) pLysS and incubated in 2xYT growth medium (16 g/L tryptone, 10 g/L yeast extract, 5 g/L NaCl; pH 7.0) at 37°C until the A600 of the cultures was 0.4. IPTG (Isopropyl-β-D-thiogalactopyranoside) was added to a final concentration of 1 mM and induction was continued overnight at 16°C. The periplasmic fractions of the harvested bacteria were isolated using osmotic shock and mild sonication and supernatants were used directly in ELISA and Western analyses. ELISA and Western analyses Wells of a 96-

well microtiter plate were coated overnight with antibody 13A3, an anti-gelonin murine monoclonal antibody and then blocked with BSA. In subsequent steps periplasmic lysates containing immunotoxin clones, rabbit anti-gelonin polyclonal antibody, and finally horseradish peroxidase (HRPO)-conjugated goat anti-rabbit IgG were added. The plate was developed with the HRPO substrate 2,2'-azino-bis-(3-ethylbenzthiazoline-6-sulfonic acid (ABTS) and the signal quantitated at 405 nm. For Western blots, periplasmic lysates were separated by 11% SDS-PAGE and transferred onto nitrocellulose. The filters were blocked in 5% BSA and then reacted with 0.1 ug/ml of rabbit anti-gelonin (or rabbit anti-scFv-23) polyclonal antibody in 1% BSA. After extensively washing the filters in Tris-buffered saline, pH 7.4 (TBS)-0.5% Tween 20, 0.1 ug/ml horseradish peroxidase-conjugated goat anti-rabbit IgG was added. Following this incubation, the filters were again washed in TBS-0.5% Tween 20 as before and developed with the Amersham ECL detection system.

EXAMPLE 17

sFv23/Gelonin cloning as a glutathione-S-transferase fusion protein

DNA encoding the sFv-23-Gelonin immunotoxin from pET-22b was digested with Nco I, blunt-ended with DNA Pol I

Klenow and dNTPs, and then purified using the PCR purification kit from Qiagen. The DNA was then digested with Hind III, gel purified using GeneClean II (Bio 101) and cloned into the Sma I and Hind III sites of the GST fusion vector pGEX-2T (Pharmacia). By virtue of this cloning method three extra N-terminal amino acids (Pro-Met-Ala) were added onto the antibody fragment of the immunotoxin.

EXAMPLE 18

Expression of sFv-23-Gelolin as a GST fusion protein in *E. coli*

Positive DNA clones encoding the GST-immunotoxin fusion were transformed into the *E. coli* strains JM109, XL1-Blue MRF', E104, and BL21 and induced under a variety of conditions. The optimal conditions were determined to be as follows: transformed cells were grown in 2xYT at 37°C to an A600 of 0.7-0.9 when IPTG was added to a final concentration of 0.2 mM; cultures were then induced at 37°C for 4 hours. Cells were centrifuged, washed with Tris-buffered saline (TBS) (pH 8) and frozen at -80°C until use. Once thawed, the bacterial pellet was resuspended in TBS, 2 mg/mL lysozyme and 1 mM phenylmethylsulfonyl fluoride (PMSF) and sonicated. Supernatants were saved and bound to Glutathione-agarose for 30 minutes at room temperature. The resin was washed

successively with an excess of 1M NaCl in TBS (pH 8) and TBS (pH 8). Fusion protein bound to the resin was then either eluted off the resin with 15 mM glutathione, reduced form (GSH) or digested directly with thrombin protease (Novagen) overnight at room temperature using standardized conditions.

EXAMPLE 19

Construction and expression of scFvZME-018-Gelonin Immunotoxins

Figure 10 shows a binding analysis of sixteen individual scFv-gelonin immunotoxin clones. Gelonin was fused to the C-terminal end of scFv-23 via a short, nonstructured Gly4Ser linker to provide a measure of flexibility between the two protein moities. The sequence chosen was one of several possible peptides that could have been designed based on small size and hydrophilicity to minimize steric crowding and aqueous solubility, respectively. The products of the first set of PCR cycles are the individual scFv antibody and toxin fragments whereas the predominant product from the second set of reactions is the full-length immunotoxin. For possible increased cytotoxicity, the endoplasmic reticulum retrieval signal KDEL was added to the C-terminal sequence of gelonin to provide for more efficient transport of the toxin to the ER from

where translocation into the cytosol occurs. The final PCR product was gel purified, digested with Nco I and Hind III, and cloned into the T7-expression vector pET-22b. Bacterial clones containing full-length immunotoxin DNA were induced with IPTG and both culture supernatants and periplasmic extracts screened by ELISA for binding to both antibody-specific hapten p185HER-2/neu on the surface of SK-OV-3 or BT474 cells and 13A3, an anti-gelonin murine monoclonal antibody. Functional immunotoxin bound to target was detected with a polyclonal rabbit anti-gelonin antibody followed by a horseradish peroxidase-conjugated goat anti-rabbit IgG antibody. Several immunotoxin clones with the highest binding titers to both BT474 cells and antibody 13A3 were chosen for sequencing and further characterization.

Different bacterial hosts and induction conditions were examined to improve expression yields and reduce the degradation problems initially observed with the immunotoxins. These manipulations were largely successful on a small scale (5 ml) but any attempts to scale up these growth conditions were largely unsuccessful. Using a combination of ion-exchange, hydroxyapatite, and affinity chromatography, yields and purity of protein obtained were unviable (data not shown) and the GST-fusion system was

selected as a possible means to improve expression yields of the immunotoxin.

EXAMPLE 20

5 Expression of scFv-23-gelonin immunotoxins as a GST-fusion

Figure 11 shows a western blot analysis of the scFvZME-gelonin and scFv-23-gelonin proteins refolded and purified from insoluble inclusion bodies. The pGEX-2T vector was used to express the scFv-23-gelonin immunotoxin as a GST fusion. Proteins expresses as GST fusions are generally purified in high yields using non-denaturing conditions in a one-step procedure with glutathione-agarose (GSH-ag) affinity chromatography. The vector has been designed so that the GST carrier can be cleaved from the target fusion protein by virtue of a thrombin cleavage site between the two protein moieties. Furthermore, any contaminating GST or undigested fusion protein can be removed by rebinding to GSH-ag. The incompatible 5' restriction sites in the vector necessitated blunt-ending the 5' end of the immunotoxin and the consequent addition of three extra amino acids (Pro-Met-Ala) to the N-terminal of scFv-23. However, this would not be expected to adversely affect its binding activity since this end of an antibody is far removed from the

binding site. Different induction conditions were used to optimize expression of soluble fusion protein.

EXAMPLE 21

5 Immunoconjugates with BACH-250

Recombinant gelonin (rGel) was covalently linked to BACH-250 with the heterobifunctional cross-linking reagent SPDP by the same method used in Example 9 to crosslink recombinant gelonin to TAB-250. Briefly, a stock solution of SPDP in dry DMF was added to a solution of either TAB-250 or BACH-250 to a final concentration of 5-fold molar excess. Excess unreacted SPDP was removed by Sephadex G-25 chromatography. SPDP-derivative antibody fractions were pooled and kept at 4°C . One milligram of purified rGel (2 mg/ml in PBS) was added to triethanolamine hydrochloride (TEA/HCl) buffer to a final concentration of 60 mM TEA/HCl, adjusted to pH 8.0, and then EDTA was added to a concentration of 1 mM. A 2-iminothiolane stock solution was added to a final concentration of 1 mM, and the sample was incubated for 90 min at 4°C under nitrogen. Excess 2-iminothiolane was removed by gel filtration. SPDP-modified antibody was mixed with an equal weight of 2-iminothiolane-modified rGel, which corresponded with a 5-fold molar

excess of gelonin as compared to antibody. The pH of the mixture was adjusted to 7.0 by adding 0.5 M TEA/HCl buffer (pH 8.0) and the mixture was incubated for 20 hr at 4°C under nitrogen. Iodoacetamide (0.1 M in H₂O) was added to a final concentration of 2 mM to block any remaining free sulfhydryl groups, and incubation was continued for an additional hour at 25°C.

To remove low molecular weight products and nonconjugated rGel, the reaction mixture was applied to a Sephacryl S-300 column (1.6 x 31 cm) previously equilibrated with PBS. Fractions were collected and the protein content measured. The high molecular weight peak fractions were applied to an affinity chromatography column of blue sepharose CL-6B (1 x 24 cm) pre-equilibrated with 10 mM PBS (pH 7.2) containing 0.1 M sodium chloride. The column then was washed with 50 ml of buffer to completely elute the non-conjugated antibody. This was done with a linear salt gradient of 0.1 to 2 M sodium chloride in 10 mM PBS (pH 7.2). The protein content of the eluted fractions was determined using the dye-binding assay described previously (30), and the samples were analyzed using non-reducing SDS-PAGE.

Analysis of the final purified sample using nonreducing SDS-PAGE showed that the final product contained a mixture of

immunotoxins containing one rGel molecule (major) and immunotoxins containing two rGel molecules (minor). No demonstrable amounts of unconjugated antibody or free gelonin toxin were detected.

5

EXAMPLE 22

Cell Binding and Competition Studies

The binding of either unmodified TAb-250 or TAb-250/rGel to antigen-positive SKOV-3 cells was assessed by an enzyme-linked immunosorbent assay. Both reagents had a similar profile, suggesting that the binding determinants of the antibody are preserved after SPDP modification and toxin conjugation.

To examine the displacement of ^{125}I -labeled BACH250 and ^{125}I -BACH250/rGel binding, SKOV-3 cells were grown to 80-90% confluence and harvested. A total of 3.6×10^4 cells were incubated on ice for 4 hr in the presence of 5-10 ng/ml of ^{123}I -labeled BACH-250 or BACH-250/rGel alone, or in combination with increasing amounts of unlabeled mouse IgG, BACH-250 or BACH-250/rGel for 4 hr. Cells then were washed and cell-associated radioactivity was determined using a gamma counter. Percentage of cells bound was determined by dividing the counts per minute,

(cpm) measured by total cpm and then multiplying the quotient by 100.

In the competitive displacement studies, BACH-250 and BACH-250/rGel competed or cross-competed with either reagent as shown in Figure 13. Both radiolabeled BACH-250 and BACH-250/rGel bound SKOV-3 target cells to the same extent, and the binding of neither agent could be displaced by nonspecific IgG. On the other hand, with the addition of unlabeled free antibody or immunotoxin, the binding of either radiolabel competed identically. The affinity of the immunotoxin appeared to be identical to that of the original antibody and the binding of the immunotoxin to target cells occurred solely through the interaction of the antibody component of the immunotoxin with the antigen on the target cells.

EXAMPLE 23

In Vitro Cytotoxicity Studies in SKOV-3 Cells

Various concentrations of BACH-250/rGel were applied to log-phase SKOV-3 cells in culture in the presence of increasing concentrations of free antibody. For the assays with immunotoxins, cultures were washed, and cells were detached using versene (edetate sodium), after which the cells were resuspended in complete

medium at a density of 5×10^4 cells/ml. Aliquots of suspended cells were then dispensed into 96-well microtiter plates and the cells allowed to adhere for 24 hours. The medium was then replaced with medium containing different concentrations of immunotoxin or rGel, and the cells were incubated for an additional 72 hours. The relative cell proliferation was analyzed using the MTT staining previously described (31). The values given are the means of duplicate experiments performed in quadruplicate.

As shown in Figure 14, the immunotoxin was almost 100% cytotoxic to target cells at doses of up to 1×10^{-10} M. The IC_{50} for the immunotoxin was 54.7 pM. In contrast, the IC_{50} for free rGel under these conditions was 2×10^{-6} M, or a concentration almost 4 logs higher than that of the immunotoxin (data not shown). The co-administration of free antibody at doses of 0.01, 0.1, and 1.0 ug/ml increased the immunotoxin IC_{50} to 63.5, 206, and 639 pM respectively (see Figure 13). Cell culture studies with the BACH-250/rGel immunotoxin showed significant cytotoxicity occurred at concentrations of more than 0.01 ug/ml (Figure 15). Maximal effects on cells were observed by day 6 of culture and growth inhibition was observed for the 10 days of the observation period.

EXAMPLE 24

Internalization studies.

The internalization of ^{125}I -TAb-250 or ^{125}I -TAb-250/rGel was assessed by determining the amount of radioactivity in acid-sensitive and insensitive compartments. Cells were harvested and resuspended in buffer containing radiolabeled antibody alone or containing excess unlabeled antibody to determine the extent of nonspecific binding. After the cell-surface binding of the radiolabeled antibody reached equilibrium, the cells were centrifuged and the pellet was washed to remove unbound antibody. The cells were warmed to 37°C to initiate internalization of the radiolabeled antibody. At the times indicated, aliquots were removed and the cells collected by centrifugation. The supernatants that contained dissociated or recycled antibody were collected. The pellets were resuspended twice in an acid wash, and the supernatants containing the surface-bound antibody were combined and counted. The tips of the centrifuge tubes containing the remaining cell-associated radioactivity were then clipped and counted.

Studies performed with various cell lines expressing different levels of *c-erb-2/HER-2/neu* showed that antibodies TAb-

250 and BACH-250 were not internalized in MDA-231 cells, which contain low levels of the c-erbB-2 protein ($\sim 0.16 \times 10^6$ sites/cell). In contrast, SKBR-3 cells ($>2 \times 10^6$ sites/cell) internalized the antibody most efficiently, and the internalization of antibody into SKOV-3 cells ($\sim 1 \times 10^6$ sites/cell) and MDA-453 cells ($\sim 0.35 \times 10^6$ sites/cell) cells was intermediate (data not shown).

EXAMPLE 25

In Vitro Cytotoxicity Studies in Six Different Cell Lines.

The cytotoxic effects of TAb-250/rGel were examined in six different cell lines that express various levels of the c-erbB-2 protein (see Figure 16). As predicted from the internalization studies, the cytotoxic activity of the conjugate was greatest against the SKBR-3 cell line with the highest number ($\sim 4 \times 10^6$ sites/cell) of cell-surface receptors. Intermediate toxicity was observed in SKOV-3 cells, further reduced cytotoxic effects were observed in MDA-MB-453 cells, and almost no cytotoxic effects were observed in MDA-231 or MCF-7 cells, which had the fewest number of sites/cell.

This experiment showed that in these cells, the cytotoxicity of the TAb-250/rGel immunoconjugate correlates well

with the number of cell-surface receptors and that the internalization of cell-surface bound antibodies or immunotoxins may depend on the number of available c-erbB-2 receptors. Because rGel must be internalized to be cytotoxic, anti-HER-2 immunotoxins
5 may be most effective in cells expressing very high levels of HER-2.

EXAMPLE 26

Pharmacokinetics of Immunoconjugate Clearance from Plasma

4-6-week old balb/c mice were injected with 0.3 μ Ci (5
10 μ g) of either labeled monoclonal antibody (mAb) or immunoconjugate. Three mice each were sacrificed by cervical dislocation at 15, 30, 45, 60, 75, 90, 105, 120, 240 min and 24 hours after injection. Blood samples were removed (chest cavity), and
15 weighed and radioactivity was determined with a gamma counter. The blood samples then were centrifuged, and supernatant was decanted and counted to determine the amount of the plasma-associated radiolabel. The amounts of radioactivity were analyzed
20 by a least-squares nonlinear regression program (RSTRIP, MicroMath, Inc.) to determine the pharmacokinetic parameters. The pattern of the clearance of radiolabeled BACH-250 and BACH-

250/rGel from the plasma of mice after intravenous administration is shown in Figure 17.

Although both agents were cleared from plasma in a similar fashion, pharmacokinetic analysis of the clearance curves (Table I) showed that the BACH-250 antibody closely fit a one-compartment model for clearance, whereas the immunotoxin was cleared biphasically. In addition, the calculated terminal-phase half-lives of the antibody and the immunotoxin were 26.9 and 75.2 hours respectively. However, the initial concentrations of the two agents in plasma were similar, suggesting that there were no significant changes in the initial apparent volume of distribution. Also, the overall clearance kinetics of the immunotoxin were not significantly influenced by the toxin component, but do appear to be related to the rate of clearance of the parent antibody.

TABLE I

**COMPARISON OF BACH-250 and BACH-250/rGel
PHARMACOKINETICS**

AGENT	Cpo (% I.D./ g)	T _{1/2} AL PHA (h)	T _{1/2} BET A (h)	Cxt (%I.D./ g x h)	RT (h)
BACH-250	15.24	----	26.9	592	38.8
BACH- 250/rGel	13.8	4.6	75.2	492	94.9

Cpo = Concentration in plasma extrapolated to t=0. Calculated from least squares, best-fit equation.

T_{1/2}ALPHA = Calculated alpha phase half-life in plasma.

T_{1/2}BETA = Calculated beta-phase half life in plasma.

Cxt = Calculated area under the concentration curve.

RT = Residence time

EXAMPLE 27

Tissue Distribution of BACH-250 Derived Immunotoxin

Balb/c athymic mice received subcutaneous implants of SKOV-3 (positive for c-erbB-2 expression) and MDA-MB-468 (negative for c-erbB-2 expression) tumor cells on the left and right hind flanks, respectively. Tumors were allowed to grow to 0.1-0.4 g,

at which time 10 animals each were injected intraperitoneally with approximately 10 μ g of either 125 I-TAb-250 or 125 I-TAb-250-gelonin (2-8 uCi/ μ g). Both of the iodinated proteins have been shown to compete comparably for the binding of unlabeled TAb-250 on SKOV-3 cells. Two animals from each group were then sacrificed at 6, 18, 24, 30 and 48 h; tumor and normal tissues were removed, and the samples were weighed and radioactivity counted. The amount of radiolabeled antibody distributed to the tumor or various organs was expressed as the percentage of the injected dose per gram of tissue, and finally expressed as a ratio to the concurrent blood levels.

The tissue distribution profiles of radiolabeled BACH-250 and BACH-250/rGel at 24, 48, 72, and 96 hours. after intravenous administration in athymic mice bearing SKOV-3 xenografts are shown in Figures 18-21 respectively. In almost all the tissues studied, the content of radiolabeled immunotoxin generally paralleled that of the parent antibody at all the same points. However, at 24 and 48 hours. after administration, the content of immunotoxin in liver and spleen appeared to be higher than that of the parent antibody, indicating that the increased uptake may be due to the effects of toxin component of the immunotoxin construct. The

content of both the antibody and the immunotoxin in normal tissues decreased significantly over time. In addition, as shown in Figure 22, the content of the immunotoxin and the parent antibody were similar over time, reaching a plateau by approximately 48 hours.

5 after administration and remaining for 96 hours.

EXAMPLE 28

BACH-250/rGel Treatment of Subcutaneous Tumors in balb/c Mice

Results of treatment of balb/c mice bearing well-established subcutaneous SKOV-3 tumors are shown in Figure 23. To examine the effects of TAb-250 or TAb-250/rGel on subcutaneous tumor growth we implanted SKOV-03 tumor cells subcutaneously in balb/c athymic mice (7-8 mice/group). Test agents were administered intraperitoneally three times per week for three weeks, starting on day 5 after implantation (tumor volumes, 0-30 mm³). Mice received PBS (control), TAb-250 (42 µg/dose) plus rGel (8 µg/dose), or the TAb-250-rGel conjugate (50 µg/dose). Tumors were measured twice per week with vernier calipers and volumes (in cubic millimeters) were calculated as a product of the length x width x height.

Treatment with the TAb-250-rGel conjugate was shown to inhibit tumor growth by 99% at day 35 and up to 93% by day 55. In contrast, animals treated with the unconjugated combination of TAb-250 plus free rGel showed a maximal tumor inhibition of only 56% compared with the control group over days 35-55. Tumors regressed or did not develop in 5 of the 7 animals in the immunotoxin (TAAb-25/rGel) treatment group. Whereas sizable tumors developed in all 7 animals treated with either PBS or the unconjugated combination of TAb-250 plus rGel. This finding showed that tumor growth was inhibited significantly more in animals treated with the TAb-250/rGel conjugate than in control mice, (99% and 94% at day 49 after implantation, respectively); and also more so than it was in mice treated with equimolar amounts of unconjugated TAb-250 plus free rGel (4% at day 35, 56% at day 49).

EXAMPLE 29

BACH-250/rGel Treatment of Intraperitoneal Tumors in balb/c Mice

To examine the effects of TAb-250 or TAb-250-rGel on intraperitoneal tumor growth, we implanted intraperitoneal SKOV-3 tumor cells in 60 balb/c athymic mice. After implantation, the mice were randomly divided into five treatment groups of 12 mice each.

Beginning on day 7, the mice in each group were given an intraperitoneal injection of PBS, 107 μ g of TAb-250 alone, 20.5 μ g of rGel alone, 107 μ g TAb-250 plus 20.5 μ g of rGel, or an equimolar amount of the TAb-250/rGel conjugate. A total of nine treatments were given, three times/wk for 3 weeks, and survival time for each mouse was recorded.

In this lethal model of SKOV-3 growing as an intraperitoneal xenograft, treatment of balb/c mice with the TAb-250/rGel conjugate lengthened the survival of mice with these tumors by an average of 40% (12.4 days) as compared with survival in mice treated with PBS, TAb-250 or unconjugated TAb-250 plus rGel (Figure 13).

EXAMPLE 30

scFv-23-TNF (tumor necrosis factor) immunotoxins.

Previous studies have demonstrated that chemical conjugates of human tumor necrosis factor (TNF) and monoclonal antibodies display significant targeted cytotoxic properties against tumor cells in culture which appears to be far superior to those of native TNF (s1-39-s1-41). see Rosenblum, et al., *Cancer Communications*, 3(1): 21-27, (1991); Rosenblum, et al.,

Immunotherapy, 40(5): 322-328, (1995); and, Rosenblum, et al.,
American Association for Cancer Research, Abstract 8802, (1989). In
addition, studies in xenograft models suggest that these
immunocytokines readily accumulate specifically in tumor tissues
5 and demonstrate superior *in vivo* antitumor activity compared to
native TNF. Therefore, the decision was made to construct a second
generation molecular construct of a recombinant single chain
antibody fused to the TNF molecule thereby incorporating both
antibody and TNF functions within the same molecule.

Therefore, a recombinant fusion toxin was constructed
composed of a scFv-23 single-chain antibody targeting the HER2/*neu*
protooncogene and human TNF as a cytotoxic effector molecule. The
cDNA encoding mature human TNF was from Dr. James Klostergaard,
M. D. Anderson (Houston, TX). The sFv23/TNF cDNA was constructed
15 by a two-step PCR reaction. The first step consisted of separate PCR
amplification of the antibody and TNF coding sequences utilizing
forward and reverse primers for each sequence. The final step
consisted of PCR reaction of the sequences utilizing overlap primers
additionally incorporating a flexible tether (G₄S) between the
20 antibody and TNF as shown in Figure 25. A flexible 14 amino acid
connected the V_H and V_L regions of the antibody as shown in Fig 25.

EXAMPLE 31

Bacterial expression of the scFv-23-TNF immunotoxin.

The PCR reaction products were cloned into a vector for bacterial expression of the recombinant insert. The complete insert
5 was submitted for dideoxynucleotide sequencing (M. D. Anderson Core Sequencing Facility) and the final gene product sequence was confirmed.

Bacterial colonies transfected with the plasmid carrying the sFv23/TNF insert were agitated in a bacterial shaker (Innova
10 4330, New Brunswick Scientific) in 1 L of TYE media (15 g tryptone, 10 g yeast extract, 5 g NaCl) containing 50 µg/ml tetracycline (Sigma) overnight at 37°C at 250 rpm. The bacterial cells were harvested by centrifugation and the pellet dispersed into 2 L of fresh TYE. Expression of the target protein was induced by the addition of
15 arabinose to a final concentration of 0.1%. The culture was further incubated at 37°C for 2-3 hours. The cells were collected by centrifugation, resuspended in 80 ml extraction buffer (50 mM Tris/HCl, pH 8, 20 mM EDTA, 0.25 mg/ml lysozyme, (Sigma) and incubated with shaking for 1 hour at room temperature. Triton X-
20 100 and sodium chloride were added to the sample at a final concentration of 2% and 0.5 M respectively, then incubated for an

additional 0.5 hours. After centrifugation, the insoluble fraction of the inclusion bodies was resuspended in 160 ml of 50 mM Tris/HCl, pH 8.0, 20 mM EDTA and sonicated (6 x 20 sec) using a Vir Sonic 300 sonicator (Virtis, Gardiner, NY). The inclusion bodies were then harvested by centrifugation, washed three times in the same buffer and stored at -80°C. Bacterial expression of the scFv23/TNF fusion construct is shown in Figure 25. After growth and induction with arabinose at 37°C, production of the construct was approximately 5-10% of total protein as assessed by SDS-PAGE.

EXAMPLE 32

Western blot analysis of scFv-23-TNF immunotoxin expression.

Western blot analysis of scFv23/TNF expression was performed on protein samples from the crude extracts of *E. coli* and the purified inclusion bodies. sFv23 and r-TNF were analyzed by SDS PAGE under reducing conditions. The gel was electrophoretically transferred overnight onto a nitrocellulose transfer and immobilization membrane (Protran, VWR, Sugar Land, TX). The membrane was then incubated in 5% BSA/TBS (20 mM Tris/HCl, 137 mM NaCl, and 0.5% Tween 20, pH 7.6) and then incubated for 1 hour with anti-sFv23 rabbit polyclonal antibody (1:10,000 dilution in

TBS/Tween). After successive washing with TBS/Tween 20, the membrane was incubated with goat-anti-rabbit IgG horseradish peroxidase (1:5000 dilution in TBS). Then, the membrane was developed using ECL reagents (Amersham Corp., Arlington Heights, IL) and exposed to X-ray film (Kodak). Estimation of production of the target fusion construct was 25-50 mg/L as assessed by Western blot analysis.

EXAMPLE 33

Purification of scFv-23-TNF immunotoxin.

Insoluble inclusion bodies were denatured by addition of 6 M guanidine, 100 mM Tris/HCl, pH 7.5, 2 mM EDTA, and 50 mM DTE to make a final concentration of 3 mg/ml protein (as assessed by Bradford protein determination). After a minimum of 2 hours agitation at room temp, the solubilized proteins were diluted 100-fold in refolding buffer (100 mM Tris/pH 7.5, 2 mM EDTA) and incubated at 12⁰C for 48 hours. The protein was subsequently bound to a small column containing SP Sepharose Fast Flow Resin (Pharmacia). The bound fraction was eluted from the column by addition of 2 M NaCl in 100 mM Tris/pH 7.5, 2 mM EDTA. Eluted protein fractions were pooled and dialyzed against TBS.

Purification of the soluble protein utilizing ion exchange chromatography resulted in essentially homogeneous material as assessed by SDS-PAGE after elution from the exchange resin. Yield of final, purified material was approximately 100 µg/L of bacterial culture. Western analysis of the product utilizing antibodies to either TNF or to scFv23 (Figure 27) demonstrated an immunoreactive species with both antibodies at the expected molecular weight (43 kDa).

EXAMPLE 34

scFv-23-TNF immunotoxin binding studies.

The binding of the sFv23/TNF fusion toxin to adherent SKBR3 cells was assessed by ELISA using an anti-TNF antibody. Log-phase SKBR-3 cells were washed in PBS and 50,000 cells/50 µl PBS were added to each well of a 96-well tissue culture plate (Falcon) and dried overnight in a 37⁰C oven. The plates were blocked by the addition of 100 µl 5% bovine serum albumin (BSA) in PBS. A 50 µl aliquot of sFv23/TNF fusion protein and serial 2-fold dilutions were then added to appropriate wells and incubated for 1 hour at room temperature. After three washes (PBS/Tween-20), anti-sFv23 rabbit polyclonal antibody (1 µg/ml in PBSA/Tween-20) was added and

incubated for 1 hour. The wells were tapped dry and then, 100 µl of horseradish peroxidase conjugated goat anti-rabbit antibody (Boehringer Mannheim) was added. The plates were developed by addition of ABTS substrate in 0.1 M citrate buffer, pH 4.2 (Sigma Chemical Co.) and incubated for 1 hour. The optical density was measured at 405 nm on a Bio-Tek Autoreader.

The binding of both the native sFv23 single-chain antibody and the sFv23/TNF fusion construct is shown in Figure 28. The binding of both agents was similar and appeared to be dose-dependent. A slightly higher binding of the fusion construct compared to the antibody was noted at the highest concentrations tested. As shown in Figure 29, optimal binding to target cells occurred after incubation with 0.75 uM concentration of the fusion construct. There was no apparent binding of the construct to antigen negative (A-375) cells tested under identical conditions (Figure 29).

EXAMPLE 35

Cytotoxicity of scFv-23-TNF immunotoxin.

The cytotoxicity of TNF and scFv23/TNF fusion toxin was determined based on cytotoxicity to the transformed murine fibroblast cell line L-929 cells. Log-phase cells in culture media

(RPMI 1640 with 1.5 mM glutamine and 10% FBS) were plated in a 96-well tissue culture plate (Falcon) at a density of 2×10^4 cells/well and incubated overnight at 37°C, 5% CO₂ atmosphere. Then, 200 µl of TNF in PBS starting at 100 units/ml and serial dilutions were added in the presence of actinomycin D (0.5 µg/ml, Sigma Chemical Co, St. Louis, MO). Likewise, serial dilutions of sFv23-TNF were added and the plate was incubated for 24 hours. The surviving adherent cells were then stained by the addition of 100 µl of crystal violet (0.5% (w/v) in ethanol). The stain was incubated on the plates for 0.5 h, excess stain was removed, the plates were washed with water, allowed to air dry and the remaining dye was solubilized by addition of 150 µl of Sorenson's Buffer (0.1 M sodium citrate, pH 4.2). The plates were read on a microplate ELISA reader at 540 nm.

IC₅₀ values were 1 and 100 pM for native TNF and the antibody-TNF fusion toxin, respectively. This demonstrated approximately a 100-fold decrease in the apparent specific activity for the construct compared to native TNF. This decrease in specific activity could be due to steric hindrance of the antibody interfering with interaction of TNF with its receptor site. Alternatively, since TNF operates optimally as a compact trimer in solution (39), the

antibody component could also interfere with optimal aggregation of the TNF component in solution.

5

EXAMPLE 36

Cytotoxicity of scFv-23-TNF immunotoxin against breast carcinoma.

Two breast carcinoma cell lines SKBR3-LP and SKBR3-HP were assessed for their relative expression of HER2/*neu* and for their relative sensitivity to TNF-induced cytotoxicity. Log phase SKBR-3 cells were diluted to 8000 cells/100 μ l in media. Aliquots (100 μ l) were added to Falcon 96 well flat-bottomed tissue culture plates and incubated 24 hours at 37⁰C with 5% CO₂. Purified sFv 23/TNF or recombinant human TNF were diluted 1:8 in culture media (McCoy's 5a with 1.5 mM glutamine and 10% fetal bovine serum). Aliquots each sample (200 μ l) were added to the plate in 2-fold serial dilutions. The plates were further incubated at 37⁰C, 5% CO₂ for 72 hours. Remaining adherent cells were stained by the addition of 150 μ l with of crystal violet (Fisher Scientific) 0.5% in 20% methanol). Dye-staining cells were solubilized by addition of 150 μ l of

Sorenson's Buffer (0.1 M sodium citrate, pH 4.2 in 50% ethanol) and the plates were read at 540 nm in an ELISA plate reader.

As shown in Figure 30, the LP variant cell line contained relatively higher levels of HER2/*neu* compared to the HP line (Figure 6, insert). In addition, the LP line demonstrated a much greater resistance to TNF-induced cytotoxic effects. The cytotoxicity of TNF and scFv23/TNF was assessed against log-phase SKBR3-HP cells and as demonstrated in Figure 31, the IC₅₀ for TNF exceeded 40,000 Units/ml while the scFv23/TNF fusion toxin demonstrated an IC₅₀ value of 2000-fold lower at approximately 20 Units/ml. In contrast, the activity of both TNF and scFv23/TNF were assessed against TNF-resistant cells expressing higher levels of HER2/*neu*. As shown in Figure 32, these cells were relatively resistant to the cytotoxic effects of both scFv23/TNF construct and to TNF. However, the IC₅₀ of TNF against these cells was 5×10^7 U/ml while that of the sFv23/TNF construct was approximately 20-fold lower (2×10^6 U/ml).

In conclusion, the present invention and the embodiments disclosed herein are well adapted to carry out the objectives and obtain the ends set forth at the outset. Certain changes can be made in the method and apparatus without parting from the spirit and scope of this invention. It is realized that

changes are possible and it is further intended that each element or step recited in any of the following claims is to be understood as referring to all equivalent elements or steps for accomplishing substantially the same results in substantially the same or
5 equivalent manner. It is intended to cover the invention broadly in whatever form its principles may be utilized. The present invention is therefore well adapted to carry out the objects and attain the ends and advantages mentioned, as well as others inherent therein.

WHAT IS CLAIMED IS:

1. A composition comprising a conjugate of an antibody exhibiting binding specificity for an extracellular epitope of c-erbB-2 protein and a plant derived toxin, wherein said toxin is pharmacologically effective against neoplastic cells and is selected from the group consisting of gelonin, full length recombinant gelonin, functional gelonin fragments and functional gelonin derivatives, wherein said plant derived toxin has a lower effect on cell death or arrest of cellular growth when not conjugated to said targeting moiety.

2. The composition of Claim 1, wherein said antibody is an intact full-length immunoglobulin.

3. The composition of Claim 1, wherein said antibody is an intact full-length immunoglobulin.

4. The composition of Claim 1, wherein said antibody is an Fv fragment, and the V_H peptide is conjugated with said toxin.

5. The composition of Claim 1, wherein said antibody is an sFv, and the V_L peptide is conjugated with said toxin.

6. The composition of Claim 1, wherein said antibody is a single chain antibody.

7. The composition of Claim 1, wherein said conjugate is a fusion protein between said antibody and said toxin.

8. The composition of Claim 7, wherein said composition is recombinantly produced by fusing the gene encoding said antibody to the gene encoding gelonin.

9. The composition of Claim 1, wherein said antibody is selected from the group consisting of TAb 250 and BACH-250.

10. A pharmaceutical composition comprising the composition of claim 1 and a pharmaceutically acceptable vehicle.

11. A method of treatment against neoplastic cells, comprising the step of administering an effective dose of the composition of Claim 1 to said cells.

5 12. The method of Claim 11, wherein said neoplastic cells are characterized by over-expression of c-erbB-2 protein.

10 13. The method of Claim 11, wherein said neoplastic cells are selected from the group consisting of mammary carcinoma cells, ovarian carcinoma cells, lung carcinoma cells, salivary gland carcinoma cells, gastric tumor cells, colon adenocarcinoma cells, and bone marrow leukemia cells.

15 14. The method of claim 11, wherein said treatment is used against neoplastic cells in a human or an animal.

20 15. A composition comprising a conjugate of tumor necrosis factor to an antibody exhibiting binding specificity for an extracellular epitope of c-erbB-2 protein.

16. The composition of Claim 15, wherein said conjugate is a fusion protein between said single chain antibody and tumor necrosis factor.

17. The composition of Claim 15, wherein said conjugate is recombinantly produced by fusing a gene encoding said single chain antibody to a gene encoding tumor necrosis factor.

5 18. The composition of Claim 15, wherein said single chain antibody is scFv-23.

19. The pharmaceutical composition, comprising the composition of claim 15 and a pharmaceutically acceptable vehicle.

20. A method of treatment against neoplastic cells, comprising the step of administering an effective dose of the composition of Claim 19 to said cells.

21. The method of Claim 20, wherein said neoplastic cells are selected from the group consisting of mammary carcinoma cells, ovarian carcinoma cells, lung carcinoma cells, salivary gland carcinoma cells, gastric tumor cells, colon adenocarcinoma cells, and bone marrow leukemia cells.

ABSTRACT OF THE DISCLOSURE

Novel immunotoxins and methods of treating neoplastic diseases are provided. More specifically, immunotoxins comprised conjugation of a c-erbB-2 targeting moiety and a cell growth
5 modulator are provided. These immunotoxins specifically and selectively kill tumor cells that over-express the c-erbB-2 protein. The novel immunotoxins would be useful in treating human mammary carcinomas, human ovarian carcinomas, lung carcinomas, gastric tumors, salivary gland adenocarcinomas, and colon
10 adenocarcinomas.

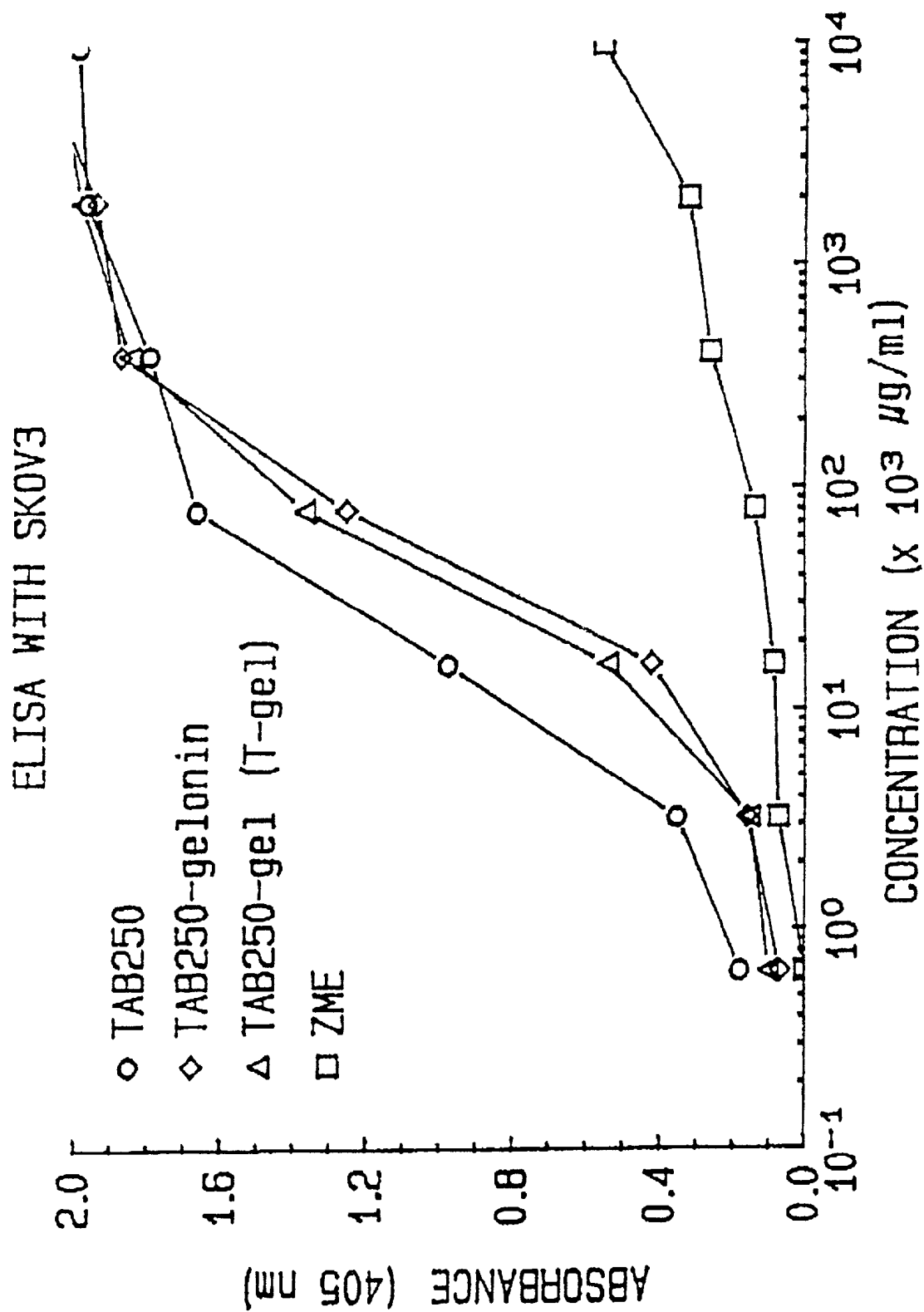


FIGURE 1

CYTOTOXICITY OF TAB250-GELONIN (SKOV3)

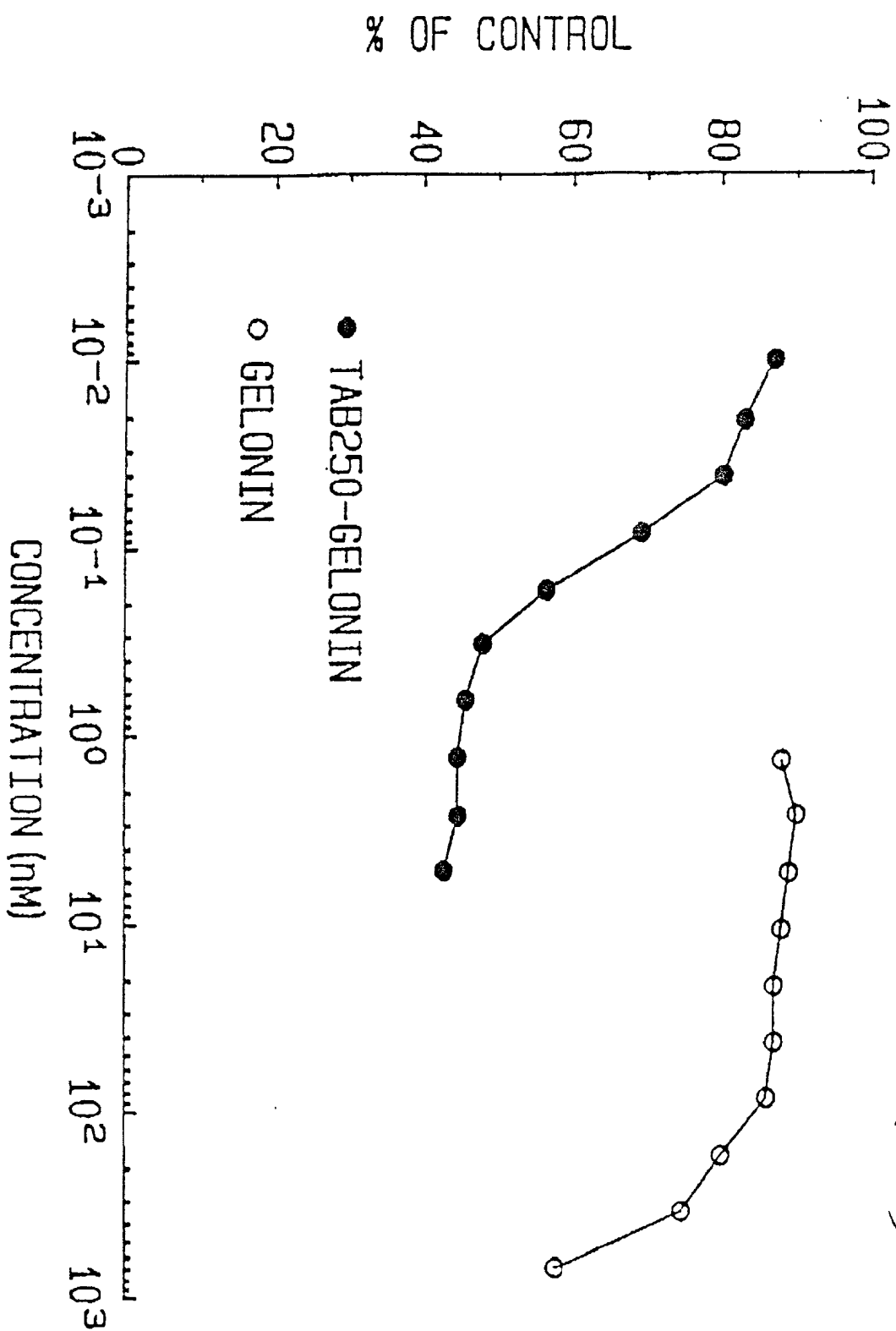


FIGURE 2

COMPETITION OF RELEVANT VS IRRELEVANT ANTIBODY WITH CONJUGATE ON SKOV3

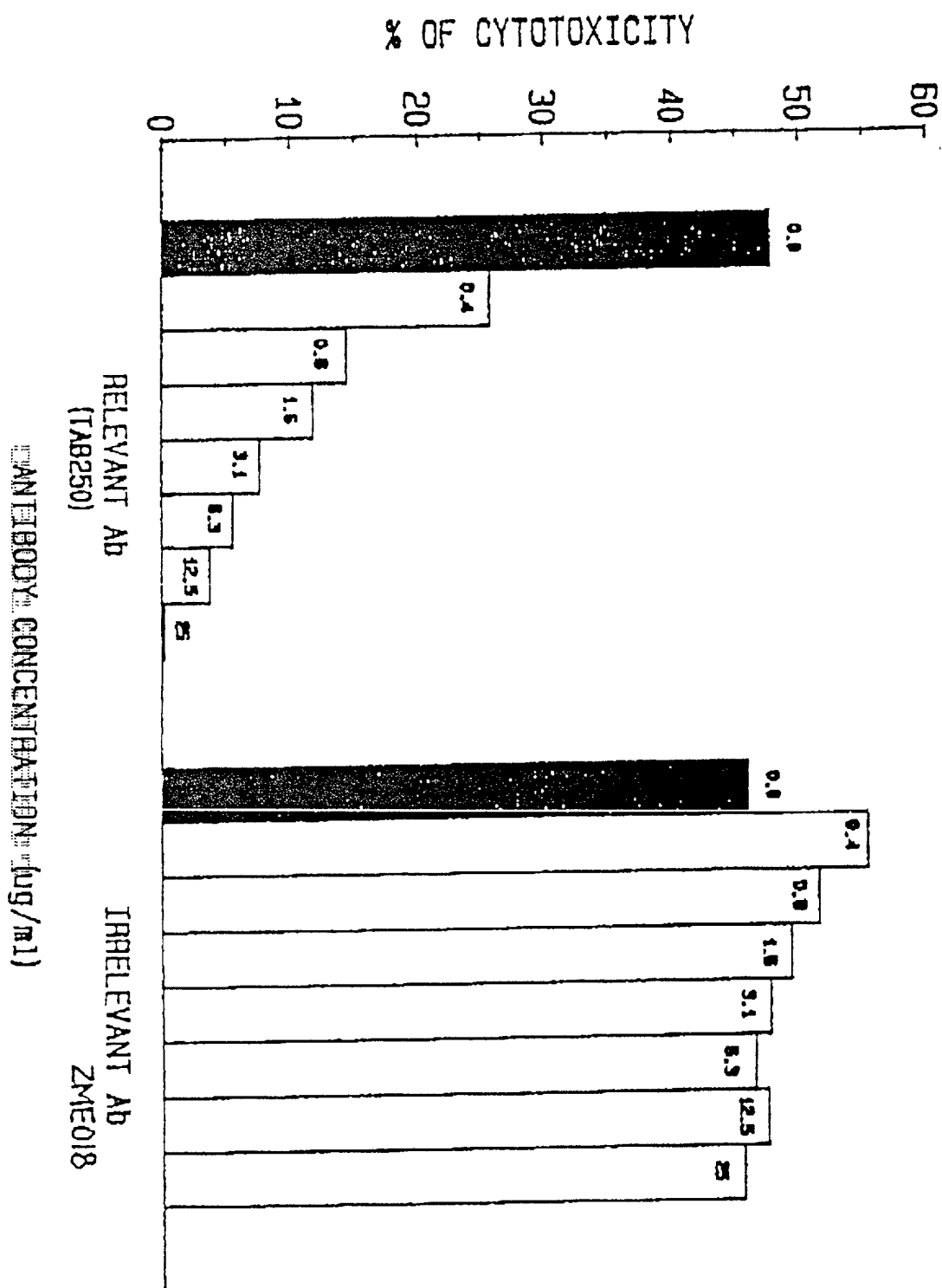


FIGURE 3

CPA
1Ab 250-Gelatin Dose Response
on SKOV3 cells

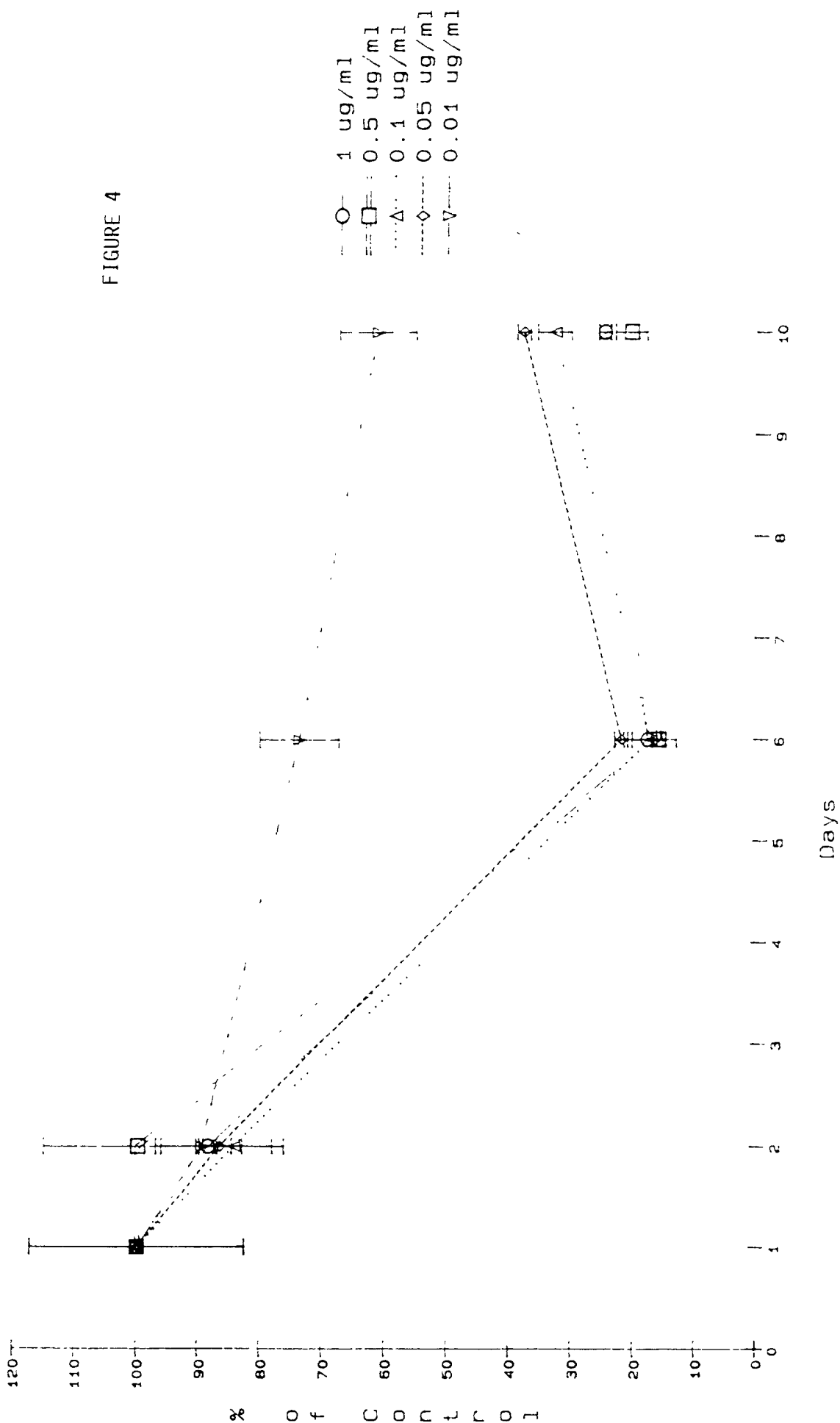


FIGURE 4

GCPA91991

CPA
Effect of TAB 250 vs. TAB 250-Gelonin
on SKOV3 cells

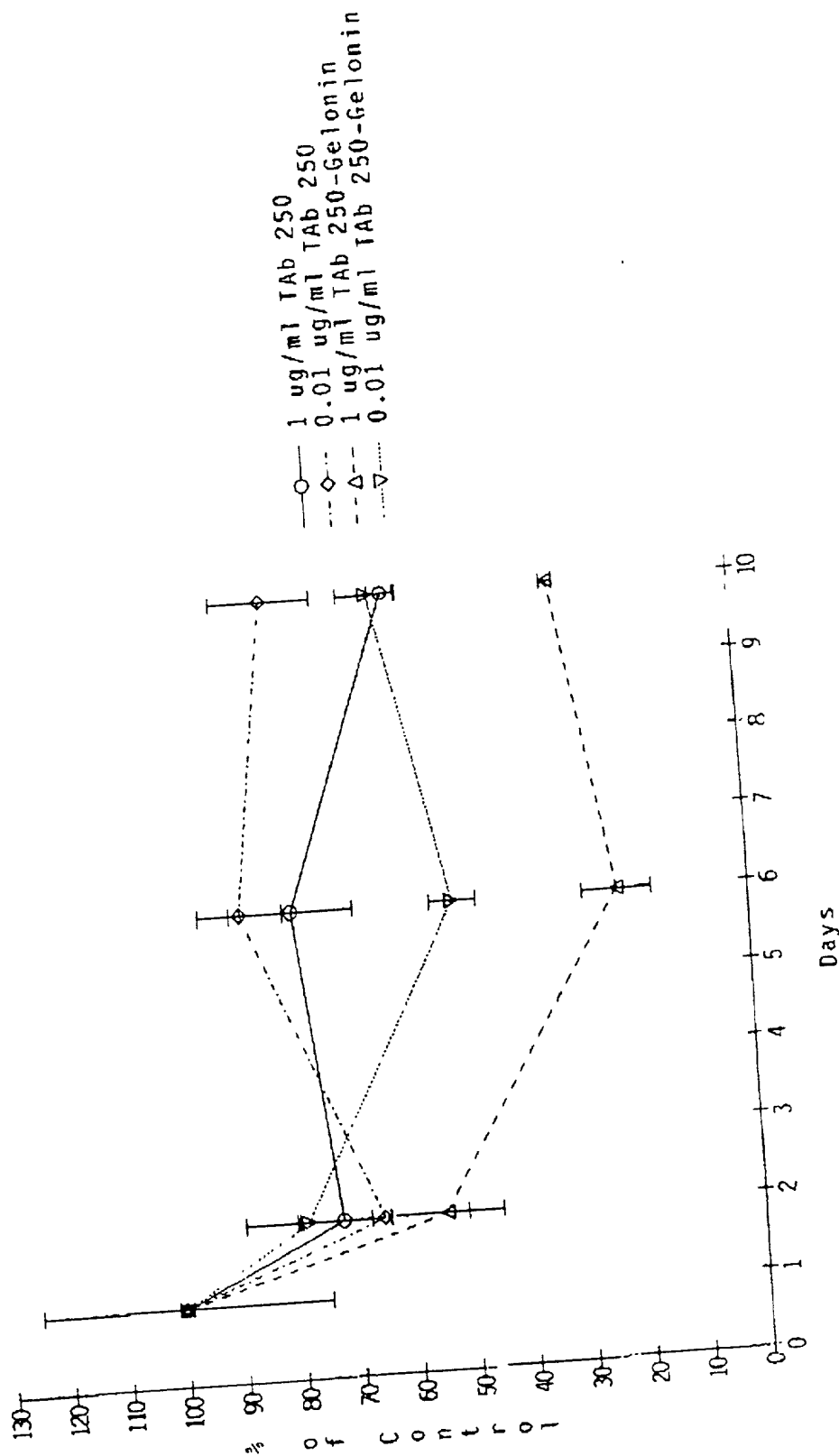
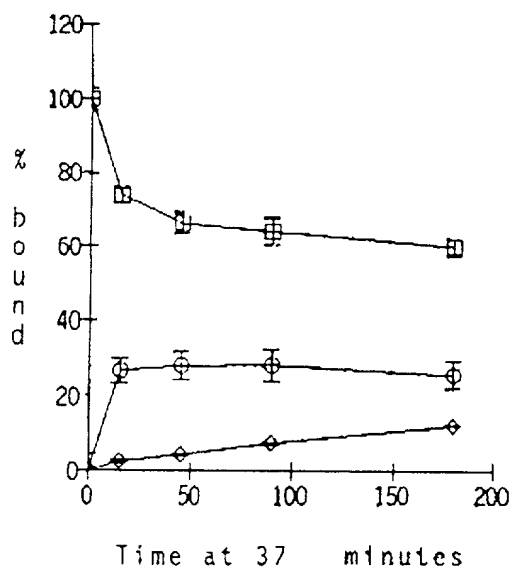


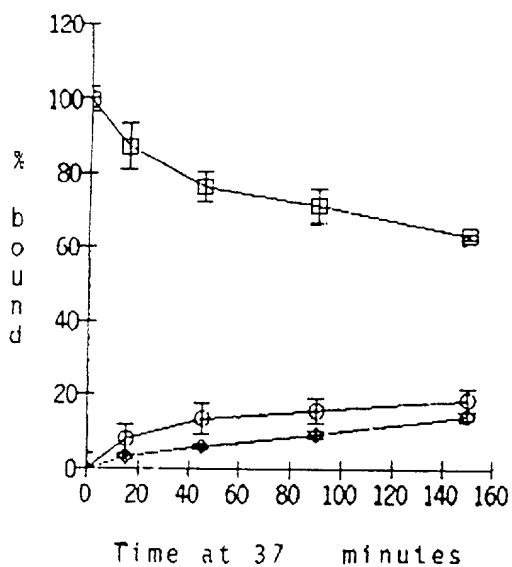
FIGURE 5

Internalization of Tab 250
in SKBR3 cells



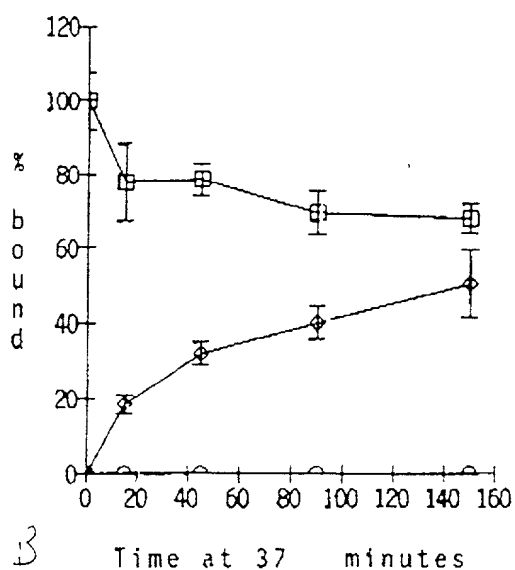
—○— internalized
—◇— dissociated/recycled
—□— surface bound

Internalization of Tab 250
in SKOV3 cells



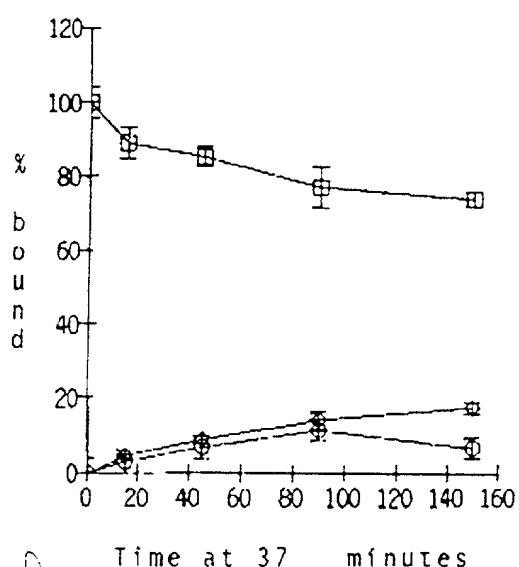
—○— internalized
—◇— dissociated/recycled
—□— surface bound

Internalization of Tab 250
in MDA 231 cells



—◇— dissociated/recycled
—□— surface bound
—○— internalized

Internalization of Tab 250
in MDA 453 cells



—○— internalized
—◇— dissociated/recycled
—□— surface bound

FIGURE 6

GM9591

TAB 250-Gelonin Dose Response MIT Assay

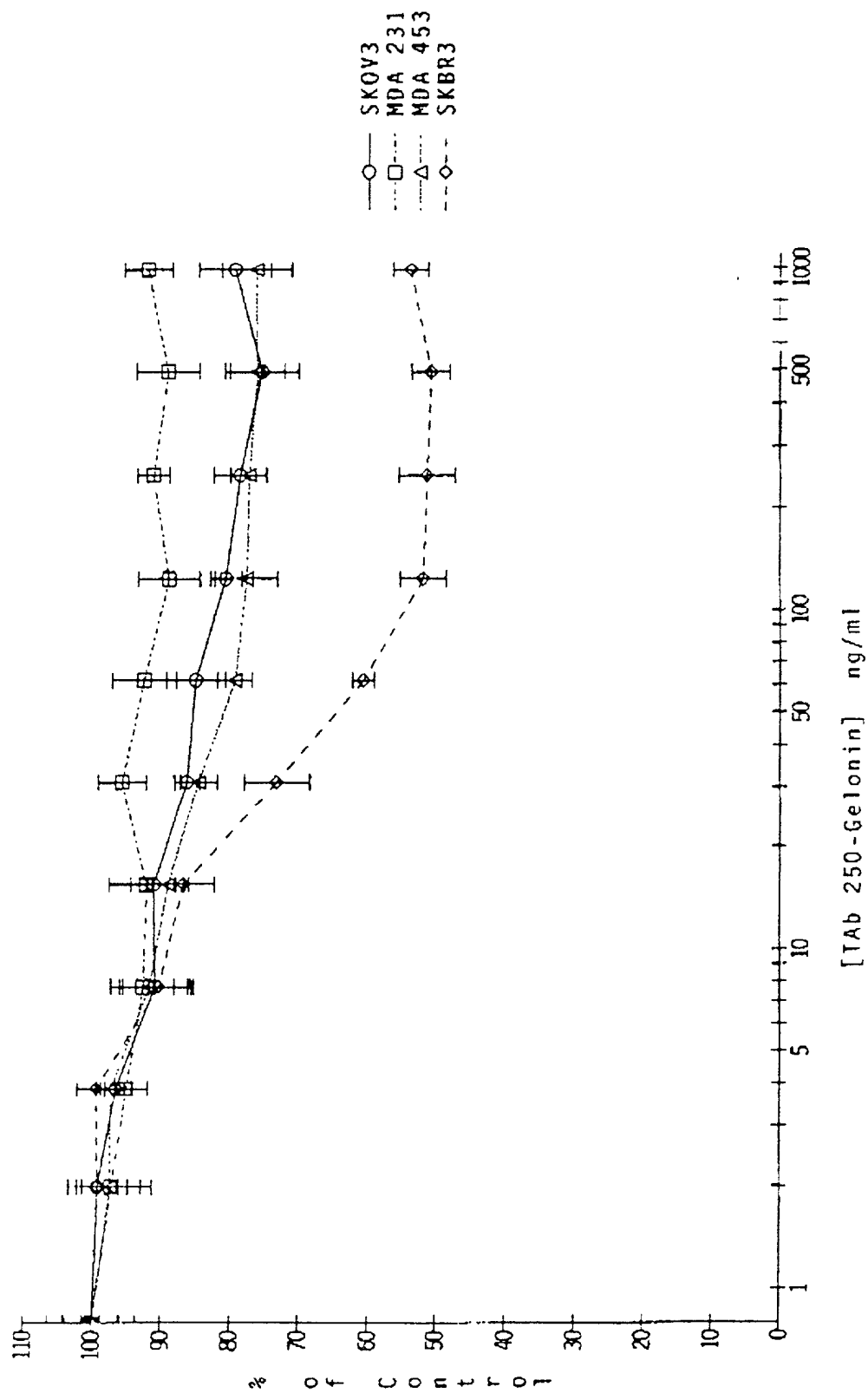


FIGURE 7

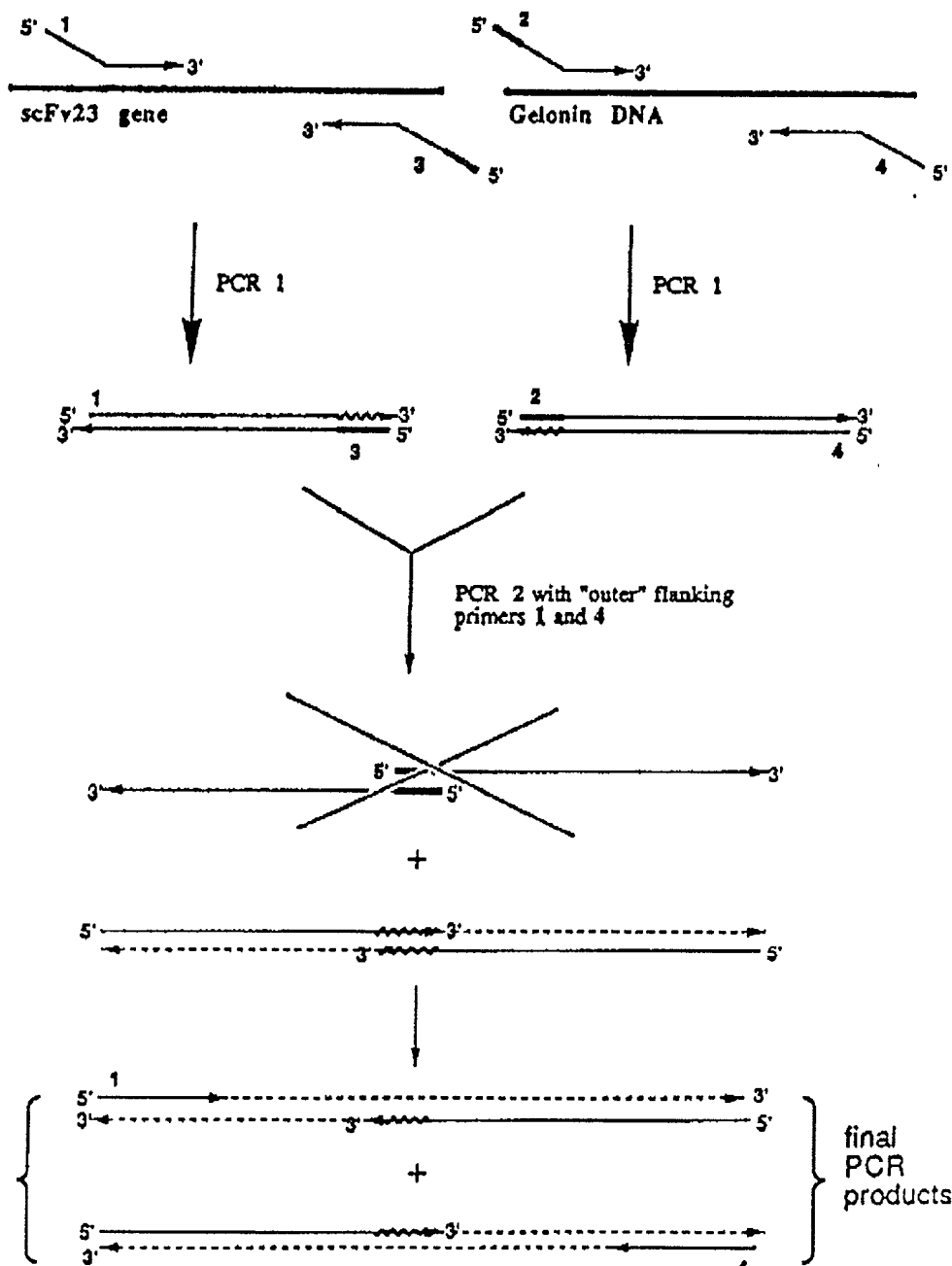
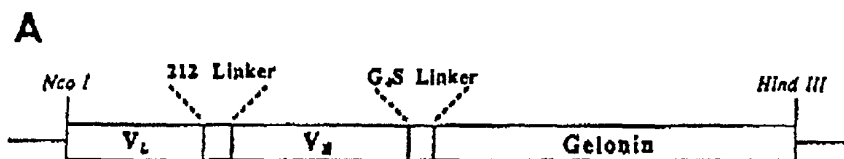


FIGURE 8

Figure 8: Construction of the scFv23 - Gelonin Immunotoxin gene. The first PCR syntheses were performed in separate tubes and one tenth of each of the reaction products were subsequently combined and used as templates for the second PCR (Davis et al. (1991) *BioTechnology* 9, 165-169). Nco I and Hind III restriction sites were included into the sequences of the "outer" flanking primers 1 and 4 for future cloning purposes. Homology between bases in the PCR primers 2 and 3 is indicated by the filled boxes at the 5' ends of each primer and represented DNA sequences encoding the Gly-Gly-Gly-Gly-Ser (G₄S) linker between the antibody and gelonin fragments of the immunotoxin.



B

V_L scFv23

CCC ATG GCG ATG TCT GAC CTC CAG CTG ACC CAG TCT CCA CCA ATC CTC TCT CCA TCT
Pro Met Ala Met Ser Asp Val Gln Leu Thr Gln Ser Pro Ala Ile Leu Ser Ala Ser

LCDR1

CCA GCG CAG AAG GTC ACA ATC ACT TGC AAG GCG AGC CCA AGC GTA AGT TAC GTC CAG
Pro Gly Glu Lys Val Thr Met Thr Cys Arg Ala Thr Phe Ser Val Ser Tyr Met Ile

LCDR2

TGG TAT CAG CAG AAG CCA GGA TCG TCG CCC AAA CCG TGG ATT TAT AGC ACA TCG AAC
Trp Tyr Gln Gln Lys Pro Gly Ser Ser Phe Lys Pro Trp Ile Tyr Thr Thr Ser Asn

LCDR3

CTC ACA CTC ACC ACA CTC CAG GCT GAA GAT GCT GCG ACT TAT TAC TCG CAG GAG TGG
Leu Thr Val Ser Arg Val Glu Ala Glu Asp Ala Ala Thr Tyr Tyr Cys Gln Gln Trp

312 linker

AGT GAG AAG CCA CCC ACC TTC CCA CCG GCG TCC AAG CTC GAA ATA AAA GGT TCT ACC
Ser Arg Ser Ser Ser Thr Phe Gly Gly Gly Ser Lys Leu Glu Ile Lys Gly Ser Thr

V_H scFv23

TCT GGT TCT GGT AAA TCT TCT GAA GGT AAA GGT GTC CAG CTC CAG CAG TCA CCA CCG
Ser Gly Ser Gly Lys Ser Ser Glu Gly Lys Val Gln Leu Glu Glu Ser Gly Pro

HCDR1

CAG CTG CTC AAG CCG CCA GGT TCA ATC AAG ATA TCG TCG AAG ACT TCT GGT TAC TCA
Glu Val Val Lys Pro Gly Gly Ser Met Lys Ile Ser Cys Lys Thr Ser Gly Tyr Ser

HCDR2

ATT CCA GTT ATT AAT CCG TAC AAT GGT GAT ACT AAT TAC AAT CAG AAG TTT AAG GCG
Ile Gly Leu Ile Asn Phe Tyr Asn Gly Arg Thr Asn Tyr Asn Glu Lys Phe Lys Gly

HCDR3

AAG GCG ACA TTT ACT GTA CAG AAG TCG TCG AGC ACA GCG TAC ATG CAG CTC CTC AGT
Lys Ala Thr Phe Thr Val Asp Lys Ser Ser Ser Thr Ala Tyr Met Glu Lys Leu Ser

312 linker

CTG ACA TCT CAG CAG TCT CCA CTC TAT TAC TCT CCA CCG AAG GGT AGC GAG TGG TAT
Leu Thr Ser Thr Asp Ser Ala Val Tyr Tyr Cys Ala Arg Arg Val Thr Asp Thr Tyr

G4S linker

TTC GAT GTC TCG GCG CCA GCG ACC ACC CTC ACC CTC TCG CTC TCG TCA GGT GCG GGT
Phe Asp Val Trp Gly Ala Gly Thr Val Thr Val Ser Val Ser Ser Gly Gly Gly

Gelonin

GAC TCG GGT GTC TAT ACC GGT ACC TTC ACC ACC AAA GCG GGT ACC TAT ATT ACC TAT
Gly Ser Gly Leu Asp Thr Val Ser Phe Ser Thr Lys Gly Ala Thr Tyr Ile Thr Tyr

GTT AAT TTC CTC AAG GAA CTC CCG GTT AAA CTC AAA CCG GAA GGT AAC ACC CAT GCG
Val Asn Phe Leu Asn Glu Leu Arg Val Lys Leu Lys Pro Glu Gly Asn Ser His Gly

ATC CCG CTC CTC CCG AAA CCG GAT GAC CCG GGT AAA TCG TTC CTC CTC GCG CCG CTC
Ile Pro Leu Leu Arg Lys Gly Asp Asp Pro Gly Lys Cys Phe Val Leu Val Ala Leu

ACC AAC GAT AAC CCG CAG CTC CCA GAA ATC CCA ATC GAT GTT ACC ACC GTC TAC GTA
Ser Asn Asp Asn Gly Gln Leu Ala Gly Ile Ala Ile Asn Val Thr Ser Val Tyr Val

GTT GGT TAT CAG GTT CCG AAC CCG ACC TAC TTC TTC AAA GAT GGT CCG GAT CCA GCG
Val Gly Tyr Gln Val Arg Asn Arg Ser Tyr Phe Phe Lys Asp Ala Pro Asp Ala Ala

TAC GAA GCG CTC TTC AAA AAC ACC ATC AAA AAC CCG CTC CTC TTC CCG AAA ACT
Tyr Glu Gly Leu Phe Lys Asn Thr Ile Lys Asn Pro Leu Leu Phe Gly Gly Lys Thr

CGT CTC CAG TTC GGT GCG ACC TAC CCG ACC CTC GAA CCG GAA AAA CCG TAC CCG GAA
Arg Leu His Phe Gly Gly Ser Tyr Pro Ser Leu Glu Gly Gly Lys Ala Tyr Arg Glu

ACT ACC GAT CTC CCG ATC GAA CCG CTC CCG ACC ATC CCG ATC AAA AAA CTC GAC GAA AAC
Thr Thr Asp Leu Gly Ile Glu Pro Leu Arg Ile Gly Ile Lys Lys Leu Asp Glu Asn

GCG ATC GAC AAC TAC AAA CCG ACC GAA ATC CCG ACC TCT CTC CTC GTT GTC ATC CAG
Ala Ile Asp Asn Tyr Lys Phe Thr Glu Ile Ala Ser Ser Leu Leu Val Val Ile Gln

ATG GTC ACC GAA CCG CCA CCG TTC ACC TTC ATC GAA AAC CAG ATT CCG AAC AAC TTC
Met Val Ser Glu Ala Ala Arg Phe Thr Phe Ile Glu Asn Gln Ile Arg Asn Asn Phe

CAG CAG CCG ATG CCG CCG CCG AAC AAC ACC ATC TCT CTC GAA AAC AAA TCG CCG AAA
Gln Gln Arg Ile Arg Pro Ala Asn Asn Thr Ile Ser Leu Glu Asn Lys Trp Gly Lys

CTG ACC TTC CAG ATC CCG ACC ACC GGT GCG AAC GGT ATG TTC ACC GAA CCG CTC GAA
Leu Ser Phe Glu Ile Arg Thr Ser Gly Ala Asn Gly Met Phe Ser Glu Ala Val Glu

CTG GAA CCG CTC AAG CCG AAA AAA TAC TAC CTC ACT CCG GTC GAT CAG CTC AAA CCG
Leu Glu Arg Ala Asn Gly Lys Lys Tyr Tyr Val Thr Ala Val Asp Gln Val Lys Pro

AAA ATC CCA CTC CTC AAA TTC CTC CAG AAA GAC CAG CTC
Lys Ile Ala Leu Leu Lys Phe Leu Glu Lys Asp Glu Leu

(SEQ ID NO. 10)
(SEQ ID NO. 11)

FIGURE 9

Figure 9: Configuration and Sequence of the scFv23 - Gelonin immunotoxin. (A) The single-chain antibody fragment was cloned in the V_L-212 linker-V_H orientation and was fused to the toxin gelonin by a short, nonstructured five amino acid peptide (G₄S). (B) The DNA sequence encoding the immunotoxin is shown with the corresponding protein sequence. The six antibody complementarity determining regions (CDRs) and the 212 linker sequence between the V_L and V_H domains are highlighted; the G₄S sequence linking the antibody and toxin fragments is italicized. DNA encoding the synthetic gene for gelonin (Rosenblum et al. *Lymphokine Cytokine Res.* in press) was used as a template for the gene fusion and, in addition, encoded the targeting signal KDEL (Lys-Asp-Glu-Leu) incorporated into the 3' end.

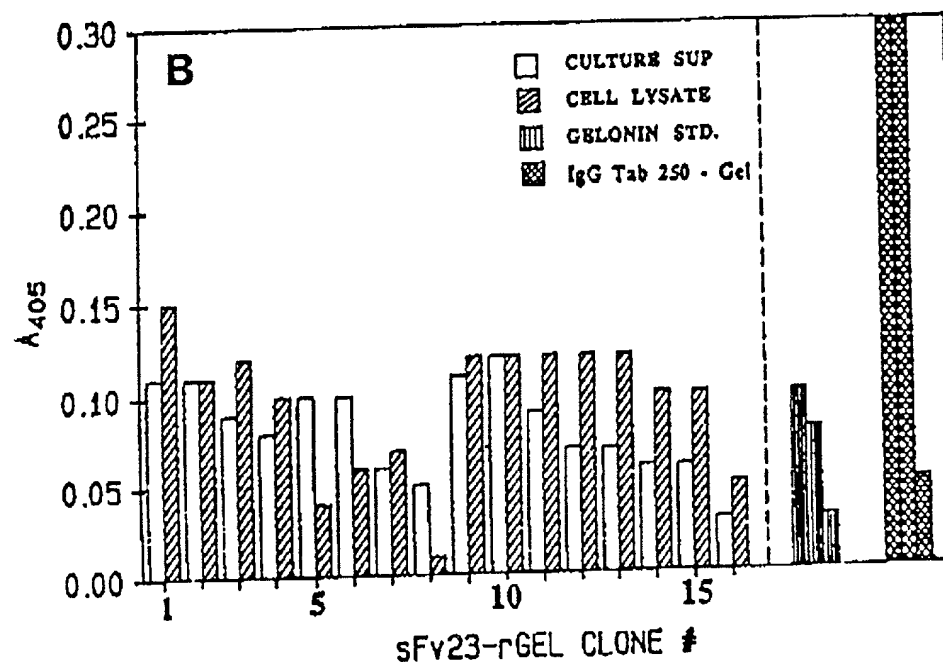
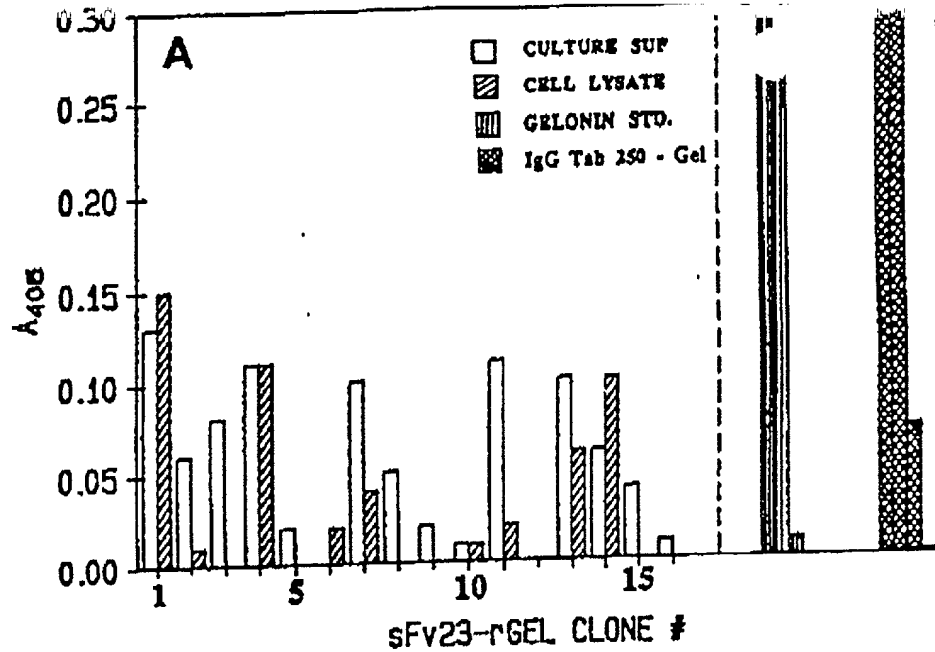


FIGURE 10

Figure 10: Binding analysis of sixteen individual scFv23 - Gelonin immunotoxin (IT) clones. Bacterial culture supernatants and periplasmic extracts of individually expressed ITs were added to wells of a 96-well ELISA plate coated with either (A) mouse *anti-gelonin* antibody 13A3, or (B) 5×10^4 antigen-positive SK-OV-3 cells. Bound IT was detected with a rabbit *anti-gelonin* polyclonal antibody followed by addition of a horseradish peroxidase (HRPO) conjugate of a goat *anti-rabbit IgG* secondary antibody. Signals were developed with the HRPO substrate ABTS and measured at 405 nm. The negative and positive controls were free gelonin (at concentrations of 50 $\mu\text{g/mL}$, 10 $\mu\text{g/mL}$, and 1 $\mu\text{g/mL}$) and a gelonin conjugate of Tab 250, a murine *anti-HER-2/neu* antibody (tested at 50 ng/mL, 10 ng/mL, and 1 ng/mL) respectively.

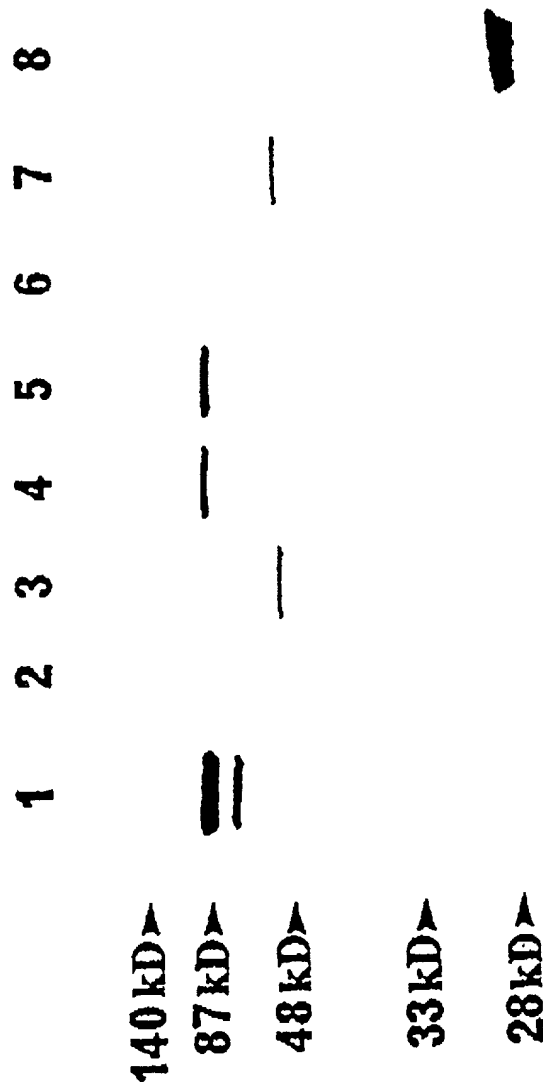


Figure 11

Figure14: Western blot analysis of purified scFvZME-Gel and scFv-23-Gel proteins refolded and purified from insoluble inclusion bodies. Refolded GST fusions of the two immunotoxins were concentrated, bound to glutathione-agarose (GSH-ag) and digested with thrombin protease. The supernatant containing cleaved protein was retreated with GSH-ag to rebind any GST and immediately bound to Blue Sepharose (B.S.) resin. Immunotoxins were subsequently eluted with 2 M NaCl and dialyzed into TBS. Nitrocellulose filters used in the Western analysis were developed using the Amersham ECL system with an exposure time of 30 sec. Molecular weight standards are indicated on the left.

Lane 1: Refolded GST-fusion of scFvZME-Gel; *Lane 2:* Flowthrough after binding scFvZME-Gel to B.S.; *Lane 3:* scFvZME-Gel eluate from B.S.; *Lane 4:* GST-scFvZME-Gel molecular weight marker; *Lane 5:* Refolded GST-fusion of scFv-23-Gel; *Lane 6:* Flowthrough after binding scFv-23-Gel to B.S.; *Lane 7:* scFv-23-Gel eluate from B.S.; *Lane 8:* Gelonin standard (40 ng)

Figure 12

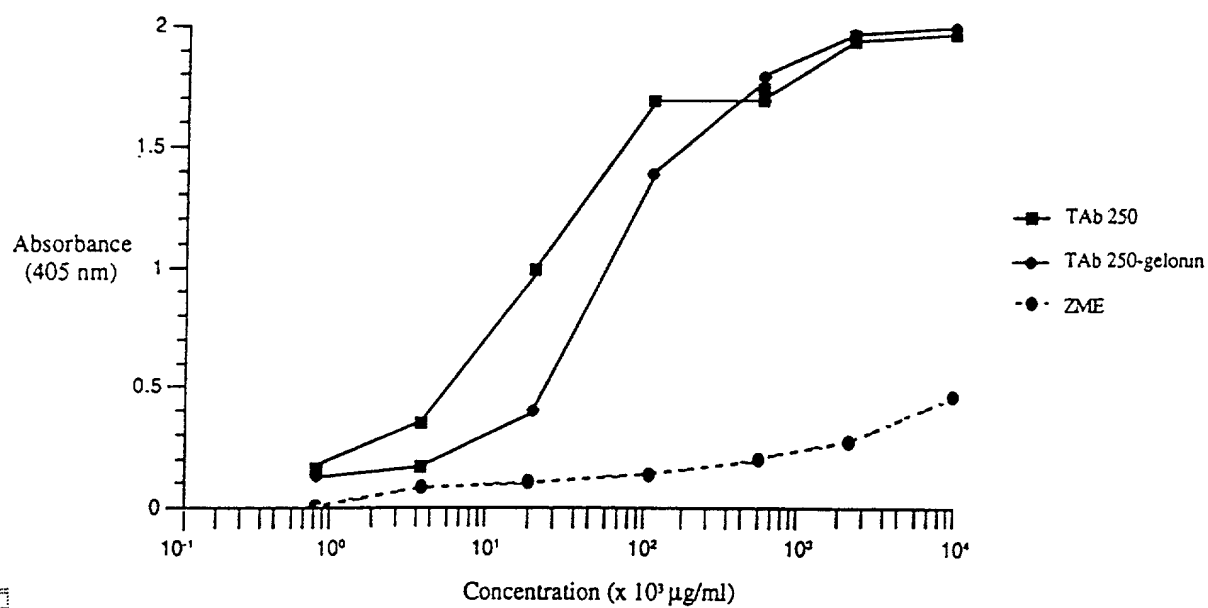


Figure 13

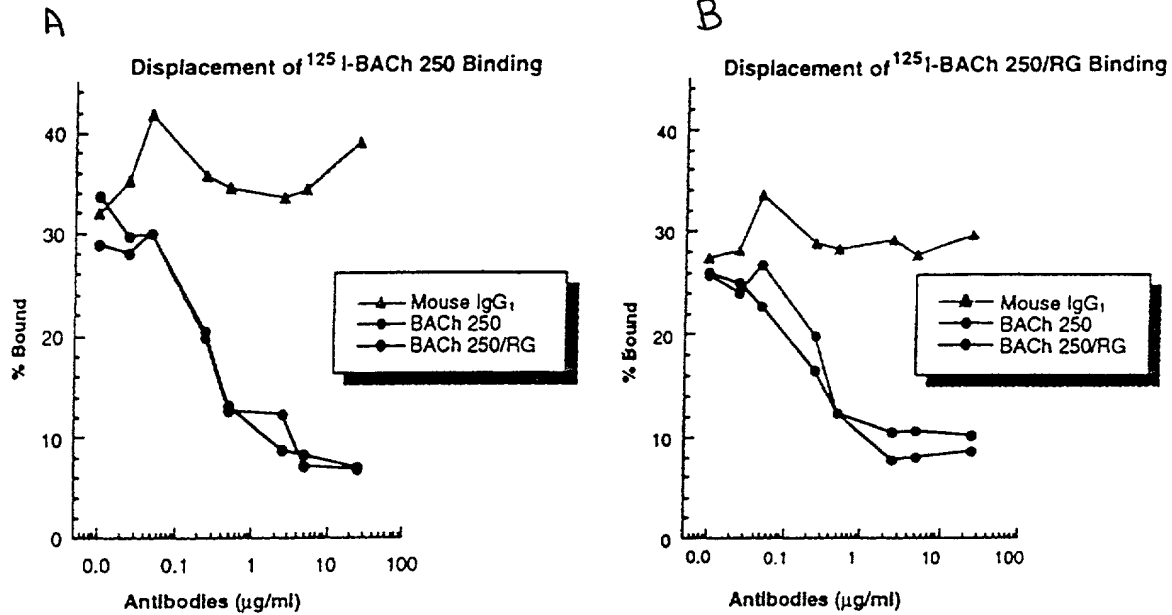


Figure 6 14

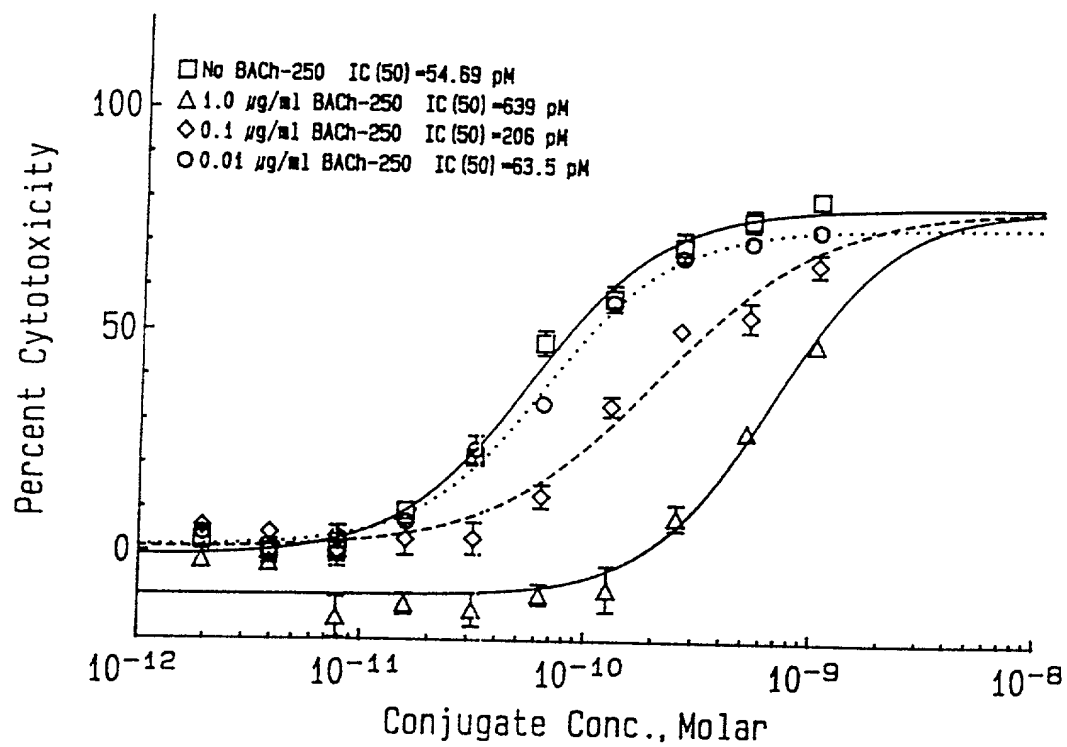


Figure 15

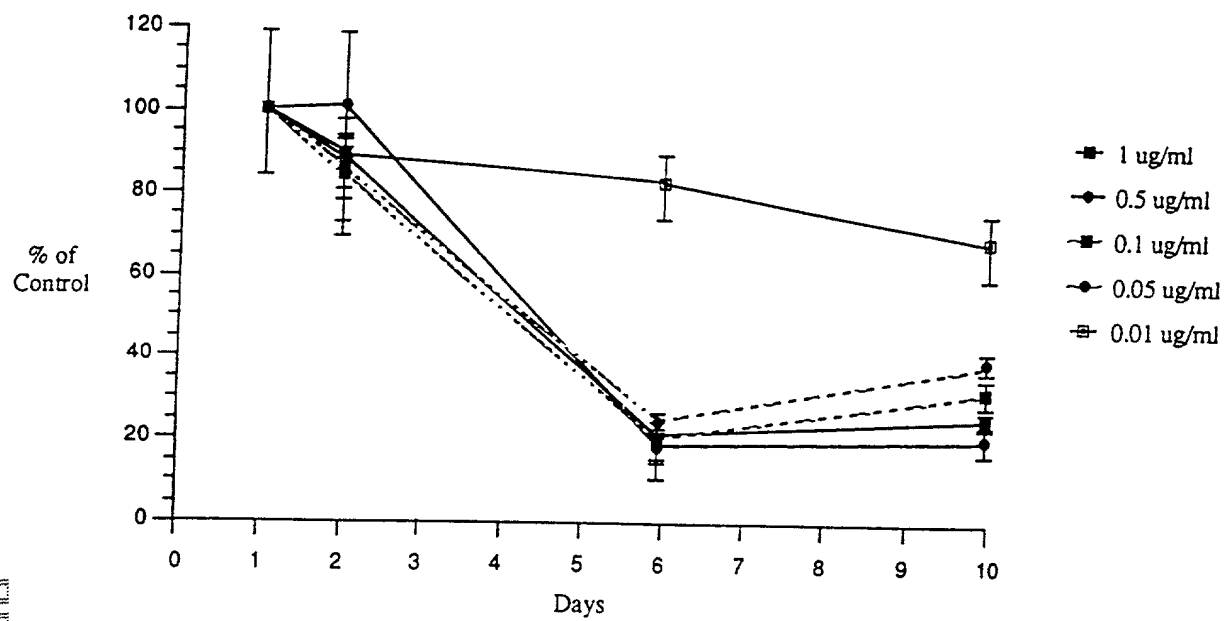
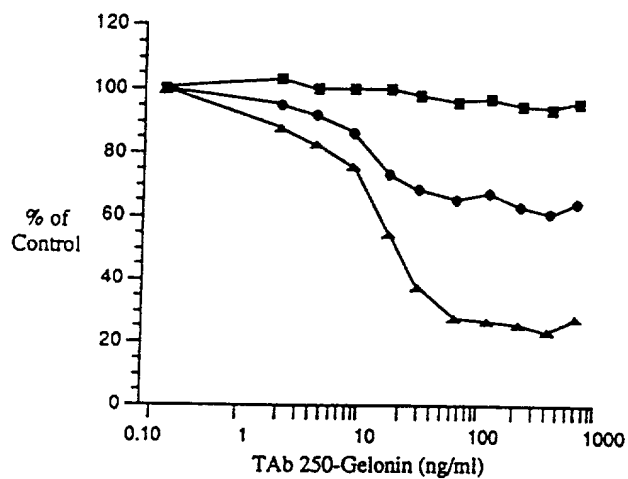
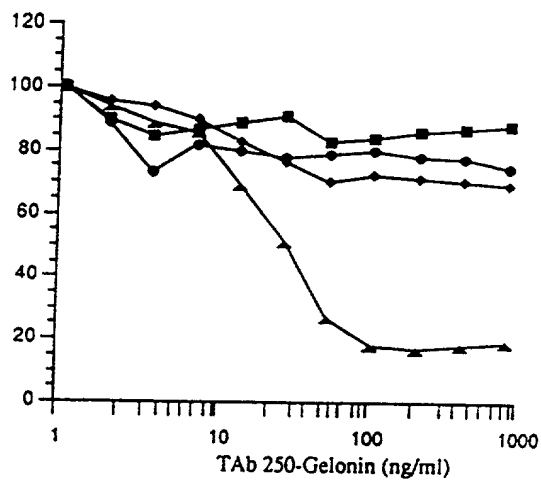


Figure 16



	# receptors
■ MDA-MB-231	16,000
◆ SKOV-3	1,200,000
▲ SKBR-3	4,000,000



	# receptors
■ MCF-7	78,000
◆ OVCAR-3	150,000
◆ MDA-MB-453	349,000
▲ SKBR-3	4,000,000

Figure 17

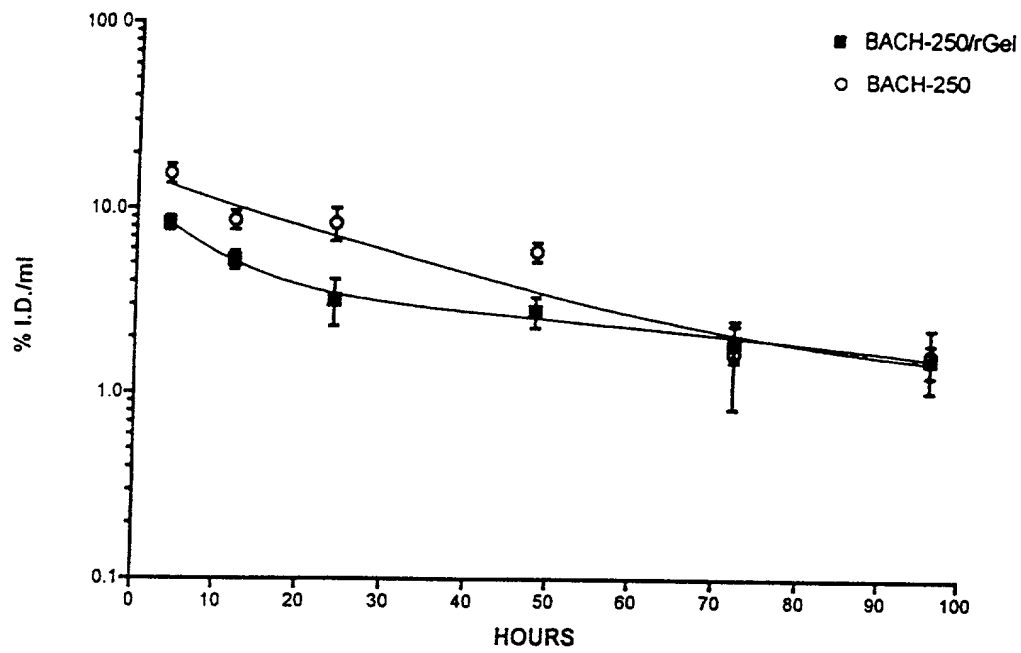


Figure 4 18

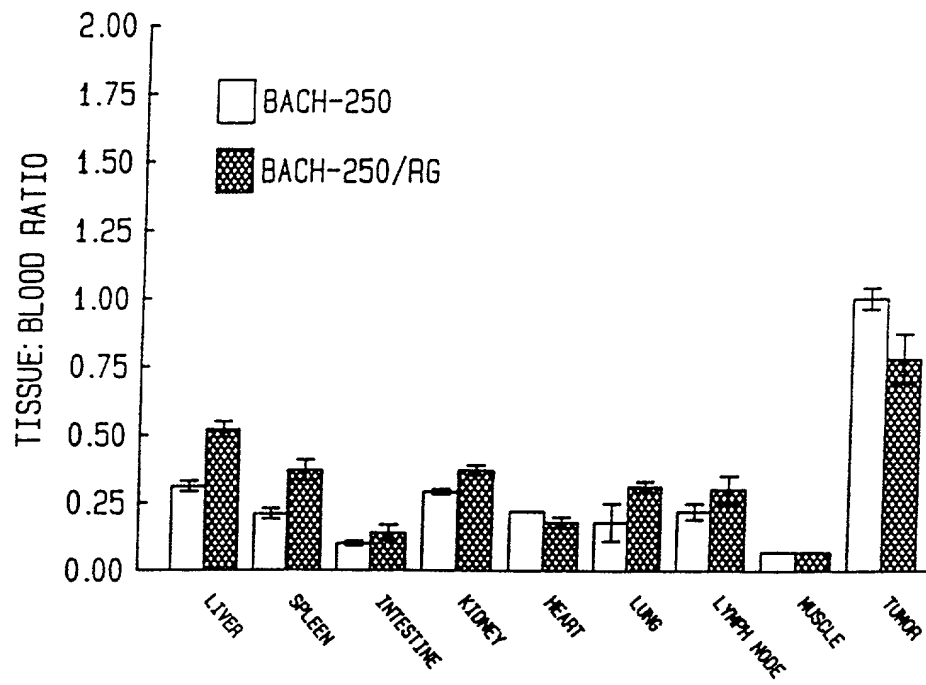


Figure 19

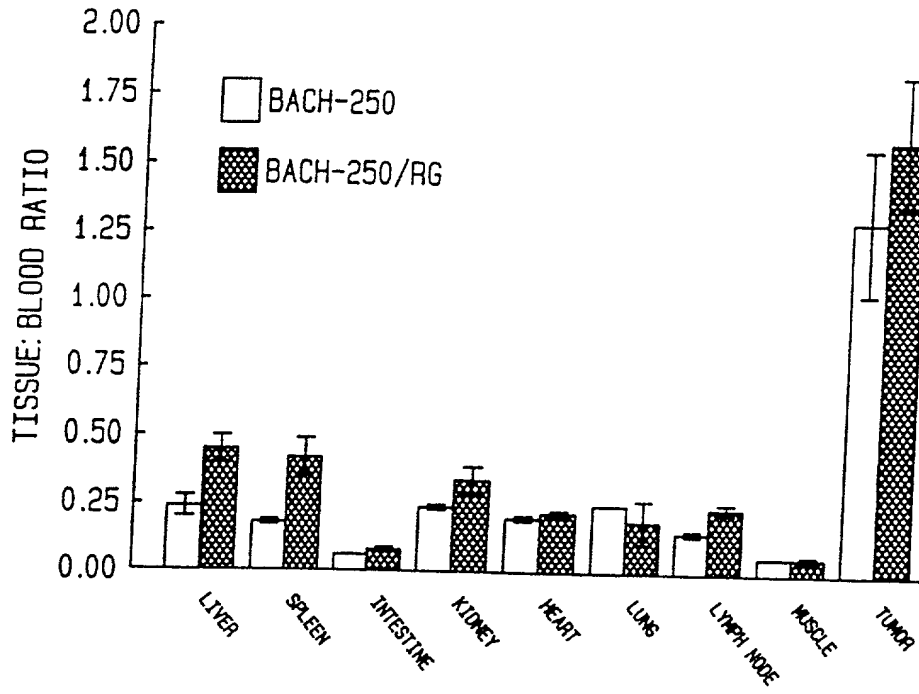


Figure 20

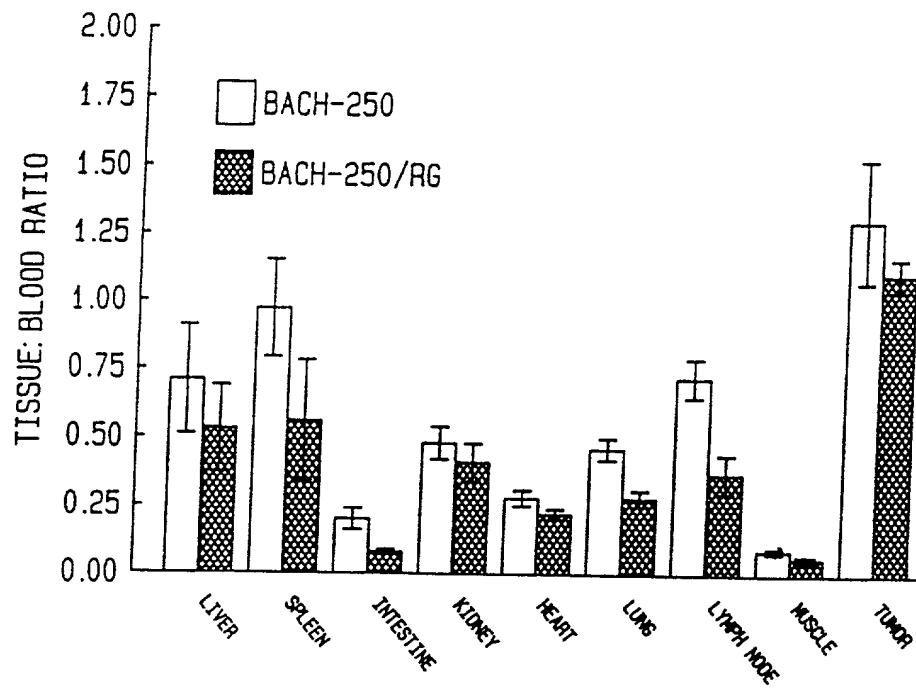


Figure 21

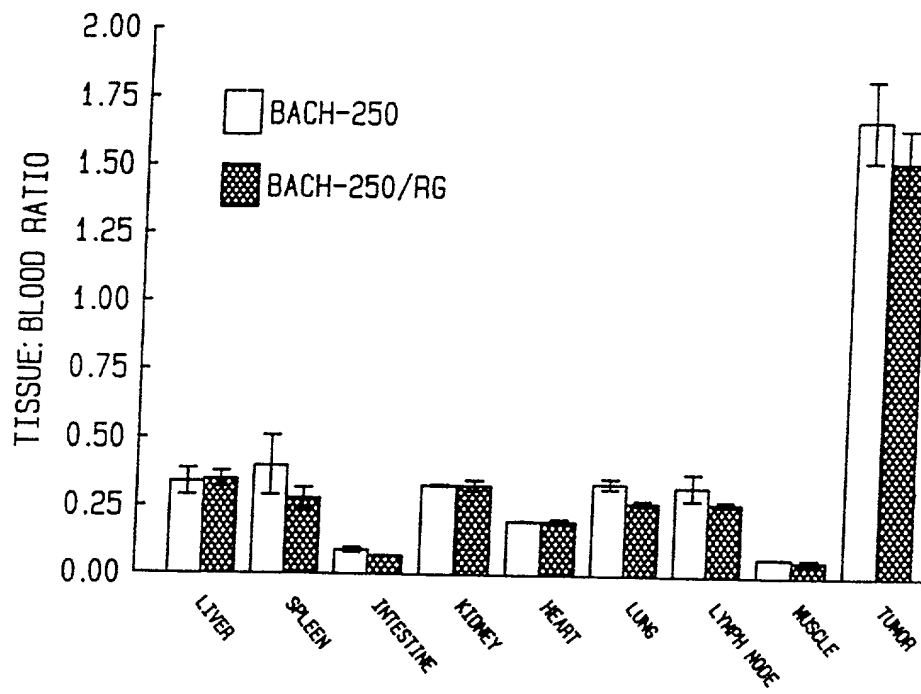


Figure 22

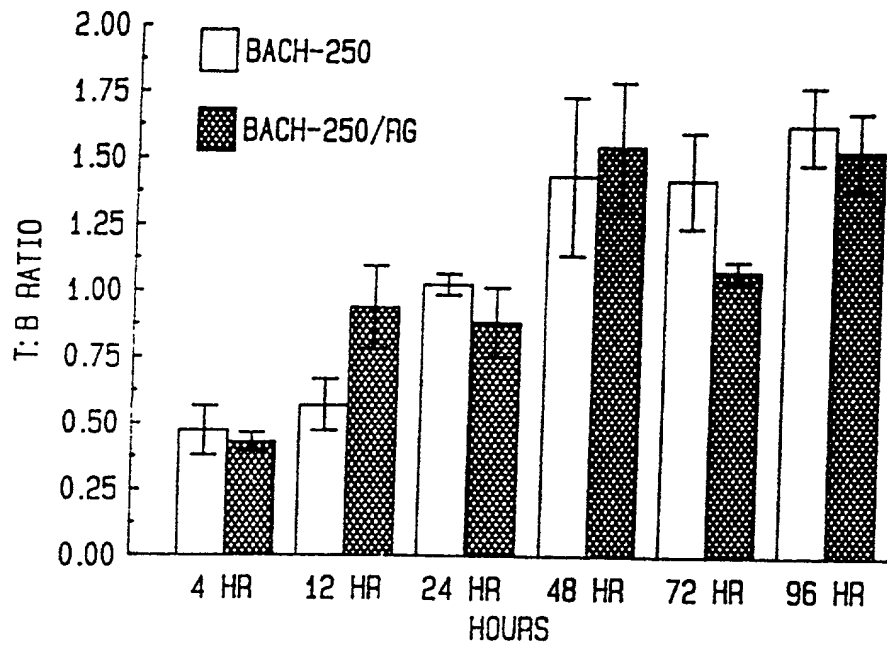


Figure 22 23

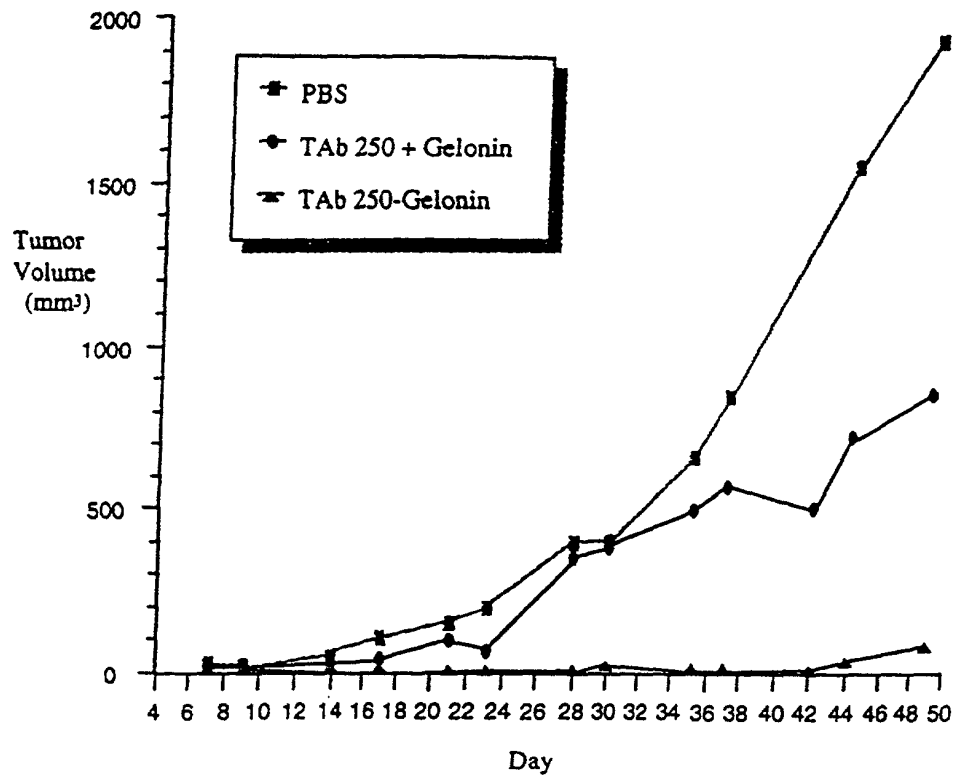


Figure 24

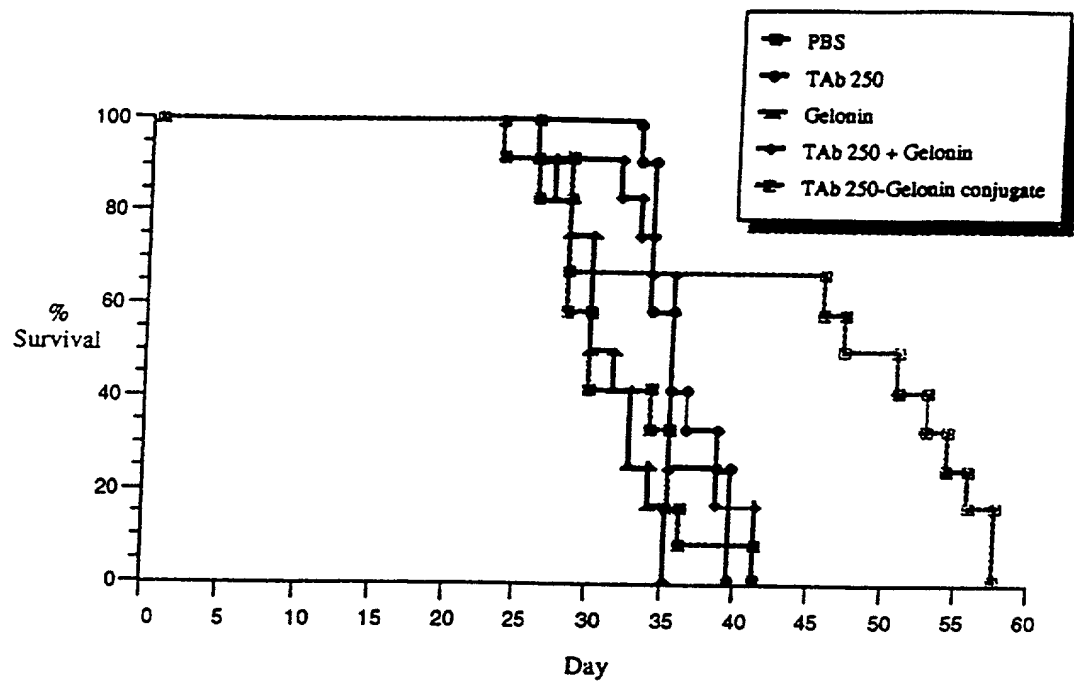


FIGURE 2 25

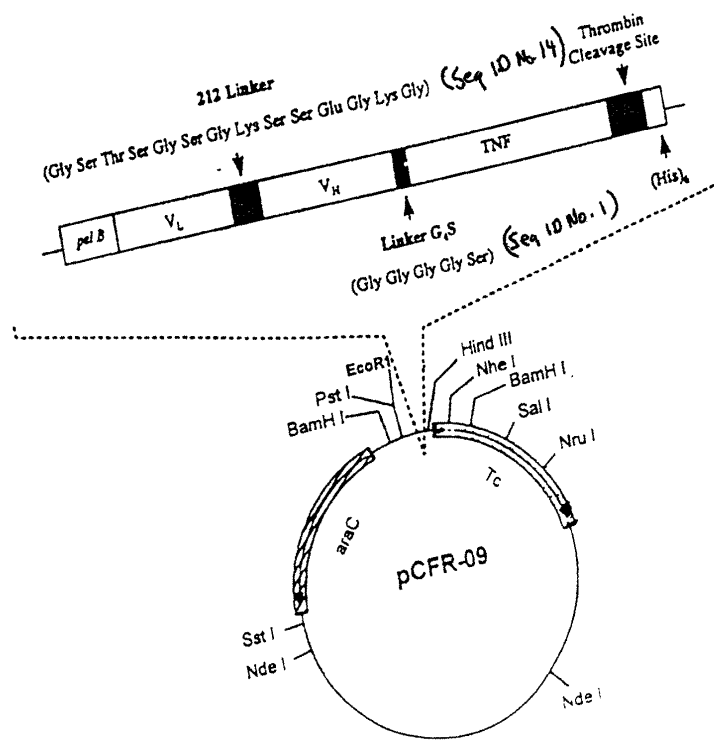


FIGURE 26

LANE SAMPLE

- A TNF(17kD) Standard
- B Uninduced sFv23 bacterial lysate
- C Induced sFv23 soluble lysate
- D Affinity (IMAC) resin prior to elution
- E sFv23 eluate from affinity resin
- F Uninduced sFv23-TNF bacterial lysate
- G Induced sFv23-TNF soluble lysate
- H Affinity (IMAC) resin prior to elution
- I sFv23-TNF conjugate from affinity resin
- J Molecular weight markers

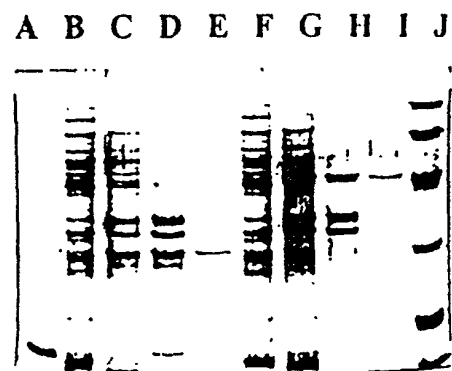


FIGURE 9 27

Coomassie

Anti-sfv23 Ab

Anti-TNF Ab

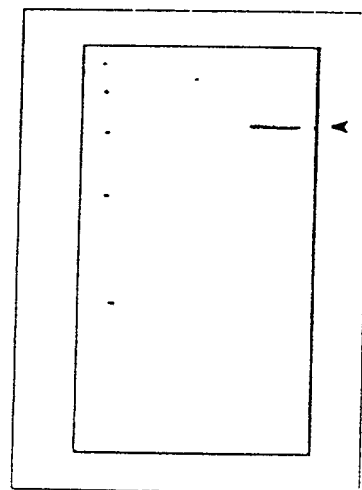
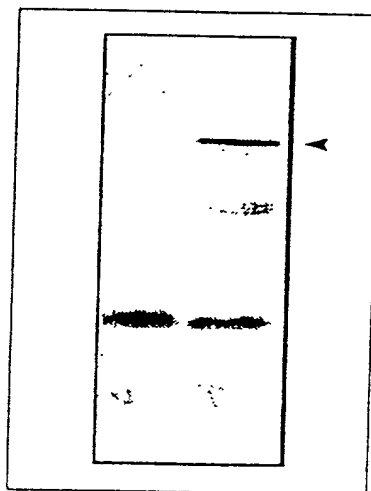
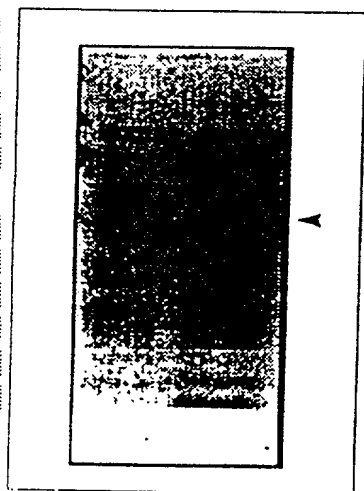


FIGURE * 28

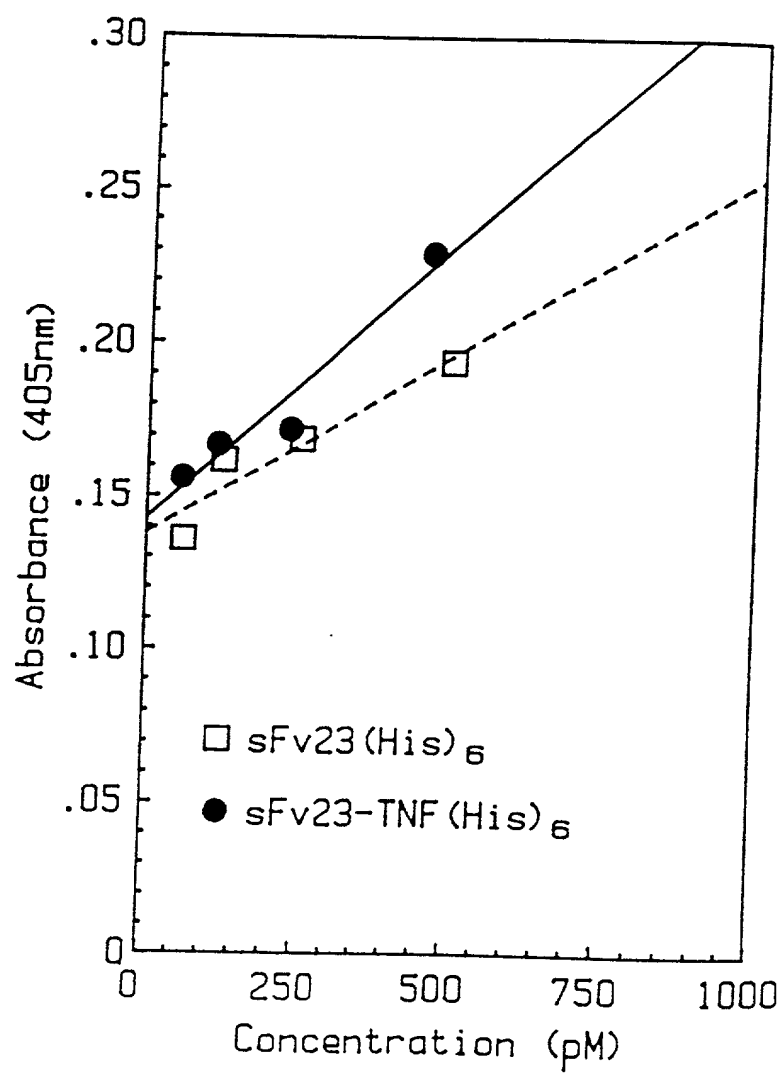


FIGURE 2 29

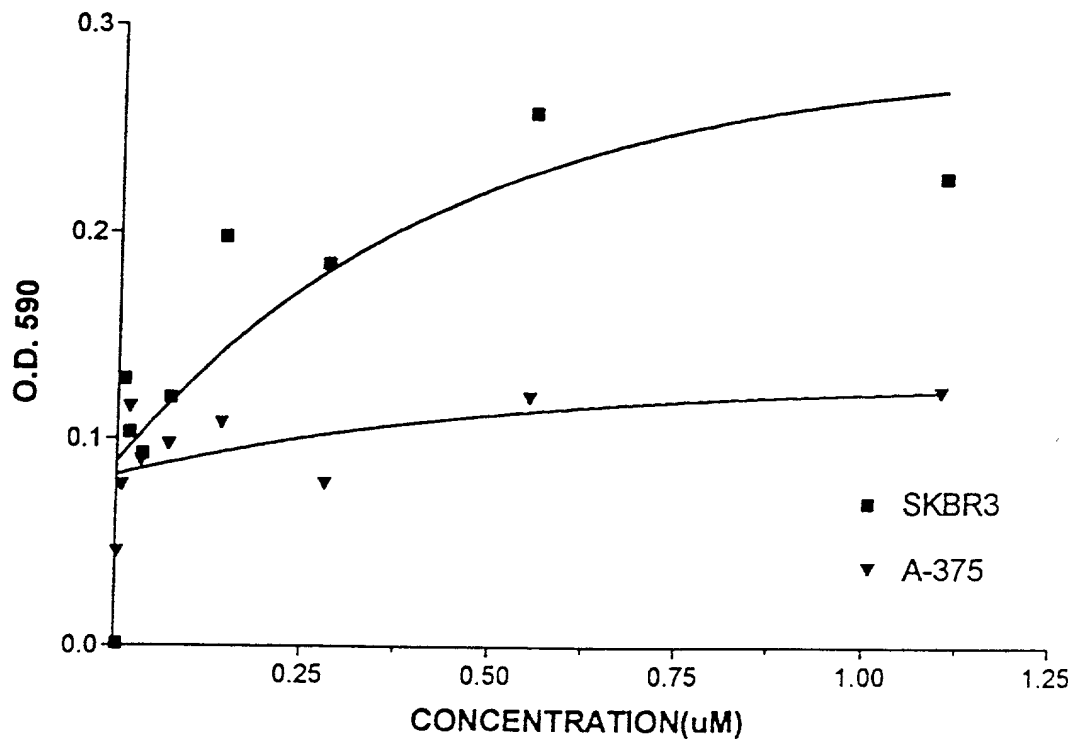


FIGURE 6 30

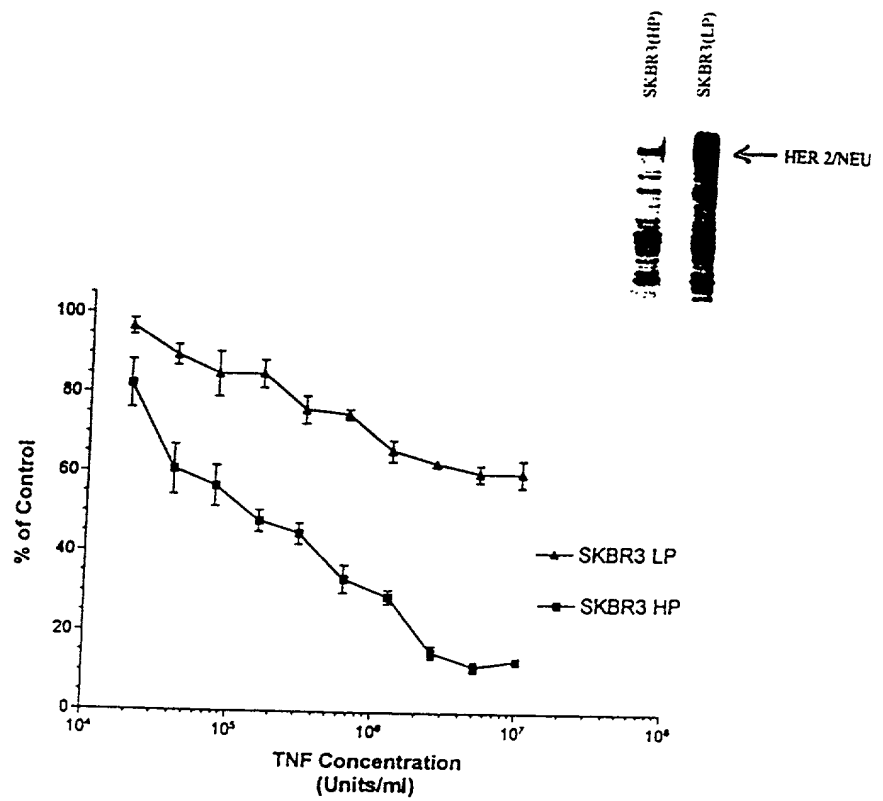


FIGURE 31

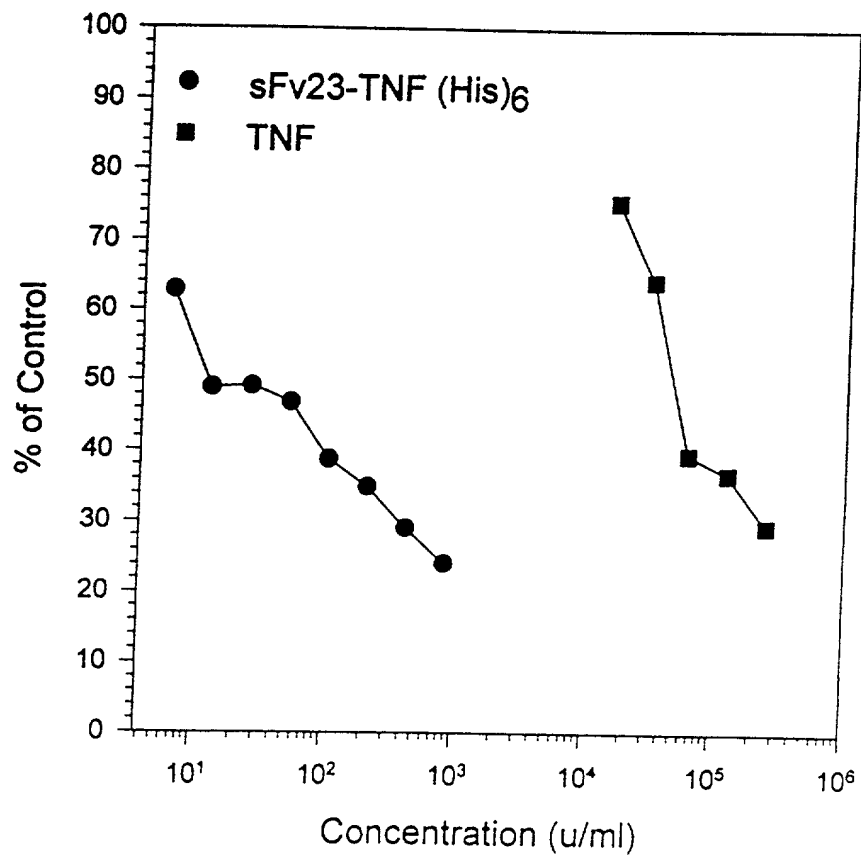
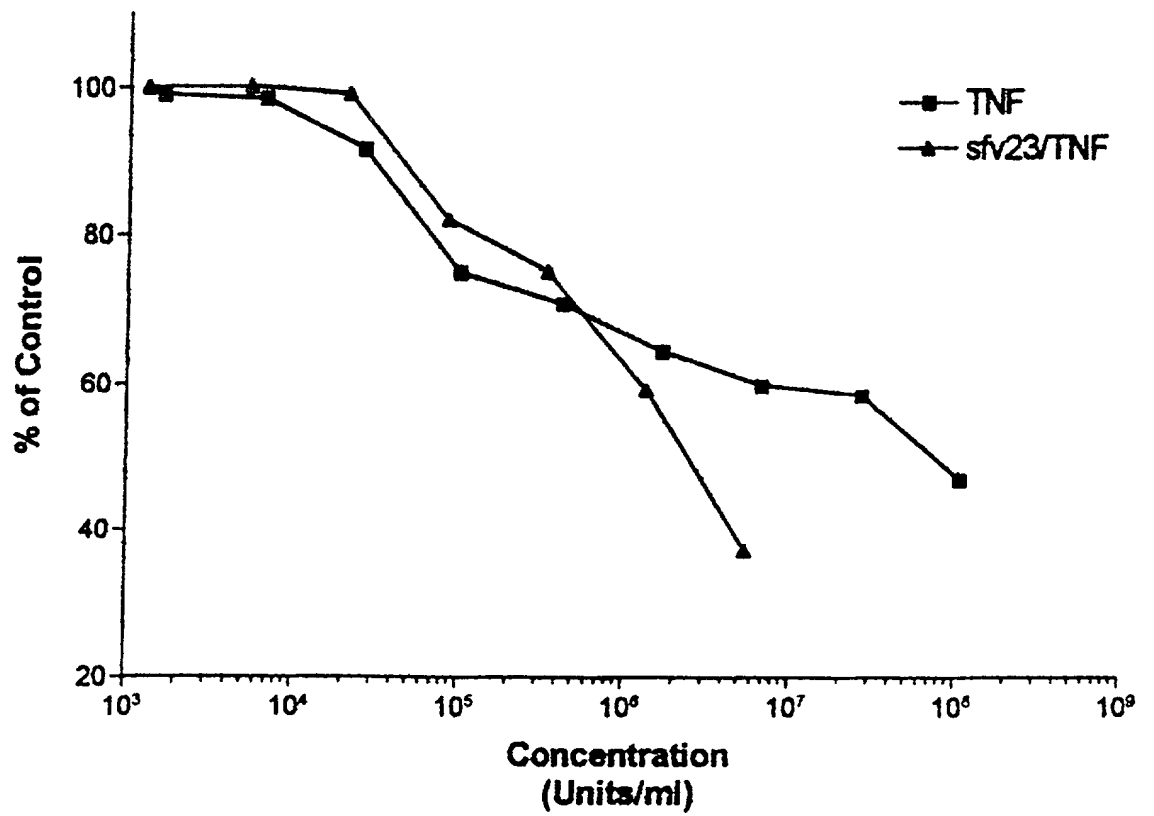


FIGURE 32



DOCKET NO: D5425CIP2

COMBINED DECLARATION AND POWER OF ATTORNEY

I, **Michael G. Rosenblum**, hereby declare that:

My residence, post office address and citizenship are as stated below next to my name. I believe I am the original, first and co-inventor, together with **Laura K. Shawver**, of the subject matter which is claimed and for which a patent is sought on the invention entitled, **Immunotoxins Directed Against c-erbB-2 (HER-2/Neu) Related Surface Antigens**; the specification of which is attached hereto and claims benefit of priority under 35 USC 120 of USSN 08/404,499 filed March 17, 1995, which is a continuation in part of US Serial No. 08/300,082, filed September 2, 1994, now abandoned, which is a continuation of US Serial No. 08/164,638, filed 12/09/93, now abandoned, which is a continuation of US Serial No. 07/867,728, filed 04/10/92, now abandoned.

I hereby state that I have reviewed and understand the contents of the above-identified specification, including the claims, as amended by any amendment referred to above. I acknowledge the duty to disclose all information I know to be material to patentability in accordance with Title 37, Code of Federal Regulations, §1.56(a), including information which became known to me between the filing date of the prior application and the national or PCT international filing date of this continuation-in-part patent application.

I hereby appoint the following attorneys and/or agents to prosecute this application and to transact all business in the Patent and Trademark Office connected therewith: Dr. Benjamin Adler, Registration No. 35,423. Address all telephone calls to Dr. Benjamin Adler at telephone number 713/777-2321. Address correspondence to Dr. Benjamin Adler, MCGREGOR & ADLER, LLP, 8011 Candle Lane, Houston, TX 77071.

I hereby declare that all statements made herein of my own knowledge are true and that all statements made on information and belief are believed to be true; and further that these statements were made with the knowledge that willful false statements and the like so made are punishable by fine or imprisonment, or both, under Section 1001 of Title 18 of the United States Code and that such willful false statements may jeopardize the validity of the application or any patents issued thereon.

Full Name of Inventor: **Michael Rosenblum**

Inventor's Signature:  Date: 5/19/99

Residence Address: 3127 Stoney Mist Dr., Sugar Land, TX 77479

Citizen of: United States of America

Post Office Address: 3127 Stoney Mist Dr., Sugar Land, TX 77479

625550 95700000

FROM : MCGREGOR&ADLER,P.C.

PHONE NO. :

May. 20 1999 12:34PM P2

DOCKET NO: D5425CIP2

COMBINED DECLARATION AND POWER OF ATTORNEY

I, **Laura K. Shawver**, hereby declare that:

My residence, post office address and citizenship are as stated below next to my name, I believe I am the original, first and co-inventor, together with **Michael G. Rosenblum**, of the subject matter which is claimed and for which a patent is sought on the invention entitled, **Immunotoxins Directed Against c-erbB-2 (HER-2/Neu) Related Surface Antigens**; the specification of which is attached hereto and claims benefit of priority under 35 USC 120 of USSN 08/404,499 filed March 17, 1995, which is a continuation in part of US Serial No. 08/300,082, filed September 2, 1994, now abandoned, which is a continuation of US Serial No. 08/164,638, filed 12/09/93, now abandoned, which is a continuation of US Serial No. 07/867,728, filed 04/10/92, now abandoned.

I hereby state that I have reviewed and understand the contents of the above-identified specification, including the claims, as amended by any amendment referred to above. I acknowledge the duty to disclose all information I know to be material to patentability in accordance with Title 37, Code of Federal Regulations, §1.56(a), including information which became known to me between the filing date of the prior application and the national or PCT international filing date of this continuation-in-part patent application.

I hereby appoint the following attorneys and/or agents to prosecute this application and to transact all business in the Patent and Trademark Office connected therewith: Dr. Benjamin Adler, Registration No. 35,423. Address all telephone calls to Dr. Benjamin Adler at telephone number 713/777-2321. Address correspondence to Dr. Benjamin Adler, MCGREGOR & ADLER, LLP, 8011 Candle Lane, Houston, TX 77071.

I hereby declare that all statements made herein of my own knowledge are true and that all statements made on information and belief are believed to be true; and further that these statements were made with the knowledge that willful false statements and the like so made are punishable by fine or imprisonment, or both, under Section 1001 of Title 18 of the United States Code and that such willful false statements may jeopardize the validity of the application or any patents issued thereon.

Full Name of Inventor: **Laura K. Shawver**

Inventor's Signature: *Laura K. Shawver*

Date: May 24, 1995

Residence Address: 216 Cotter Street, San Francisco CA 94112

Citizen of: United States of America

Post Office Address: 216 Cotter Street, San Francisco, CA 94112

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